

APPLICATION OF GAS CHROMATOGRAPHY TIME-OF-FLIGHT HRMS MASS SPECTROMETRY TO FOOD ANALYSIS

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During the last years the introduction of liquid chromatography high resolution mass spectrometry (LC-HRMS) has become popular in pesticide residues laboratories. The advantages in getting exact mass of the analytes have been evaluated giving an important and new solution to common problems of these analyses. But, till now practically all applications have been focused on LC. Recently new GC-HRMS are intending to cover a similar position for typical GC pesticide residues. Some questions remain to clarify if the established workflows in LC can be applicable for GC-HRMS considering of the differences between ionization systems as ESI or EI.

This work reports the development and evaluation of a rapid automated screening method for determining pesticide residues in food using gas chromatography time-of-flight mass spectrometry (GC-TOF-MS) based on the use of an accurate-mass database. The database (including 87 GC amenable pesticides) created include, ions obtained under electron impact ionization at 70 eV and retention times of each pesticide under retention time locking of pesticides at constant flow.

This customized database was associated to commercially available software which extracted all the potential compounds of interest from the GC-TOF MS raw data of each sample and matched them against the database to search for targeted compounds in the sample.

The automatic identification with the developed workflow has been tested in tomato, orange, potato and onion extract spiked at 10, 20, 50 and 100 µg/ml level, most pesticides present in the sample were identified correctly, the identification criteria were a retention time window of 0.5 min, and a mass error tolerance of 10 ppm for at least two fragments for each pesticide.

GC MASS SPECTROMETRIC CONDITIONS

Injection volume: 1µL
Mode: Splitless MM Inlet
Injector temperature: 280°C
2 columns: HP5MSI (350°C) 15m x 250µm x 0.25µm
Full Scan EI Mode
Constant Flow
Column 1 = 1.225ml/min
Column 2 = 1.425ml/min



Agilent 7200 Q-TOF GC/MS

OVEN PROGRAM

Rate(°C/min)	Value(°C)	Hold Time(min)	Run Time(min)
	60	1	1
	40	120	0
	5	310	0
			40.5

BACKFLUSH CONFIGURATION



Post Run: 310°C

Post Run Time: 2 min

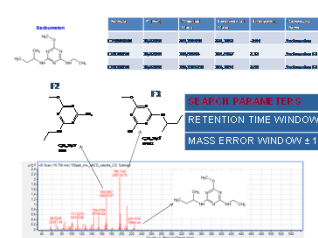
Ion source temperature: 280°C

Transfer line temperature: 280°C

ACURRATE MASS DATABASE

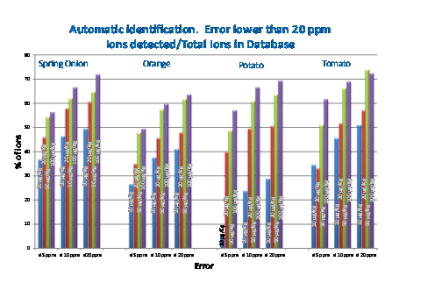
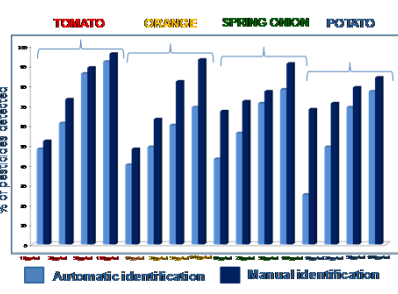
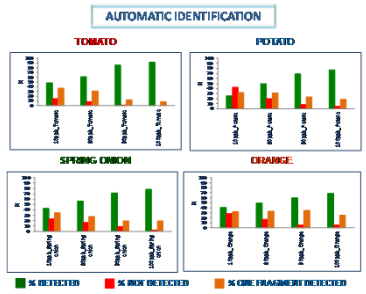
87 PESTICIDES
2 FRAGMENTS AT LEAST FOR EACH PESTICIDE
RETENTION TIMES OF EACH PESTICIDE

#Pesticide	Retention Time	Mass	Chemical name	Comments
1	8.17	174.0386	Imidacloprid	
2	11.18	185.0457	Imidacloprid	
3	11.18	185.0457	Imidacloprid	
4	11.18	185.0457	Imidacloprid	
5	11.18	185.0457	Imidacloprid	
6	11.18	185.0457	Imidacloprid	
7	11.18	185.0457	Imidacloprid	
8	11.18	185.0457	Imidacloprid	
9	11.18	185.0457	Imidacloprid	
10	11.18	185.0457	Imidacloprid	
11	11.18	185.0457	Imidacloprid	
12	11.18	185.0457	Imidacloprid	
13	11.18	185.0457	Imidacloprid	
14	11.18	185.0457	Imidacloprid	
15	11.18	185.0457	Imidacloprid	
16	11.18	185.0457	Imidacloprid	
17	11.18	185.0457	Imidacloprid	
18	11.18	185.0457	Imidacloprid	
19	11.18	185.0457	Imidacloprid	
20	11.18	185.0457	Imidacloprid	
21	11.18	185.0457	Imidacloprid	
22	11.18	185.0457	Imidacloprid	
23	11.18	185.0457	Imidacloprid	
24	11.18	185.0457	Imidacloprid	
25	11.18	185.0457	Imidacloprid	
26	11.18	185.0457	Imidacloprid	
27	11.18	185.0457	Imidacloprid	
28	11.18	185.0457	Imidacloprid	
29	11.18	185.0457	Imidacloprid	
30	11.18	185.0457	Imidacloprid	
31	11.18	185.0457	Imidacloprid	
32	11.18	185.0457	Imidacloprid	
33	11.18	185.0457	Imidacloprid	
34	11.18	185.0457	Imidacloprid	
35	11.18	185.0457	Imidacloprid	
36	11.18	185.0457	Imidacloprid	
37	11.18	185.0457	Imidacloprid	
38	11.18	185.0457	Imidacloprid	
39	11.18	185.0457	Imidacloprid	
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41	11.18	185.0457	Imidacloprid	
42	11.18	185.0457	Imidacloprid	
43	11.18	185.0457	Imidacloprid	
44	11.18	185.0457	Imidacloprid	
45	11.18	185.0457	Imidacloprid	
46	11.18	185.0457	Imidacloprid	
47	11.18	185.0457	Imidacloprid	
48	11.18	185.0457	Imidacloprid	
49	11.18	185.0457	Imidacloprid	
50	11.18	185.0457	Imidacloprid	
51	11.18	185.0457	Imidacloprid	
52	11.18	185.0457	Imidacloprid	
53	11.18	185.0457	Imidacloprid	
54	11.18	185.0457	Imidacloprid	
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63	11.18	185.0457	Imidacloprid	
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73	11.18	185.0457	Imidacloprid	
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75	11.18	185.0457	Imidacloprid	
76	11.18	185.0457	Imidacloprid	
77	11.18	185.0457	Imidacloprid	
78	11.18	185.0457	Imidacloprid	
79	11.18	185.0457	Imidacloprid	
80	11.18	185.0457	Imidacloprid	
81	11.18	185.0457	Imidacloprid	
82	11.18	185.0457	Imidacloprid	
83	11.18	185.0457	Imidacloprid	
84	11.18	185.0457	Imidacloprid	
85	11.18	185.0457	Imidacloprid	
86	11.18	185.0457	Imidacloprid	
87	11.18	185.0457	Imidacloprid	

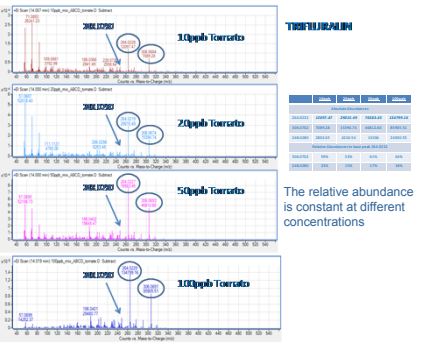
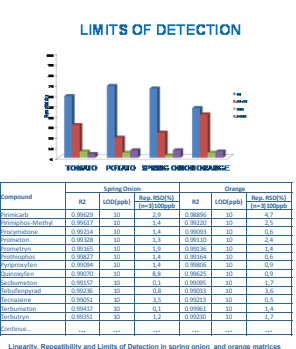


SEARCH PARAMETERS:
RETENTION TIME WINDOW ± 0.5 min
MASS ERROR WINDOW ± 10 ppm

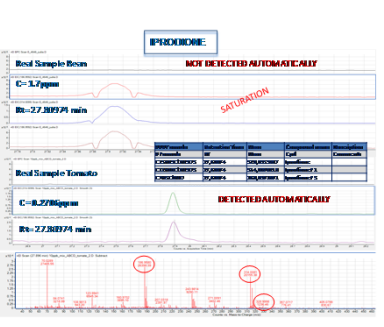
AUTOMATIC IDENTIFICATION VS MANUAL IDENTIFICATION



VALIDATION STUDY



ANALYSIS OF REAL SAMPLES



REAL SAMPLES

Matrix	GC-TOF automatic	GC-QQQ	GC-TOF manual
Pepper	Imidacloprid (20µg/kg)	Imidacloprid (20µg/kg)	Imidacloprid (20µg/kg)
Tomato_1	Imidacloprid (20µg/kg)	Imidacloprid (20µg/kg)	Imidacloprid (20µg/kg)
Tomato_2	Imidacloprid (20µg/kg)	Imidacloprid (20µg/kg)	Imidacloprid (20µg/kg)
Tomato_3	Imidacloprid (20µg/kg)	Imidacloprid (20µg/kg)	Imidacloprid (20µg/kg)
Tomato_4	Imidacloprid (20µg/kg)	Imidacloprid (20µg/kg)	Imidacloprid (20µg/kg)
Tomato_5	Imidacloprid (20µg/kg)	Imidacloprid (20µg/kg)	Imidacloprid (20µg/kg)
Tomato_6	Imidacloprid (20µg/kg)	Imidacloprid (20µg/kg)	Imidacloprid (20µg/kg)
Tomato_7	Imidacloprid (20µg/kg)	Imidacloprid (20µg/kg)	Imidacloprid (20µg/kg)
Tomato_8	Imidacloprid (20µg/kg)	Imidacloprid (20µg/kg)	Imidacloprid (20µg/kg)
Tomato_9	Imidacloprid (20µg/kg)	Imidacloprid (20µg/kg)	Imidacloprid (20µg/kg)
Tomato_10	Imidacloprid (20µg/kg)	Imidacloprid (20µg/kg)	Imidacloprid (20µg/kg)
Tomato_11	Imidacloprid (20µg/kg)	Imidacloprid (20µg/kg)	Imidacloprid (20µg/kg)
Tomato_12	Imidacloprid (20µg/kg)	Imidacloprid (20µg/kg)	Imidacloprid (20µg/kg)
Tomato_13	Imidacloprid (20µg/kg)	Imidacloprid (20µg/kg)	Imidacloprid (20µg/kg)
Tomato_14	Imidacloprid (20µg/kg)	Imidacloprid (20µg/kg)	Imidacloprid (20µg/kg)
Tomato_15	Imidacloprid (20µg/kg)	Imidacloprid (20µg/kg)	Imidacloprid (20µg/kg)
Tomato_16	Imidacloprid (20µg/kg)	Imidacloprid (20µg/kg)	Imidacloprid (20µg/kg)
Tomato_17	Imidacloprid (20µg/kg)	Imidacloprid (20µg/kg)	Imidacloprid (20µg/kg)
Tomato_18	Imidacloprid (20µg/kg)	Imidacloprid (20µg/kg)	Imidacloprid (20µg/kg)
Tomato_19	Imidacloprid (20µg/kg)	Imidacloprid (20µg/kg)	Imidacloprid (20µg/kg)
Tomato_20	Imidacloprid (20µg/kg)	Imidacloprid (20µg/kg)	Imidacloprid (20µg/kg)
Tomato_21	Imidacloprid (20µg/kg)	Imidacloprid (20µg/kg)	Imidacloprid (20µg/kg)
Tomato_22	Imidacloprid (20µg/kg)	Imidacloprid (20µg/kg)	Imidacloprid (20µg/kg)
Orange	Imidacloprid (20µg/kg)	Imidacloprid (20µg/kg)	Imidacloprid (20µg/kg)

CONCLUSIONS

The development of an accurate mass database in GC-TOF-MS has been performed, in a first step the pesticides were analyzed under EI at 70eV and with retention time locked. Their mass spectrum were investigated, each experimental ion has been assigned to a part of the molecule in order to calculate the theoretical exact mass of each fragment. In most of cases the molecular ion was not present in the mass spectrum.
The accurate mass data base created includes at least two fragments for each pesticide and its retention time.
The accurate mass data base was tested for automatic identification in four matrices: tomato, spring onion, orange and potato in four different concentrations. In all cases the error was lower than ±20ppm and in around a 65% of the detected ions the error was lower than ± 5ppm.
The automatic identification was compared with manual identification and differences arise especially at low concentrations.
15 real samples were processed for the automatic identification with the developed data base, results were compared with those obtained by GC-QQQ-MS, from a total of 22 pesticides presents in the real samples 14 were correctly identified automatically, and three pesticides were not identified correctly by GCQTOF because the high concentration detected saturate the detector.
This work is a first approach to the accurate mass database for automatic identification in GC-TOF-MS. It can be considered as a starting point for a large database and for an improved methodology for automatic identification.