

# *IN-TUBE EXTRACTION (ITEX) AND DETERMINATION OF 2-CHLOROETHANOL*



MARIA DEL MAR GÓMEZ RAMOS  
LORENA MANZANO SÁNCHEZ

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## Introduction

The sample is heated and / or agitated in a sealed vial.

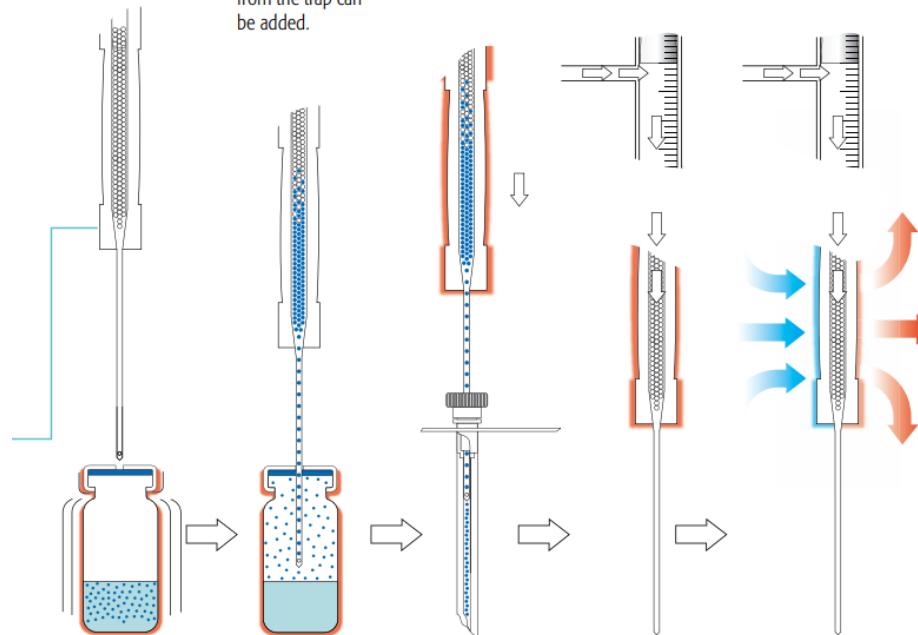
The ITEX needle pierces the sealed vial and the heated syringe pumps the headspace gas through the cold trap.

An additional step to remove water from the trap can be added.

The loaded ITEX trap is flash heated up to 350°C and analytes are desorbed into the hot GC injector.

After thermal desorption the hot ITEX trap is cleaned with inert flush gas.

Active cooling allows for short cycle times.



ITEX Tool



## Introduction

### Sorbent Materials

Sorbent	Sorbent type	Specific surface area ( $\text{m}^2 \text{ g}^{-1}$ )	Temperature limit (°C)	Water affinity	Typical applications
Carbopack C	Graphitized carbon black	10	500	Relatively low	Low to medium boilers (C12–C20)
Carboxen 1000	Carbon molecular sieve	1200	225	Moderate	Permanent gases, volatiles (C2–C5)
Carbosieve SIII	Carbon molecular sieve	975	400	Moderate	Volatile organics (C2–C5)
Tenax GR	70 % porous organic polymer/30 % graphitized carbon	24	350	Low	Volatiles, flavors
Tenax TA	Porous organic polymer	35	350	Low	Volatiles and semi-volatiles (C7–C26)
HayeSep D	Porous organic polymer	795	290	Low	Volatiles (C1–C6)
MWCNT	Multi-walled carbon nanotubes	211 <sup>a</sup>	n.a.	n.a.	n.a.
PDMS	Silicone rubber	Absorbent	250	Low	Nonpolar volatiles, semi-volatiles
Carbowax 20M	Polyethylene glycol	Absorbent	225	High	Polar semi-volatiles

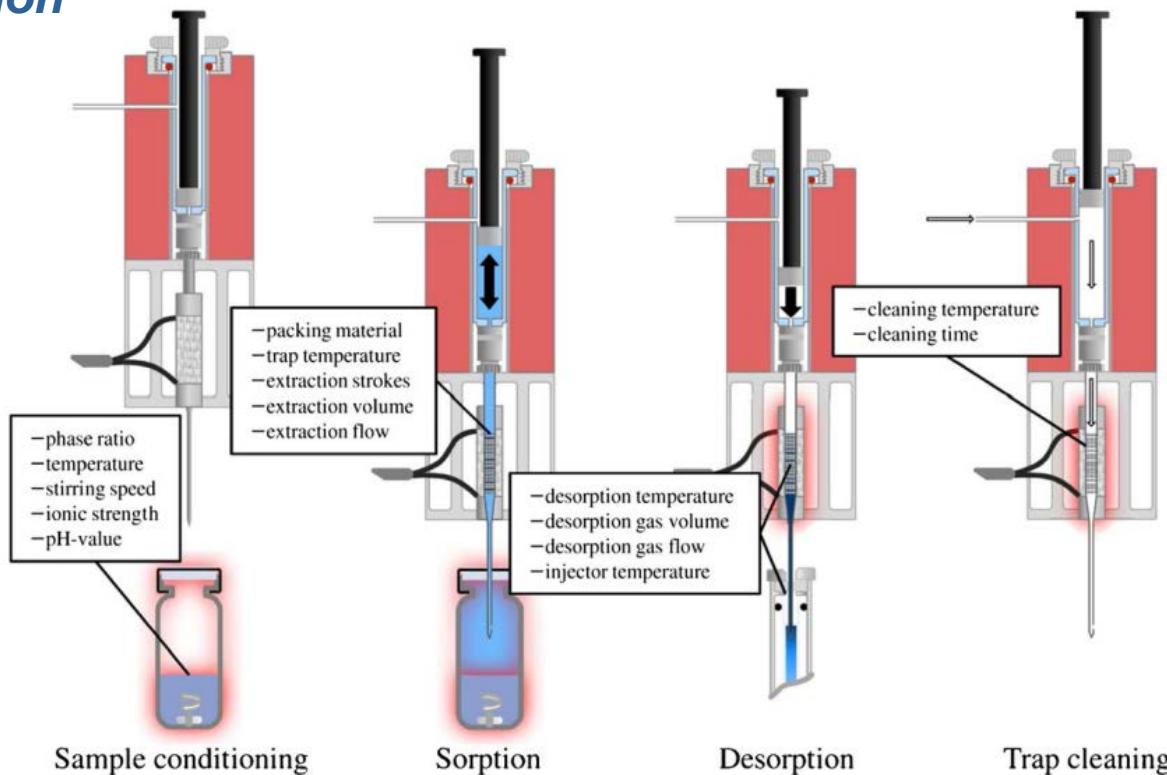
*n.a.* not available



Source: Laaks, J.; Jochmann, M. A.; Schilling, B.; Schmidt, T. C. Optimization Strategies of In-Tube Extraction (ITEX) Methods. *Anal. Bioanal. Chem.* **2015**, 407 (22), 6827–6838.  
<https://doi.org/10.1007/s00216-015-8854-4>

## Introduction

### Parameters



## Introduction

### Parameters

- Sorbent and sample temperature**

Sorbent material is placed in a tube outside the heated sample vial and that the trap temperature can be controlled independently from the conditioning temperature of the sample. However, when the temperature difference between the sample vial and the trap becomes too large, problems with condensation of water on the sorbent material can arise.

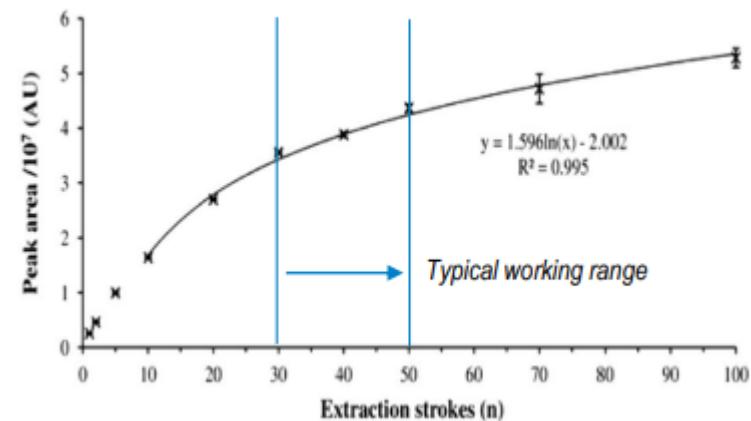
- Extraction Flow and extraction strokes**

Number of extraction strokes:

- Flexible adjustment of analyte enrichment
- Defines sampled volume
- No loss of analytes, “closed” system

Extraction Flow:

- Analyte specific
- Yield decreased towards higher extraction flows
- Stronger effect at lower analyte concentrations



Source: Laaks, J.; Jochmann, M. A.; Schilling, B.; Schmidt, T. C. Optimization Strategies of In-Tube Extraction (ITEX) Methods. *Anal. Bioanal. Chem.* **2015**, 407 (22), 6827–6838.  
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## Introduction

### Volatile compounds

Description	Abbreviation	Boiling Point Range (°C)	Example Compounds
Very volatile (gaseous) organic compounds	VVOC	<0 to 50-100	Propane, butane, methyl chloride
Volatile organic compounds	VOC	50-100 to 240-260	Formaldehyde, d-Limonene, toluene, acetone, ethanol (ethyl alcohol) 2-propanol (isopropyl alcohol), hexanal
Semi volatile organic compounds	SVOC	240-260 to 380-400	Pesticides (DDT, chlordane, plasticizers (phthalates), fire retardants (PCBs, PBB))

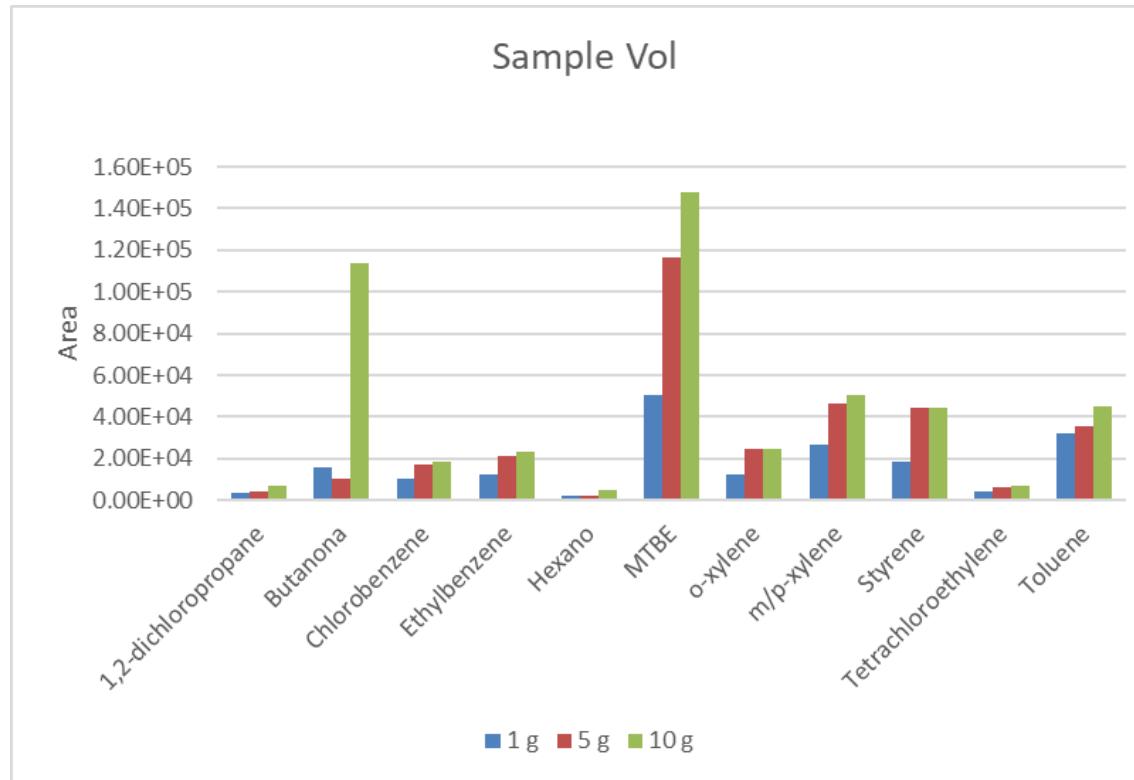
Source: EPA <https://www.epa.gov/indoor-air-quality-iaq/technical-overview-volatile-organic-compounds>

Compound	Vap Pressure (at 25 °C) mmHg	Boiling Point (°C)
1,2-Dichloropropane	40	96
Butanone	78	65
Chlorobenzene	11	132
Ethylbenzene	7	136
Hexane	120	69
MTBE	245	55
Xylenes	18	139
Styrene	12.4	145
Tetrachloroethylene	18.47	121
Toluene	28.7	111

## Optimization of parameters (Oil sample)

### Parameters

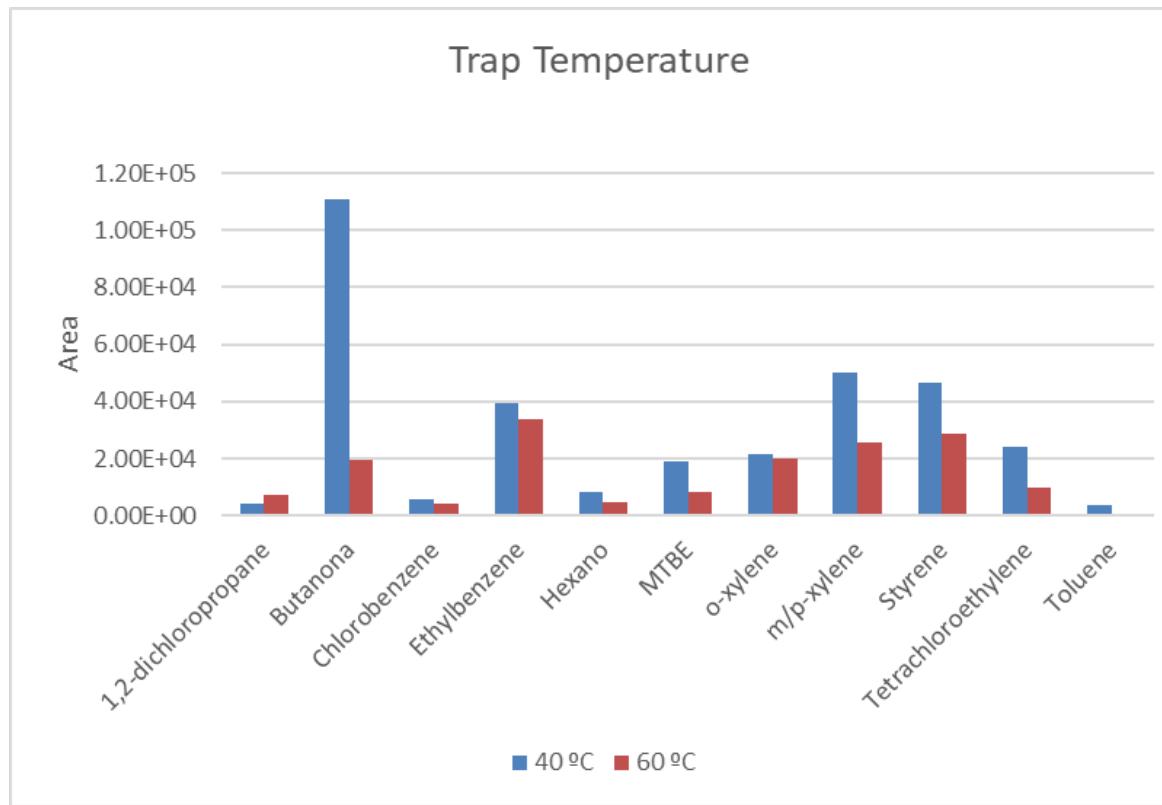
- Sample Volume



## Optimization of parameters (Oil sample)

### Parameters

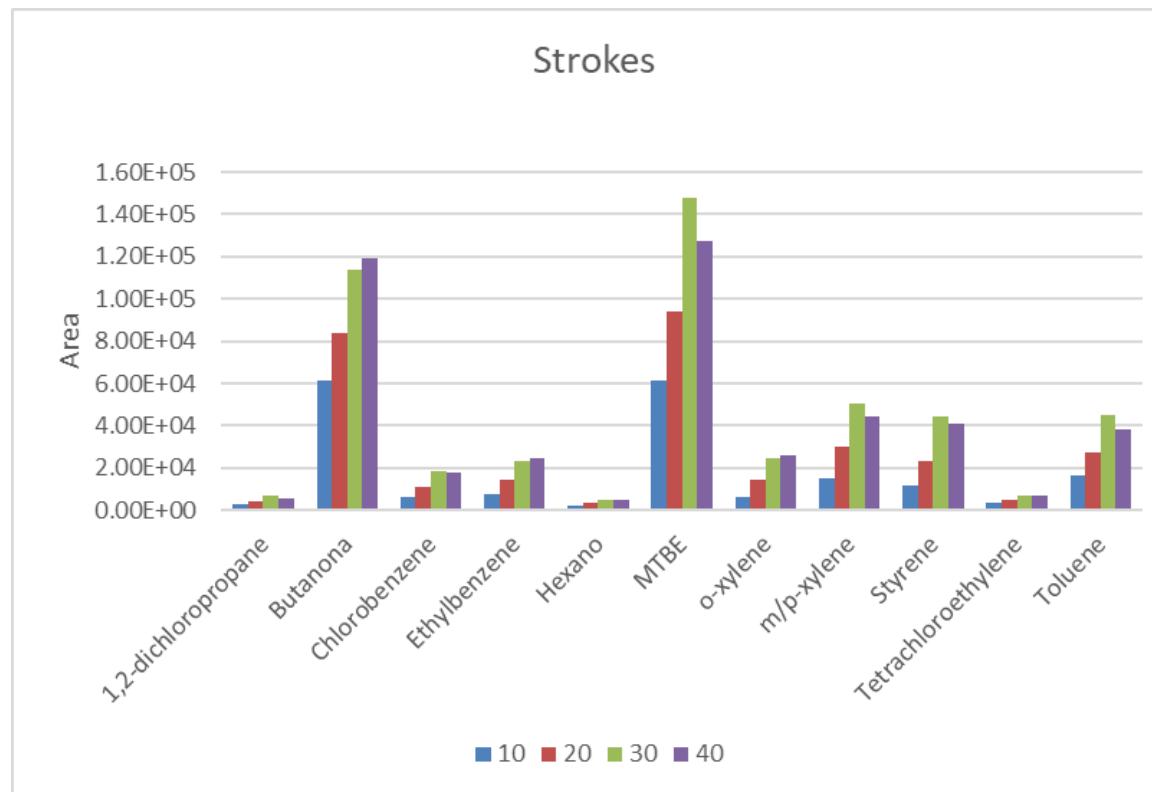
- Trap Temperature

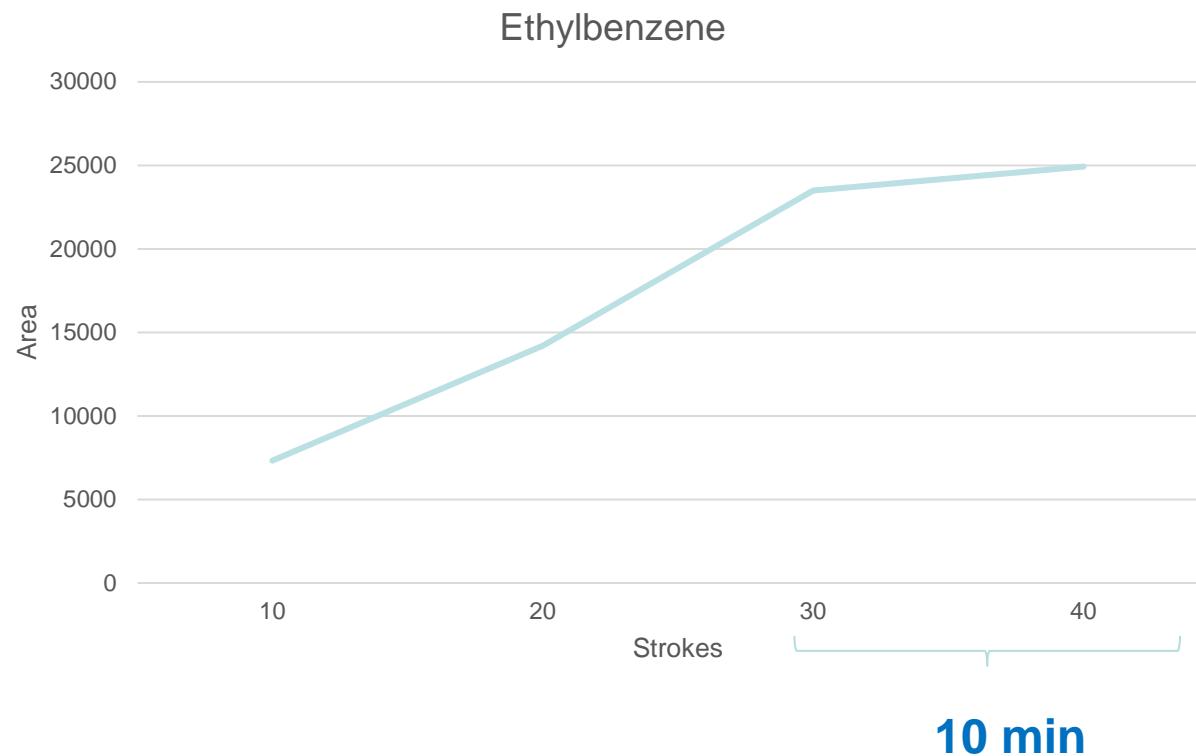


## Optimization of parameters (Oil sample)

### Parameters

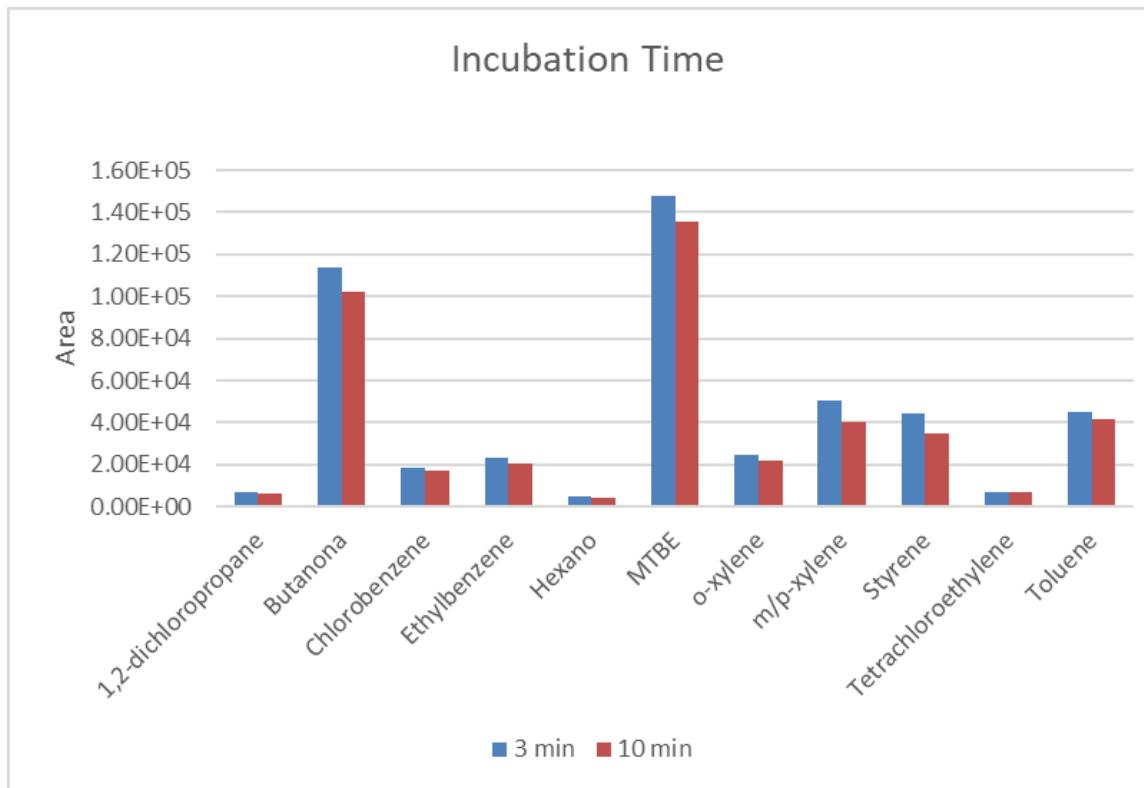
- Strokes





## Optimization of parameters (Oil sample)

- Incubation Time

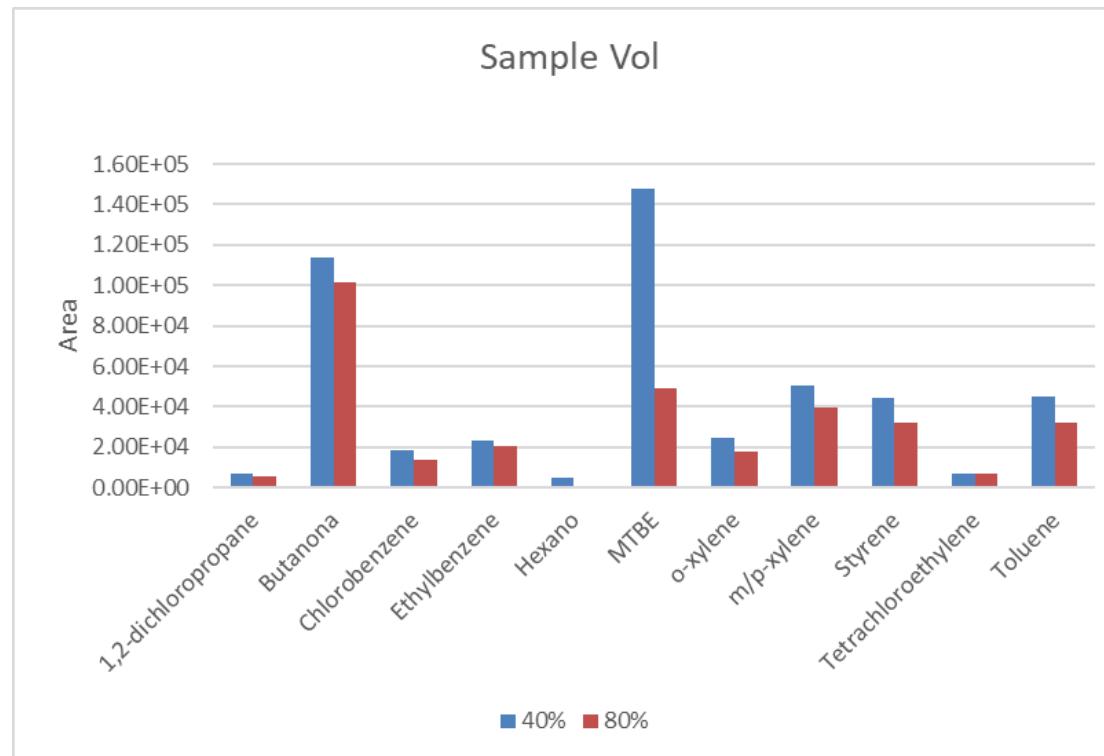




## Optimization of parameters (Oil sample)

### Parameters

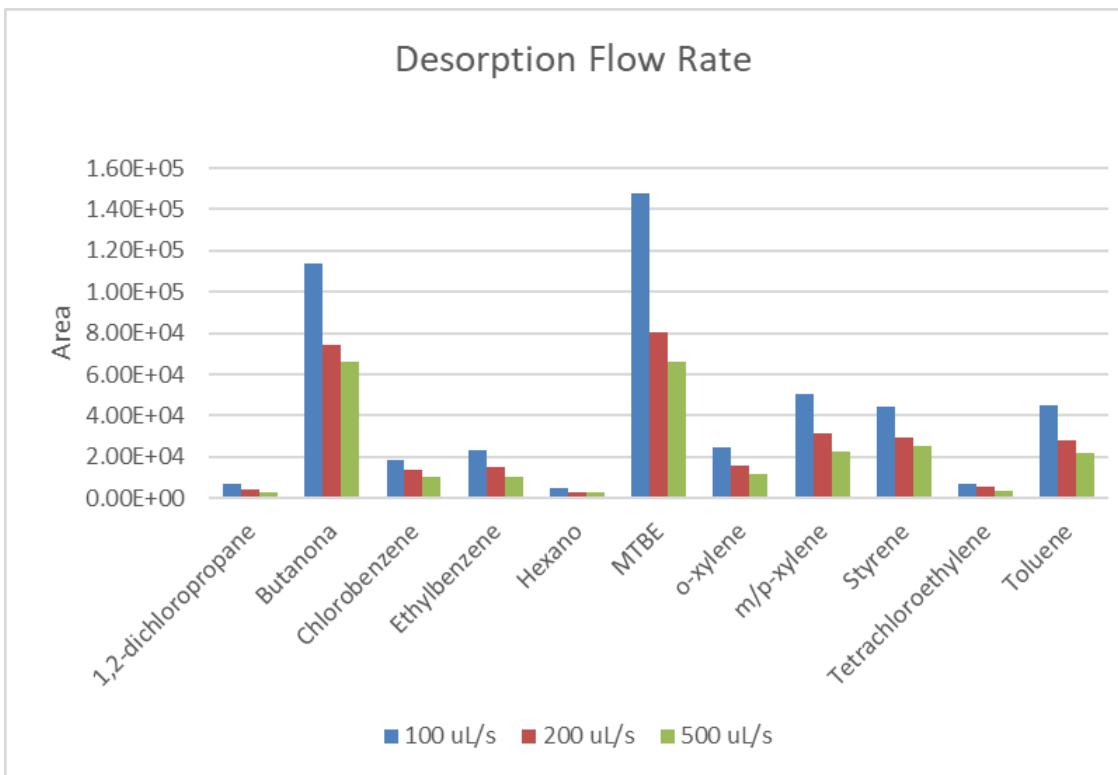
- Sample Prefill



## Optimization of parameters (Oil sample)

### Parameters

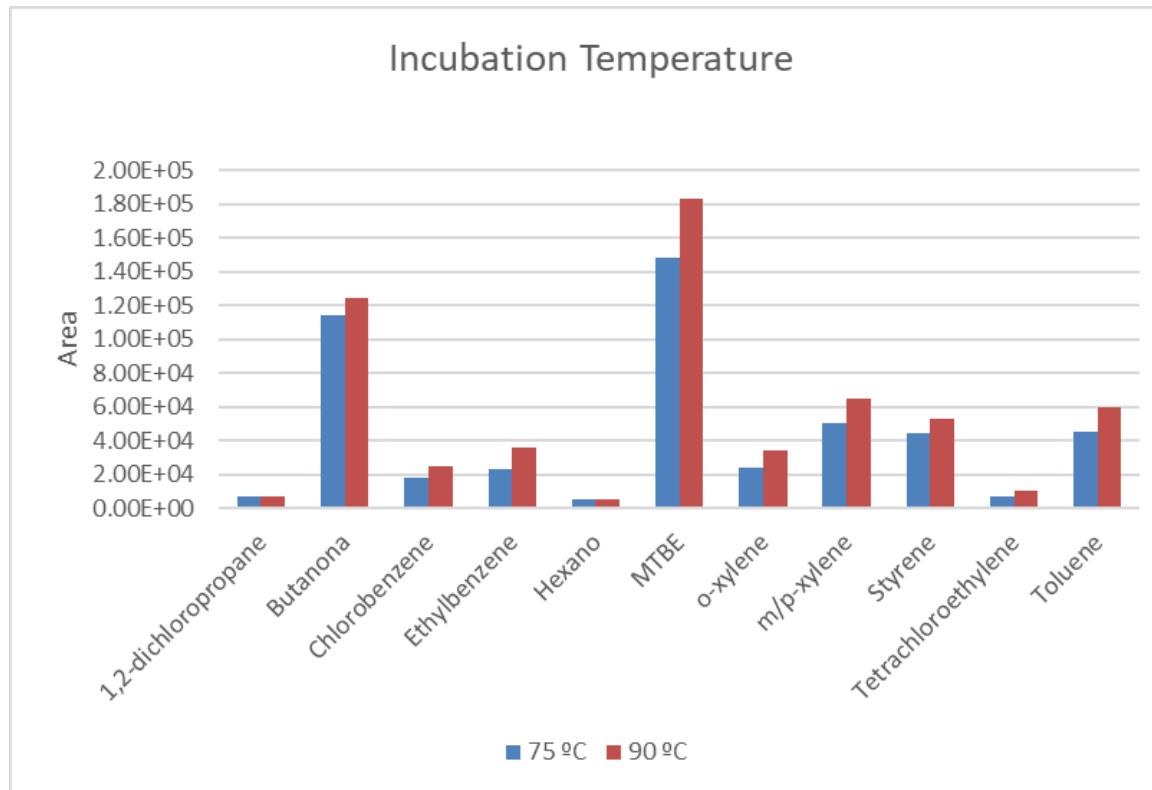
- Desorption Flow Rate



## Optimization of parameters (Oil sample)

### Parameters

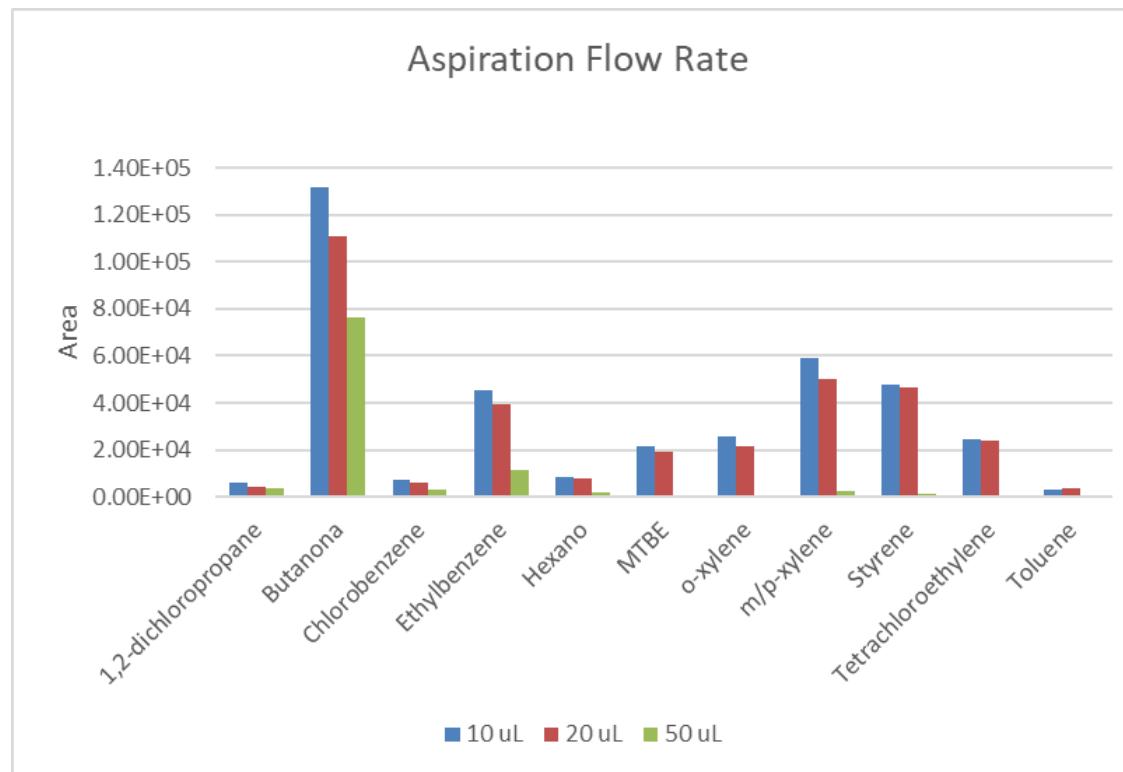
- Incubation Temperature



## Optimization of parameters (Oil sample)

### Parameters

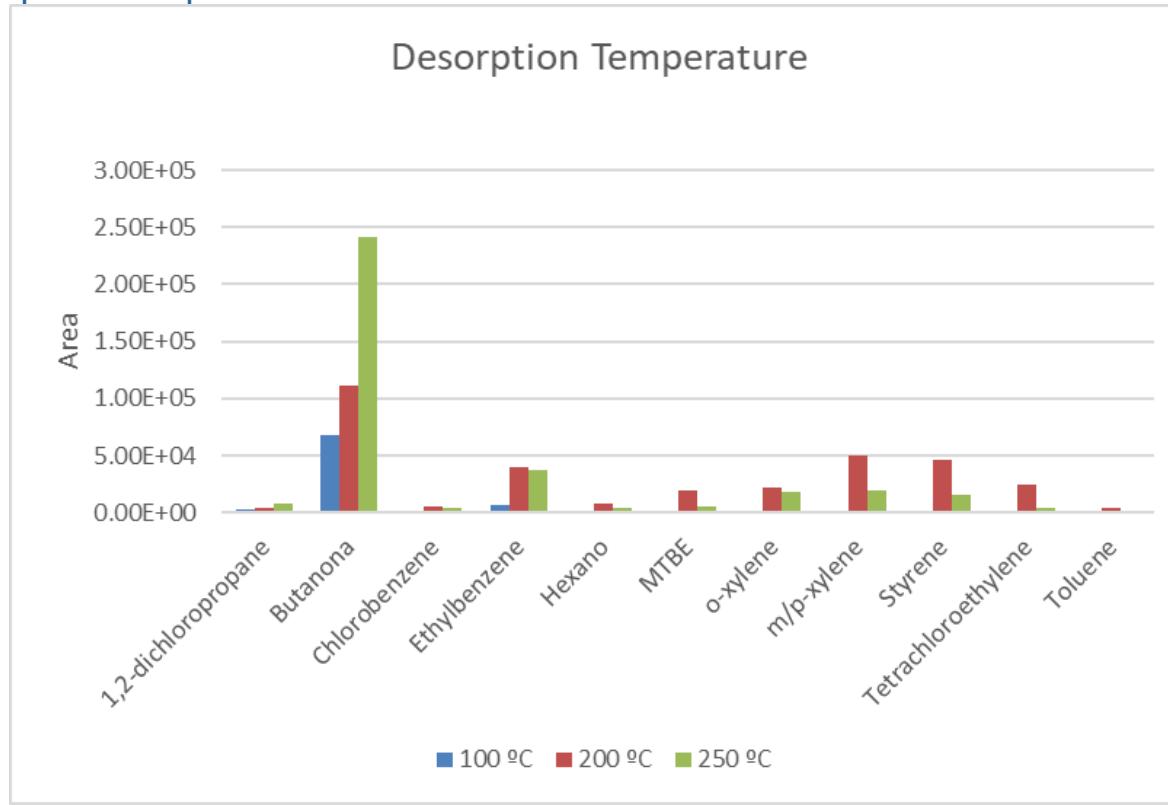
- Extraction Aspirate Flow Rate



## Optimization of parameters (Oil sample)

### Parameters

- Desorption Temperature



## Optimization of parameters (Oil sample)

Tool	ITEX 1
GC Cycle Time	35 min
Trap Preclean Time	500 s
Trap Preclean Temperature	250 °C
Agitator	Agitator 1
Heat Agitator	True
Incubation Temperature	90 °C
Incubation Time	3 min
Syringe Temperature	90 °C
Trap Purge Time	50 s

### Sample

Trap Extraction Temperature	40 °C
Sample Volume	500 uL
Sample Vial Penetration Depth	12 mm
Extraction Aspirate Flow Rate	20 uL/s
Extraction Dispense Flow Rate	200 uL/s
Extraction Strokes	30
Extraction Volume	1000 uL
Extraction Pullup Delay	5 s
Sample Prefill Ratio	40 %

### Inject Sample

Desorption Flow Rate	100 uL/s
Desorption Temperature	200 °C
Inlet Penetration Depth	35 mm
Inlet Penetration Speed	50 mm/s
Injection Signal Mode	Plunger Up

### Post Purge Syringe

Trap Post Cleaning Time	500 s
Trap Post Cleaning Temperature	250 °C

### Remove Water

### Advanced

Agitator Speed	500 rpm
Set Syringe Standby Temperature	Automatic
Agitator On Time	5 s
Syringe Standby Temperature	90 °C
Agitator Off Time	1 s
Set Trap Standby Temperature	Automatic
Trap Standby Temperature	40 °C
Agitator Standby Temperature	90 °C
Set Agitator Standby Temperature	Automatic

## Analysis Time

ITEX: 30 min aprox

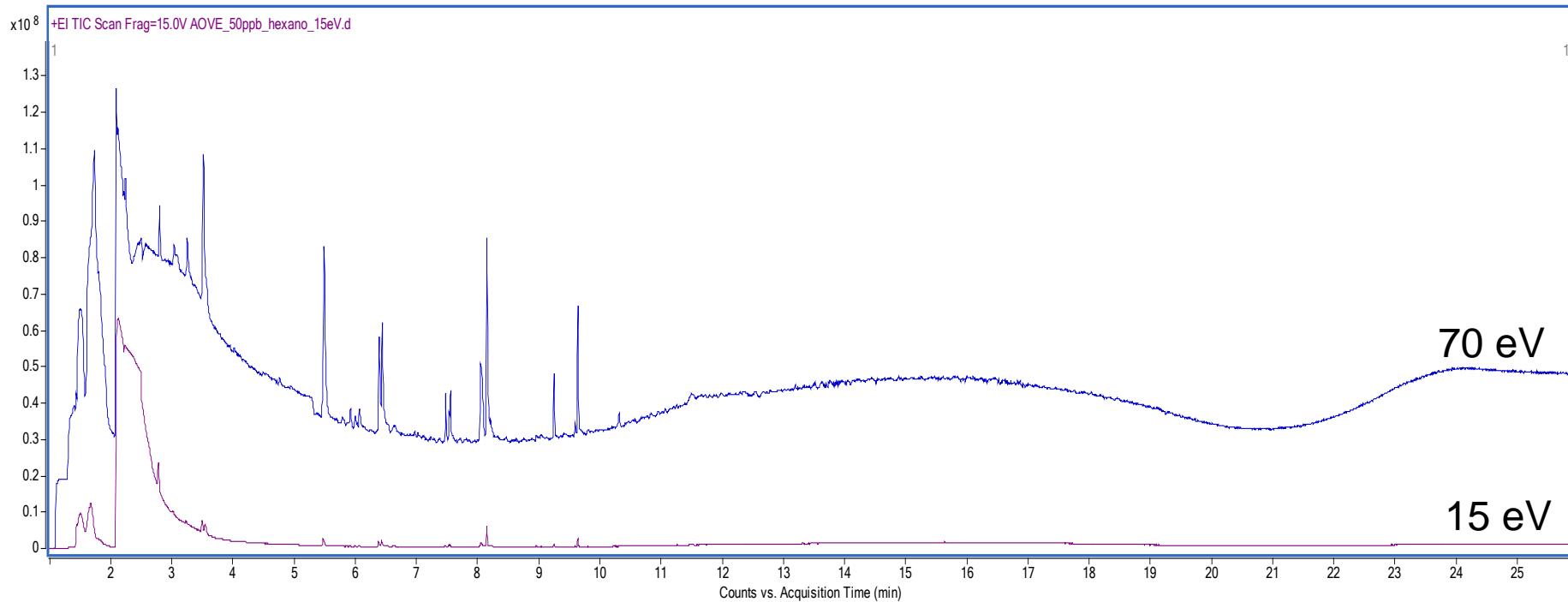
Chromatography: 14 min

## Source of GC-QTOF

EI MODE

*Evaluation of different electron energy: 70 eV and 15 eV*

TIC of Olive Oil Sample

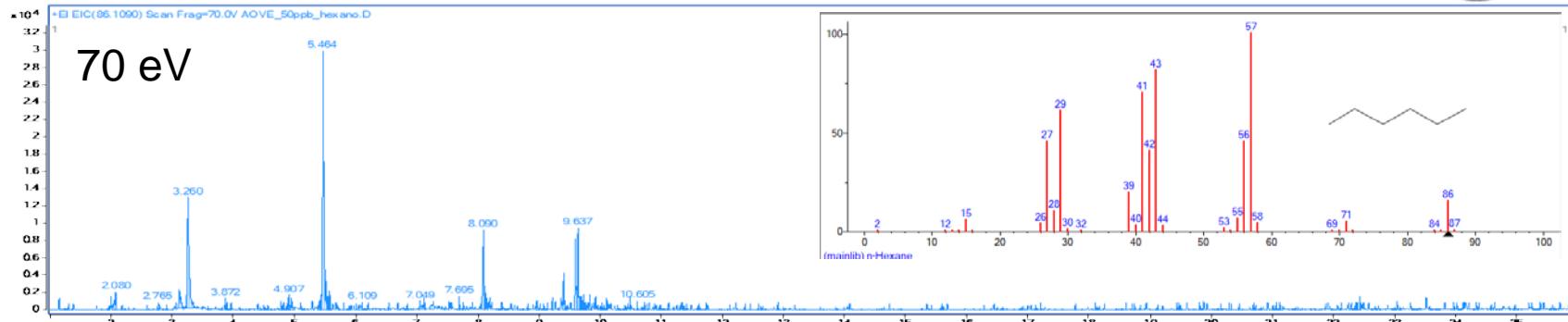
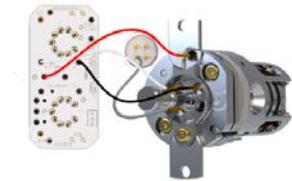


## Source of GC-QTOF

### EI MODE

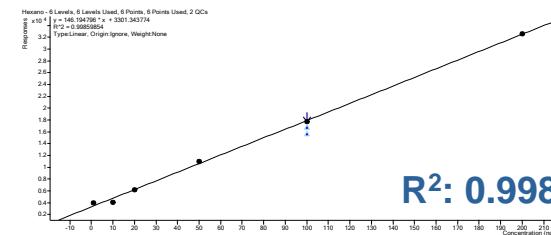
*Evaluation of different electron energy: 70 eV and 15 eV*

**Hexane Radical Ion ( $m/z$ ): 86.109**

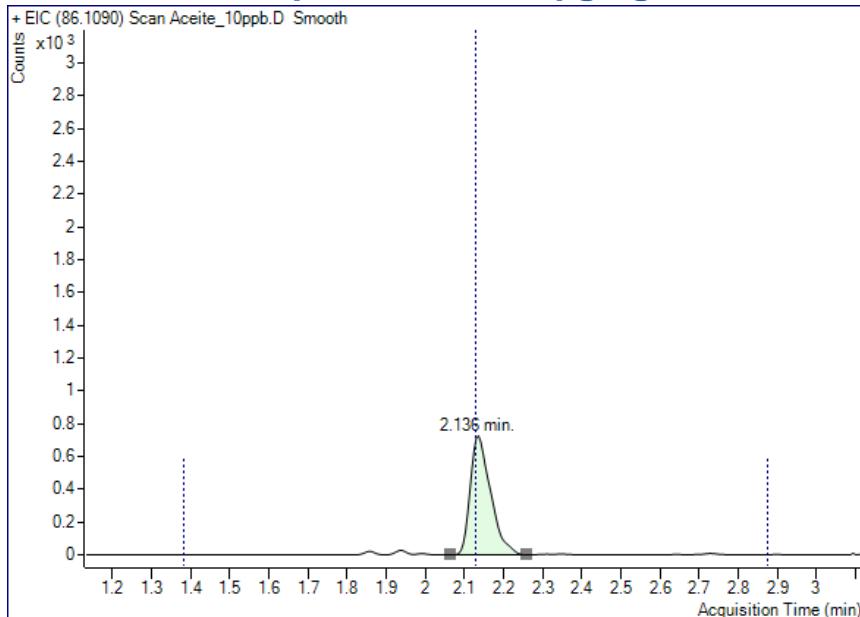


Counts vs. Acquisition Time (min)

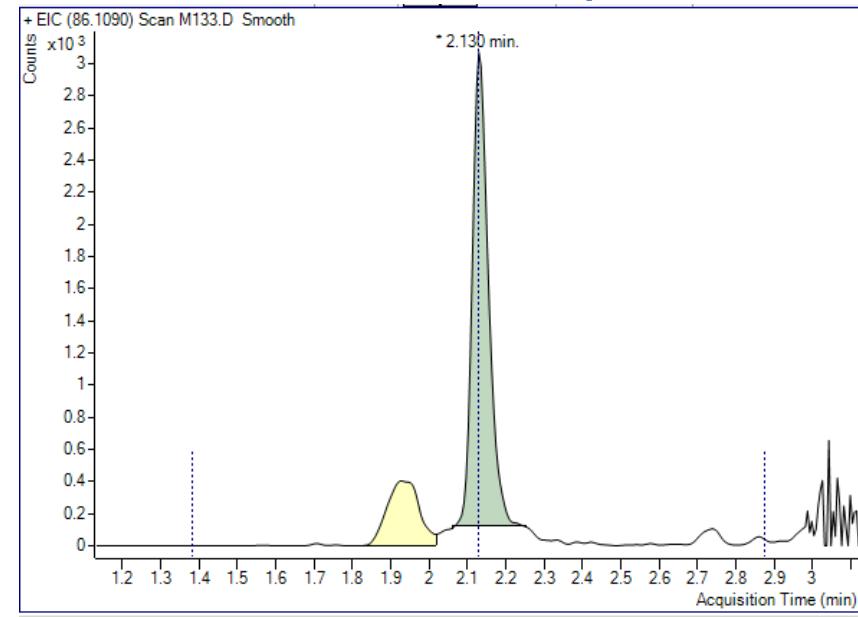
## Determination of hexane in real simples (MRL: 1 mg/kg)



**Spiked Oil at 10 µg/kg**



**Olive Oil Sample**



**Hexane: 48**  
**µg/kg**

## Determination of 2-chloroethanol in sesame seeds

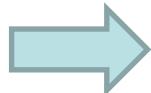


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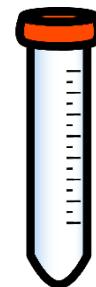
**10 mL Ethyl Acetate  
0.3 g NaCl  
1.6 g MgSO<sub>4</sub>**

**2 g**

**Centrifuge (5 min 3500 rpm)**



**4 mL supernatant  
100 mg C18 + 100 mg PSA**



Amplitude: 100 mm  
Speed: 2.5 m/s  
Acceleration: 80 m/s<sup>2</sup>  
Jerk: 6  
Time: 900 se

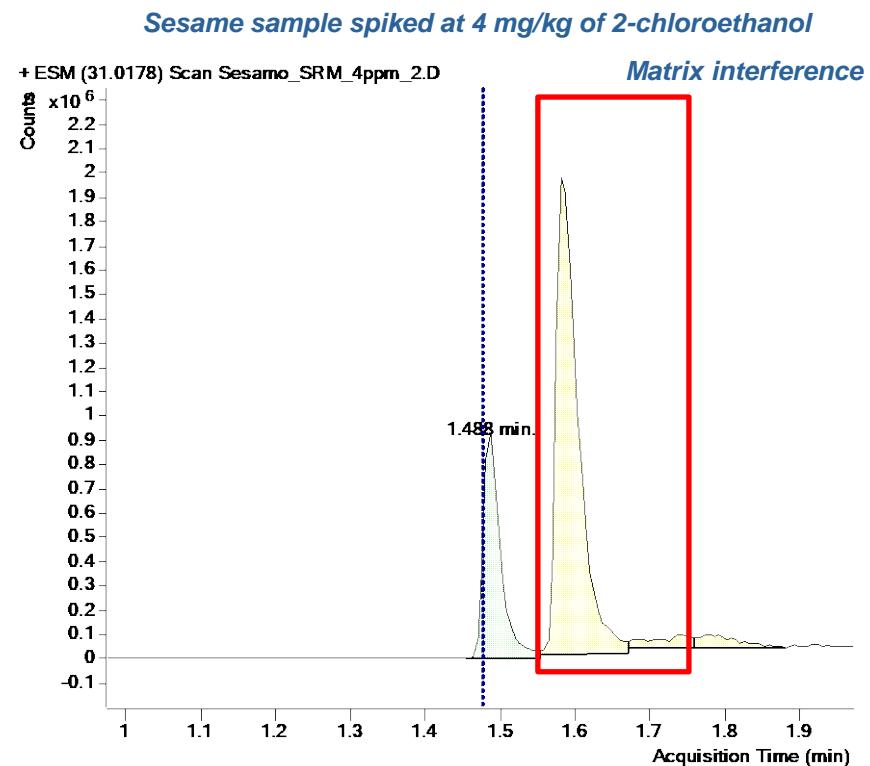
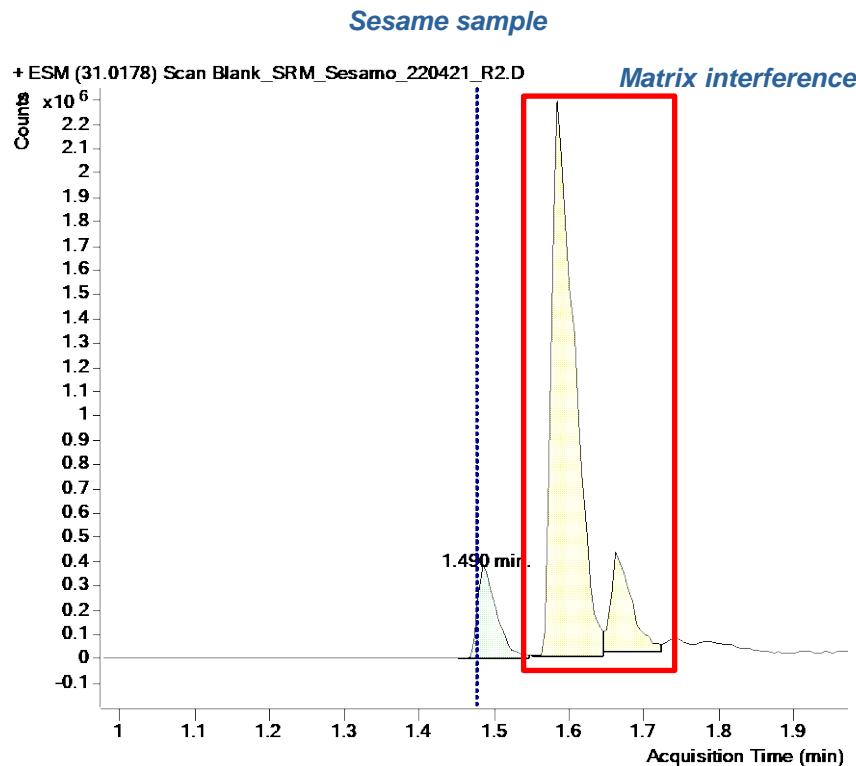
**Centrifuge (5 min 3500 rpm)**



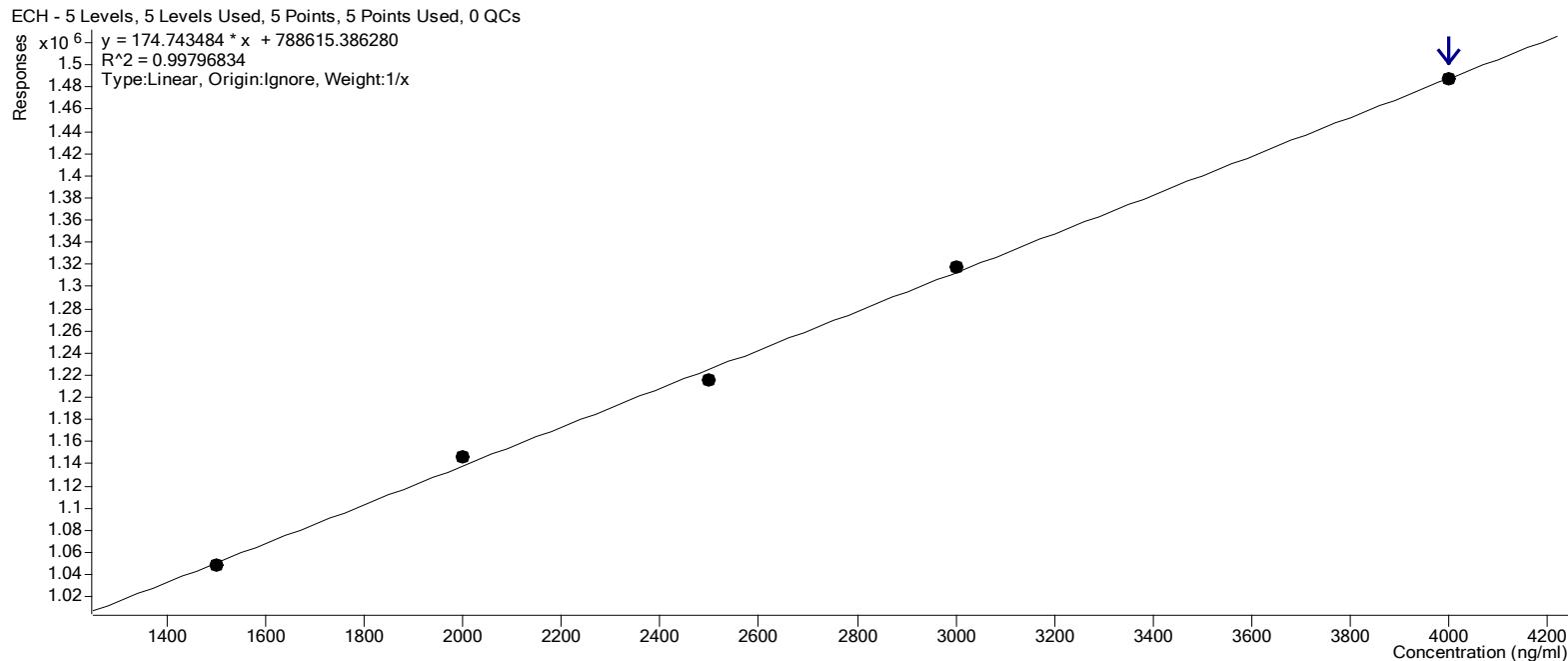
**Analysis of an aliquot of supernatant  
by GC-QTOF  
(vial dilution 1:2 with ethyl acetate)**



## Standard addition (2-chloroethanol)



## Standard addition (2-chloroethanol)



Concentration of 2-chloroethanol in sesame: 3.2 mg/kg

**Thank You  
for Your Attention**



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