

PROFICIENCY TEST 6, 2004

Incurred Residues of Pesticides in Tomato Homogenate

Final Report

Organiser:

Dr. Amadeo R. Fernández-Alba

Universidad de Almería
Edificio Químicas CITE I
Ctra. Sacramento s/n
04120 Almería
Spain
Phone: +34 950015034
Fax: +34 950015645
e-mail: amadeo@ual.es
<http://www.ual.es/GruposInv/EUPT6/presentation.htm>

Organising team at the University of Almería:

Ms. Paula Medina, Chemist
Mr. Octavio Malato, Chemist
Dr. Mariano Contreras, senior Chemist
Dr. Ana Agüera, senior Chemist
Dr. Milagros Mezcua, senior Chemist
Mrs. Susana López, Chemist

University of Almeria
University of Almeria
COHEXPHAL Laboratory
University of Almeria
University of Almeria
University of Almeria

Advisory Group:

Mr. Arne Andersson, Head of Division.
Dr. Antonio Valverde, senior Chemist.

National Food Administration, Uppsala, Sweden
Technical Auditor for pesticide residues of the Spanish Body for Accreditation (ENAC), Spain

Dr. Roma Tauler, senior Chemist.
Dr. Jerome Vial, senior Chemist.

University of Barcelona, UAB, Spain
Ecole Supérieure de Physique et de Chimie Industrielles, ESPCI, Paris, France

Dr. Miguel Gamón, senior Chemist.

Technical Auditor for pesticide residues of the Spanish Body for Accreditation (ENAC) Spain

Dr. André de Kok, senior Chemist.

Inspectorate for Health Protection, Amsterdam, The Netherlands

Dr. Tuija Pihlström, Chemist.
Mr. Stewart Reynolds, senior Chemist.

National Food Administration, Uppsala, Sweden
Central Science Laboratory York, United Kingdom

CONTENTS

- 1. INTRODUCTION**
- 2. TEST MATERIALS**
 - 2.1 Analytical methods
 - 2.2 Preparation of treated test material
 - 2.3 Preparation of blank test material
 - 2.4 Homogeneity test
 - 2.5 Stability tests
 - 2.6 Distribution of test material and protocol to participants
- 3. STATISTICAL METHODS**
 - 3.1 Background
 - 3.2 False positives and negatives
 - 3.3 Estimation of the assigned concentration
 - 3.4 Estimation of the target standard deviation
 - 3.5 z-Scores
 - 3.6 Combined scores
- 4. RESULTS**
 - 4.1 Summary of results reported
 - 4.2 Assigned concentration and target standard deviations
 - 4.3 Assessment of the laboratory performance
- 5. CONCLUSIONS**
- 6. SUGGESTIONS FOR FUTURE WORK**
- 7. REFERENCES**
- 8. ACKNOWLEDGEMENTS**
- APPENDIX 1. Homogeneity data
- APPENDIX 2. Histograms of residue data for each pesticide for all the laboratories
- APPENDIX 3. Results (mg/kg) and z-Scores for FFP and Qn
- APPENDIX 4. Graphical representation of z-Scores for FFP (25%)
- APPENDIX 5. Combined scores RSZ and SSZ for FFP (25%)
- APPENDIX 6. z-Score, 'Weighted Sum of z-Score' and graphical representation for laboratories classified in Category A.
- APPENDIX 7. Methods used by participants for determining pesticides
- ANNEX 1. Protocol and instructions. List of pesticides to be sought
- ANNEX 2. List of laboratories invited to participate in PT6

EUROPEAN COMMISSION'S PROFICIENCY TEST 6 ON PESTICIDE RESIDUES IN FRUIT AND VEGETABLES, 2004

The 6th European Commission's Proficiency Test was performed in 2004 using tomato homogenate. The tomatoes were grown in the south of Spain, in Almeria, and were treated with two different types of pesticide applications: a field treatment similar to normal commercial practice and a post harvest treatment where commercial formulations were applied using a microspray. Thirteen pesticides were used for the treatments. Participating laboratories were provided with a 'blank' tomato homogenate as well as the treated test material.

One hundred and thirty laboratories agreed to participate in the proficiency test.

The medians from the analytical data submitted were used to calculate the assigned concentrations of the thirteen pesticide residues. Using these, together with the standard deviations, z-Scores were calculated for every laboratory for the thirteen pesticides. The standard deviation was calculated in two ways, using a fit-for-purpose (FFP) target standard deviation based on the experience of the Advisory Group and also using Qn, robust statistics.

Classical procedures for summing z-Scores (SSZ and RSZ) were calculated for all the individual z-Scores of the participating laboratories. For the assessment of the overall laboratory performance a new criteria has also been applied. Only laboratories that fulfilled the criteria of detecting at least eleven of the thirteen pesticides (~90%) have been classified as having 'sufficient scope', and have been assigned to Category A. Within this category, the laboratories have been classified as 'good', 'satisfactory' or 'unsatisfactory'. All the other laboratories have been put into Category B, and classified as having 'insufficient scope'. For laboratories in Category B, individual z-Scores have been calculated, but their overall performance using the new criteria has not been assessed.

These results are presented in this report.

1. INTRODUCTION

The Council Directives (86/362/EEC and 90/642/EEC) provide for the organization and financial support for regular proficiency testing of the laboratories that perform analyses for their national monitoring programmes. These proficiency tests are performed in order to assure the quality, accuracy and comparability of the residue data sent by EU Member States to the European Commission and to the other Member States on an annual basis. The EU Commission's Proficiency Tests 1-5 were carried out from 1996-2003, and were all organized by the Swedish National Food Administration. This year, the University of Almeria, Spain, organized the 6th European Proficiency Test. Participation in this proficiency test was open to all official national or regional analytical laboratories involved in the determination of pesticide residues in fruit and vegetables in Member States of the EU and, additionally, Norway and Romania as invited countries.

A total of one hundred and thirty laboratories in Europe agreed to participate in the 6th Proficiency Test. The test material, 400 g of tomato homogenate containing residues of pesticides, together with 400 g of blank tomato homogenate, were shipped to participants on 17th, 18th, 25th and 31st May, 2004. The deadline for submission of results to the organiser was 12th July 2004. The participants were provided with a list of fifty-seven pesticides (Annex 1), which might be present in the test material and they were asked to determine the levels of all the pesticides they detected. Participants were also asked to analyse the blank test material and report residues of the pesticides they found from the same list of pesticides. This blank material was intended for use in performing recovery experiments with the pesticides found in the test material and for the preparation of matrix-matched standards, if necessary.

2. TEST MATERIALS

2.1 Analytical methods

Three analytical methods, described briefly below, were used for the homogeneity and stability tests as well as other procedures by the organisers at the laboratory of Coexphal (Almeria):

- GC method [1]: ethyl acetate extraction in the presence of sodium sulfate, filtration, addition of more sodium sulfate, evaporation, re-dissolution in cyclohexane and determination by GC-MS/MS.
- LC method [2]: ethyl acetate extraction in the presence of sodium sulfate and addition of sodium hydroxide, filtration, addition of more sodium sulfate, evaporation, re-dissolution in methanol, and determination by LC-MS/MS.
- For dithiocarbamates [3]: acid digestion was carried out, followed by UV determination of CS₂

Thiabendazole, imidacloprid and azoxystrobin (Table 1) were determined using the LC-MS/MS method. All other pesticides except dithiocarbamates, (Table 1) were analysed using the GC method. For confirmation purposes, MS/MS spectra were used except in the case of dithiocarbamates.

2.2 Preparation of treated test material

Before the preparation of the test material, the pesticides and suitable residue levels for the study were selected and recommended by the Advisory Group, which had been appointed for Proficiency Test 6. The tomatoes were grown in the south of Spain, in Almeria. A field treatment was undertaken (Table 1). A portion of the crop was sampled and analysed to check the residue levels that had been incurred. Following the field treatment one hundred kilograms of tomatoes were sampled and treated post-harvest with a commercial formulation dissolved in water and applied to the tomatoes using a microspray (Table 1). Different formulations were applied for each of the pesticides allowing one hour between applications. A portion was taken and analysed to check the residue levels. When the tomatoes contained residue levels close to those agreed by the Advisory Group they were frozen and chopped using liquid nitrogen and a mincer, then mixed in a constantly spinning container especially designed and built up by the organiser until a homogeneous material was obtained. One pesticide, metalaxyl, was found to be present as a contaminant in the test material (but not in the blank), either because of cross contamination in the field, or because a contaminant was present in one of the commercial formulations. The concentration of metalaxyl was below the Limit of Determination (LOD) or minimum required performance level (MRPL) recommended by the PT6 Advisory Group that must be used by the participating laboratories. Therefore the residue data reported for metalaxyl is presented in this report for information purposes only. Portions of 400 g per bottle, of the well-mixed homogenate, were weighed out into screw-capped polyethylene plastic bottles, sealed, and stored in a freezer at about -20 °C prior to distribution to participants.

Table 1. Pesticides, field-treated and following harvest.

Pesticide	Field-treatment	Post-harvest treatment
Acrinathrin		Yes
Azoxystrobin		Yes
Bromopropylate		Yes
Chlorothalonil		Yes
Diazinon		Yes
Dimethoate		Yes
Endosulfan		Yes
Imazalil		Yes
Imidacloprid	Yes	
Dithiocarbamates (CS ₂)		Yes
Oxydemeton-methyl		Yes
Procymidone		Yes
Thiabendazole	Yes	

2.3 Preparation of blank test material

The tomatoes to be used for the production of the blank test material were organically grown in the same area as the test material, in the south of Spain, in Almería. The homogenate was prepared in the same way as the incurred test material described above. The blank test material did not contain any residues of the pesticides listed in Annex 1, at, or above the reporting levels applied.

2.4 Homogeneity test

Ten bottles were randomly chosen from those stored in the freezer and analyses were performed on duplicate portions taken from each bottle. The sequence of analyses was determined using a table of randomly generated numbers. The injection sequence of the 20 extracts analysed by GC and LC was also randomly chosen. The quantification by GC and LC was performed using a 3-point calibration curve and matrix matched standards. One standard mixture was used, for GC and LC. 'Dithiocarbamates' were determined as carbon disulfide, following acid digestion and using a UV detector.

The statistical evaluation was performed according to the international harmonized protocol prepared by AOAC, ISO and IUPAC [4, 5]. The individual residue data from the homogeneity tests are given in Appendix 1. The results of the statistical analyses are given in Table 2A-B. The acceptance criteria to be sufficiently homogenous was that Fcritical > F. The F-test was passed for all pesticides (p = 0.05).

Table 2A. Statistical evaluation of the homogeneity test data (n=20 analyses)

Pesticide	Acrinathrin	Azoxystrobin	Bromopropylate	Chlorothalonil	Diazinon	Dimethoate	Endosulfan
Mean, mg/kg	0.298	0.241	0.468	1.65	3.97	0.16	0.38
SD, mg/kg	0.006	0.005	0.001	0.01	0.102	0.001	0.004
F-critical	3.18	3.18	3.18	3.18	3.17	3.18	3.18
F	1.15	1.02	1.11	1.37	1.02	1.56	1.21
Passed/Failed	Passed	Passed	Passed	Passed	Passed	Passed	Passed

Table 2B. Statistical evaluation of the homogeneity test data (n=20 analyses)

Pesticide	Imazalil	Imidacloprid	Dithiocarbamates (CS ₂)	Metalaxyl	Oxydemeton-methyl	Procymidone	Thiabendazole
Mean, mg/kg	0.188	0.238	0.787	0.048	0.199	0.418	0.331
SD, mg/kg	0.007	0.002	0.021	0.02	0.001	0.002	0.005
F-critical	3.18	3.18	3.18	3.18	3.18	3.18	3.18
F	1.10	1.44	1.02	1.25	2.20	1.20	1.45
Passed/Failed	Passed	Passed	Passed	Passed	Passed	Passed	Passed

Thus, overall, the F-test showed that the material was sufficiently homogeneous for the PT with respect to all 13 pesticides and the SDs demonstrated that the analytical precision was acceptable.

2.5 Stability tests

The three analytical methods described briefly above (at point 2.1) were also used for the stability tests. The tests were performed on two occasions. On each occasion, one bottle stored in the freezer was chosen randomly and duplicate analyses were performed.

The two occasions were:

- Day 1: coinciding with the first sample shipment, which took place 17th /18th of May 2004.
- Day 2: shortly after the deadline for reporting results, 12th July 2004.

The individual results are given in Tables 3A-B. In general, these tests did not show any significant decrease in the levels of the pesticides and were considered to be acceptable as a stable test material. However, there was a significant decrease (>10%) for dithiocarbamates from Day 1 to Day 2 although it was considered a stable test material.

Table 3 A. Statistical test for analytical precision

Pesticide	Acrinathrin	Azoxystrobin	Bromopropylate	Chlorothalonil	Diazinon	Dimethoate	Endosulfan
Day 1 (mg/kg)							
1st analysis	0.311	0.250	0.464	1.657	4.023	0.153	0.384
2nd analysis	0.305	0.257	0.469	1.659	4.027	0.157	0.385
Mean 1	0.308	0.253	0.467	1.658	4.025	0.155	0.385
Day 2 (mg/kg)							
1st analysis	0.288	0.231	0.438	1.610	3.678	0.147	0.372
2nd analysis	0.294	0.234	0.432	1.609	3.680	0.145	0.375
Mean 2	0.291	0.233	0.435	1.610	3.679	0.146	0.374
(Mean 1-Mean 2)	0.055	0.079	0.069	0.029	0.086	0.058	0.029
Mean 1							

Table 3 B. Statistical test for analytical precision

Pesticide	Imazalil	Imidacloprid	Dithiocarbamates (CS ₂)	Metalaxyl	Oxydemeton -methyl	Procymidone	Thiabendazole
Day 1 (mg/kg)							
1st analysis	0.197	0.244	0.857	0.050	0.202	0.417	0.325
2nd analysis	0.200	0.240	0.850	0.050	0.205	0.420	0.328
Mean 1	0.199	0.242	0.854	0.050	0.204	0.419	0.327
Day 2 (mg/kg)							
1st analysis	0.193	0.239	0.759	0.044	0.194	0.425	0.345
2nd analysis	0.195	0.234	0.752	0.043	0.198	0.42	0.344
Mean 2	0.194	0.237	0.756	0.044	0.196	0.423	0.345
(Mean 1-Mean 2)							
Mean 1	0.025	0.021	0.115	0.120	0.039	-0.010	-0.055

2.6 Distribution of test material and protocol to participants

One bottle of incurred test sample and one bottle of blank test material were shipped in boxes containing dry ice. The samples were sent on the 17th and 18th May, 2004. Unfortunately the 20th May was a holiday in nearly all European countries and only one country notified the organiser of this in advance. Because of this problem, the receipts of many shipments were delayed and the test materials had thawed out. This meant that the organisers were obliged to send out repeat shipments to some countries. After this delay, all labs received their test material (with an exception of one lab that did not accept it). Because of the delay, the deadline for reporting results was extended by one week.

Before shipment of the samples, the laboratories received full instructions (Annex 1) for the receipt, storage and analysis of the test materials, although they were encouraged to use their normal sample receipt procedure and methods of analysis. These instructions were uploaded onto the web page built especially for this Proficiency Test. There were two parts, *the list of pesticides MRPLs*, which was the minimum required performance level, and the *Protocol*, with a common code to access through the web page. This information was notified by email to all participant labs so confidentiality was maintained throughout the entire Proficiency Test 6.

3. STATISTICAL METHODS

3.1 Background

3.1.1 Proficiency Tests 1-5

For the previous Proficiency Tests 1-5, different methods for the estimation of the assigned values and standard deviations (SDs) of the analytical data were used. The arithmetic mean values, after elimination of outliers, were used for Proficiency Tests 1-2, while the SDs were calculated using the Horwitz equation.

In Proficiency Test 3, the median of the results was chosen as the best estimate of the true concentration. Estimates of the standard deviations were compared using four methods. The traditional approach (ISO 5725-Part 2) with outlier elimination: robust statistics using both the Qn and Swiss methods; and a fixed target value (fit-for-purpose) as stipulated by the Advisory Group. The two robust statistical methods, for the estimation of the SD gave only marginal differences in the results, whilst the other two methods produced results that differed significantly.

In Proficiency Test 4, the same two robust statistical methods were compared and again only marginal differences were found.

z-Scores were used in all the five previous proficiency tests for the assessment of the laboratory performances for individual pesticides.

The sums of scores, the RSZ, SSZ and RLP were also calculated as for the last Proficiency Test (PT5).

3.1.2 Proficiency Test 6

In this proficiency test the median was used to obtain the assigned concentration values for each pesticide.

This was then used to calculate the z-Score. The SD was calculated by using the FFP value, which was assigned as 25%. Furthermore Qn was also calculated as SD value.

Sum of z-Scores: The RSZ and SSZ were also used but an additional new criterion was introduced for PT6 in agreement with the Advisory Group as well as DG Sanco. This was used as the basis to separate the labs into two categories, A and B. To be in Category A, sufficient scope had to have been demonstrated. This scope was that 11 or more results must have been submitted with no false positives. All labs reporting less than 11 results, or having reported a false positive were put into Category B.

3.2 False positives and negatives

3.2.1 False positives

In principle, pesticide residues unknown to the organizer that were reported by a small numbers of participant laboratories were treated as false positives, considering them only when the reported concentration was above their RL or MRPL. If the residue was below the laboratory's RL, then the organisation recommends to these labs should, in future PTs, ensure that they comply with the instructions issued by the organiser for reporting results.

3.2.2 False negatives

Results that did not reveal pesticide residues in the test material, although they were actually present, were considered to be false negatives, although there was one exception. Before taking a decision, the true value, the MRPL and the labs RL were all considered. Only if the RL was above the assigned value, and considering what was stated in the Protocol, the laboratories were not penalised, and no z-Score was calculated. If the RL was below the true value but above the MRPL, it was considered to be a false negative result. Laboratories should have adjusted their reporting levels to meet the MRPLs requested by the organisation.

z-Scores for false negatives were calculated in two ways: if the RL was above the MRPL, then the MRPL was used, and if the RL was below the MRPL then the RL was used, if labs did not report a value for their RL, then the MRPL was used.

Metalaxyl was at a lower residue level than the MRPL, and was therefore not considered in the overall statistical assessment of laboratory performance.

3.3 Estimation of the assigned concentration

To calculate the assigned values, the medians of all the reported results were used. Individual results reported that expressed uncertainty such as detected (**D**) or using a '**>**' sign were not used.

3.4 Estimation of the target standard deviation

Two procedures were used to calculate the standard deviation (SD); one using fixed standard deviations (fit-for-purpose, FFP) based on individual's experience and as agreed by the Advisory Group. The other was using the robust statistic Qn. (Appendix 3).

The standard deviation (SD) was calculated as a result of applying these procedures and multiplying by the assigned value or true concentration.

3.4.1 Fixed standard deviations (fit-for-purpose)

The relative standard deviation (RSD) was considered to be 25 %, according to the Advisory Group agreement, and also what was concluded from the discussion session on proficiency testing at EPRW 2004 in Stockholm, Sweden. This was to apply the same RSD to all the pesticides.

3.4.2 Robust statistic: Qn

Using Qn [6] the SD was calculated from the results after the differences between all laboratory results had been calculated. The SD is the 25th percentile of these differences, multiplied by a factor of 2.2194.

3.5 z-Scores

A z-Score for each laboratory/pesticide combination was calculated according to the following equation:

$$z = (x - X) / \sigma \quad \text{Eq.1}$$

Where:

- x is the result reported by the participant or the RL or MRPL for those labs not having detected the pesticide present in the sample (the MRPL was only used when no RL was reported)
- X is the assigned value or true concentration
- σ is the target standard deviation (using the median and FFP of 25%)

No calculation of z-Score has been performed for false positive results or laboratories with a RL above the true assigned value.

3.6 Combined scores

In order to evaluate each laboratory's overall performance, taking into account all the pesticides analysed, three methods were used to combine z-Scores the re-scaled sum of z-Scores (RSZ), the sum of squared z-Scores (SSZ) and a new procedure for combining z-Scores called the 'Weighted sum of z-Scores'.

3.6.1 RSZ

The RSZ was calculated for all z-Score values for each laboratory according to:

$$RSZ = \Sigma z / n$$

Where n is the number of z-Scores

The RSZ gives an averaged score for all pesticides analysed and indicates if a laboratory has a consistent bias in its results.

3.6.2 SSZ

The SSZ is the sum of all squared z-Scores. It was calculated for all z-Scores for each laboratory according to:

$$SSZ = (z\text{-Score}_1)^2 + (z\text{-Score}_2)^2 + \dots + (z\text{-Score}_n)^2$$

Where n is the number of z-Scores

3.6.3 Weighted sum of z-Scores

This function has been applied this year as a first approach for improvement on how to get a global evaluation of the participating laboratories and it was only applied to labs with sufficient scope (those in Category A), i.e. those labs that have reported 11 or more results with no false positives. The individual z-Scores were multiplied by weighted factors.

$$\text{'Weighted sum of z-Scores' (Z)} = \frac{\sum_{i=0}^{i \leq 2} |z| \cdot 1 + \sum_{i>2}^{i \leq 3} |z| \cdot 3 + \sum_{i>3}^{\infty} |z| \cdot 5}{n}$$

So for each lab,

- The first factor is the sum of all their z-Scores between zero to two, multiplied by one.
- The second factor is the sum of all their z-Scores greater than two but less than or equal to, three, multiplied by three.
- The third factor is the sum of all their z-Scores greater than three, multiplied by five.

The three factors were then divided by the number of reported results (n) from each lab.

The 'Weighted sum of z-Scores' has then been used to produce an overall classification of laboratories as 'good', 'satisfactory' or 'unsatisfactory' according to:

$ z \leq 2$	good
$2 < z \leq 3$	satisfactory
$ z > 3$	unsatisfactory

In this way, a simple, single combined value, very similar to the single z-Scores, is produced that should help to promote laboratories to analyse of a greater number of target compounds.

This function has not been applied to those participants with insufficient scope, in Category B, which is for the laboratories reporting less than 11 results, or with false positives.

4. RESULTS

4.1 Summary of results reported

One hundred and thirty laboratories agreed to participate in this proficiency test and one hundred and twenty-six submitted results, of which one arrived after the deadline.

One laboratory declined to participate for a special reason, two submitted combined results together, and the two others did not submit any results.

The results of the one hundred twenty-six participating laboratories are presented in this report.

One hundred and twenty eight from the 130 laboratories were from EU Member States, of which twenty-eight were from the new EU countries.

A summary of the results reported can be seen in Table 4.

Table 4 Types of Results

Pesticides	Nº of Results Reported	Nº of NA Reported	Nº of ND Reported	% of the Total Results *
Acrinathrin	69	53	4	94.5
Azoxystrobin	92	27	7	92.9
Bromopropylate	118	7	1	99.2
Chlorothalonil	114	9	3	97.4
Diazinon	124	2	0	100
Dimethoate	114	4	8	93.4
Endosulfan	119	5	2	98.3
Imazalil	88	23	15	85.3
Imidacloprid	52	69	5	91.1
Dithiocarbamates expressed as CS ₂	78	45	3	96.3
Metalaxyl (informative)	73	13	40	64.3
Oxydemeton-methyl	42	73	11	79.3
Procymidone	120	6	0	100
Thiabendazole	88	31	7	92.6

* The % of the total results are calculated from the results of the labs that sought these compounds excluding the number of NAs

NA = Not analysed

ND = Not detected

The laboratories that agreed to participate are listed in Annex 2. All data reported by the participants, including the analytical results (Appendix 3), recoveries and analytical methods used, are given in Appendix 7. For an explanation of the symbols used in the tables, see Annex 1.

The participating laboratories did not report any residues in the blank test material from the pesticides listed in Annex 1.

4.1.1 False positives

Some laboratories reported finding additional pesticides in the incurred test materials.

These pesticides and the residue levels reported are presented in Table 5. They were not considered to be false positives, if the concentration was below the laboratory's reporting limit or the MRPLs, or if the pesticides were not intended to be sought from the list of the fifty-seven pesticides included in the EUPT6 (Annex I).

Three labs would have been classified in Category A, but have been put into Category B, because they reported a false positive.

Table 5. Laboratories that reported false positives

Pesticides	Lab Code	Concentration	RL (mg/kg)	MRPL (mg/kg)
Carbendazim	061	0.037	0.02	0.1
	064	0.135		0.1
	103	0.145	0.05	0.1
Fenhexamid	026	0.343	0.05	0.05
Lambda-cyhalothrin	090	0.10	0.02	0.02
	092	0.026	0.02	0.02
	105	0.043	0.02	0.02
Pirimiphos-methyl	120	0.15	0.01	0.05
Tolyfluanid	119	0.060	0.03	0.05
Vinclozolin	105	0.125	0.05	0.05

4.1.2 False negatives

Results reported as not detected, for pesticides that were present in the samples, were considered to be false negatives except for those having a reporting limit above the assigned value. These are given in Table 6.

Table 6 Types of non-detected (ND) results reported by participants

Nº of False negatives Median	Acrinathrin (MRPL 0,05) 0,284	Azoxystrobin (MRPL 0,05) 0,225	Bromopropionate (MRPL 0,05) 0,490	Chlorothalonil (MRPL 0,01) 1,626	Diazinon (MRPL 0,02) 3,955	Dimethoate (MRPL 0,02) 0,132	Endosulfan (MRPL 0,05) 0,344
ND: RL> X	0	0	0	0	0	0	0
ND: RL≤MRPL	5	5	1	3	0	5	2
ND: RL>MRPL	0	2	0	0	0	3	0
Total	5	7	1	3	0	8	2

RL is the reporting limit given by the laboratory

MRPL is the minimum required performance level

X is the true concentration

Nº of False negatives Median	Imazalil (MRPL 0,02) 0,167	Imidacloprid (MRPL 0,05) 0,232	Dithiocarbamates expressed as CS ₂ (MRPL 0,05) 0,810	Oxydemeton-methyl (MRPL 0,02) 0,199	Procymidone (MRPL 0,02) 0,412	Thiabendazole (MRPL 0,05) 0,313
ND: RL> X	3	0	0	1	0	1
ND:RL≤MRPL	8	5	1	7	0	3
ND: RL>MRPL	7	0	2	4	0	4
Total	15	5	3	11	0	7

RL is the reporting limit given by the laboratory
 MRPL is the minimum required performance level
 X is the true concentration

4.1.3 Distribution of data

The distributions of the individual mean values, including false negative results, for the fourteen pesticides (metalaxyl, for information only) are plotted as histograms from the analytical data supplied by the 126 laboratories.

4.2 Assigned concentrations and target standard deviations

To calculate the assigned values the medians of all the reported results were used. The standard deviation was obtained in two ways, FFP and Qn. z-Scores were calculated for both.

The results for the assessment of the assigned residue levels and for the different calculations of standard deviations are given in Appendix 3.

For acrinathrin, azoxystrobin, chlorothalonil, dimethoate, imazalil, dithiocarbamates, oxydemeton-methyl and thiabendazole the z-Score using the Qn value was higher than that using the FFP value. It was decided therefore by the organisation and the Advisory Group, that the z-Scores using Qn for the RSD should not be represented graphically, and the combined z-Scores were not calculated.

4.3 Assessment of the laboratory performance

4.3.1 z-Scores

The criteria for defining the acceptability of the z-Scores are:

$ z \leq 2$	acceptable
$2 < z \leq 3$	questionable
$ z > 3$	unacceptable

In Appendix 3 the individual z-Scores are presented, together with the median for each laboratory and pesticide. Both the z-Scores calculated using the RSD from FFP and Qn are given. False negatives are included in the z-Scores.

In Appendix 4 the graphic representations of the z-Scores using the RSD from FFP are presented. No z-Scores from false negatives have been presented.

4.3.2 Combined z-Scores

The classical combined z-Scores: RSZ and the SSZ values are listed in Appendix 5 for all laboratories, used as accepted formulas.

Appendix 6 shows a table with the values of individual z-Scores for each pesticide and the new combined 'Weighted sum of z-Scores' for those labs in Category A. It is important to mention that it is a new criterion and it has been used as a first approach and further improvement can be expected. In this category are the laboratories that have reported 11 or more results and have not reported any false positives. A graphical representation of the results for these laboratories can also be found.

Sixty from the one hundred and twenty-six laboratories are in Category A (48%), from which 60% are classified as 'good', 18% as 'satisfactory' and 21% as 'unsatisfactory'.

Sixty-six of the one hundred and twenty-six laboratories reported results for less than eleven pesticides. Several of these labs could also have been classified as 'good', but they failed to report results for the required 11 compounds. Thirty of the sixty-six laboratories detected nine or ten pesticides.

Table 7 Laboratories that are either in Category A, or in Category B.

Category A Laboratories: those with sufficient scope, having reported 11 to 13 results, and classified into good, satisfactory and unsatisfactory.

Num of Reported Pesticides	Lab Code	Scope	Classification
13	Lab001	Sufficient	Good
13	Lab018	Sufficient	Good
13	Lab019	Sufficient	Good
13	Lab020	Sufficient	Good
13	Lab021	Sufficient	Good
13	Lab023	Sufficient	Good
13	Lab025	Sufficient	Good
13	Lab028	Sufficient	Good
13	Lab045	Sufficient	Good
13	Lab078	Sufficient	Good
13	Lab085	Sufficient	Good
13	Lab086	Sufficient	Good
13	Lab095	Sufficient	Good
13	Lab096	Sufficient	Good
13	Lab097	Sufficient	Good
13	Lab098	Sufficient	Good
13	Lab100	Sufficient	Good
13	Lab124	Sufficient	Good
13	Lab125	Sufficient	Good
13	Lab130	Sufficient	Good
13	Lab030	Sufficient	Satisfactory
13	Lab031	Sufficient	Satisfactory
13	Lab032	Sufficient	Satisfactory
13	Lab038	Sufficient	Satisfactory
13	Lab123	Sufficient	Satisfactory
13	Lab128	Sufficient	Satisfactory
13	Lab024	Sufficient	Unsatisfactory
13	Lab039	Sufficient	Unsatisfactory

Num of Reported Pesticides	Lab Code	Scope	Classification
13	Lab041	Sufficient	Unsatisfactory
13	Lab043	Sufficient	Unsatisfactory
13	Lab044	Sufficient	Unsatisfactory
13	Lab065	Sufficient	Unsatisfactory
12	Lab007	Sufficient	Good
12	Lab009	Sufficient	Good
12	Lab013	Sufficient	Good
12	Lab022	Sufficient	Good
12	Lab027	Sufficient	Good
12	Lab042	Sufficient	Good
12	Lab057	Sufficient	Good
12	Lab071	Sufficient	Good
12	Lab087	Sufficient	Good
12	Lab107	Sufficient	Good
12	Lab122	Sufficient	Good
12	Lab129	Sufficient	Good
12	Lab029	Sufficient	Satisfactory
12	Lab035	Sufficient	Satisfactory
12	Lab046	Sufficient	Satisfactory
12	Lab017	Sufficient	Unsatisfactory
12	Lab047	Sufficient	Unsatisfactory
12	Lab076	Sufficient	Unsatisfactory
12	Lab127	Sufficient	Unsatisfactory
11	Lab002/3	Sufficient	Good
11	Lab004	Sufficient	Good
11	Lab099	Sufficient	Good
11	Lab121	Sufficient	Good
11	Lab034	Sufficient	Satisfactory
11	Lab106	Sufficient	Satisfactory
11	Lab012	Sufficient	Unsatisfactory
11	Lab016	Sufficient	Unsatisfactory
11	Lab033	Sufficient	Unsatisfactory

Category B Laboratories: those with an insufficient scope, having reported less than 11 results or a false positive.

Num of Reported Pesticides	Lab Code	Scope	Reported False positive
12	Lab061	Insufficient	Yes
12	Lab119	Insufficient	Yes
11	Lab090	Insufficient	Yes
10	Lab005	Insufficient	
10	Lab008	Insufficient	
10	Lab014	Insufficient	
10	Lab015	Insufficient	
10	Lab037	Insufficient	
10	Lab058	Insufficient	
10	Lab064	Insufficient	Yes

Num of Reported Pesticides	Lab Code	Scope	Reported False positive
10	Lab066	Insufficient	
10	Lab070	Insufficient	
10	Lab077	Insufficient	
10	Lab082	Insufficient	
10	Lab092	Insufficient	Yes
10	Lab101	Insufficient	
10	Lab103	Insufficient	Yes
10	Lab110	Insufficient	
10	Lab118	Insufficient	
9	Lab026	Insufficient	Yes
9	Lab036	Insufficient	
9	Lab062	Insufficient	
9	Lab067	Insufficient	
9	Lab068	Insufficient	
9	Lab069	Insufficient	
9	Lab072	Insufficient	
9	Lab075	Insufficient	
9	Lab079	Insufficient	
9	Lab094	Insufficient	
9	Lab104	Insufficient	
9	Lab113	Insufficient	
9	Lab114	Insufficient	
9	Lab115	Insufficient	
8	Lab011	Insufficient	
8	Lab040	Insufficient	
8	Lab054	Insufficient	
8	Lab056	Insufficient	
8	Lab063	Insufficient	
8	Lab073	Insufficient	
8	Lab093	Insufficient	
8	Lab102	Insufficient	
8	Lab111	Insufficient	
8	Lab112	Insufficient	
8	Lab116	Insufficient	
8	Lab117	Insufficient	
7	Lab048	Insufficient	
7	Lab049	Insufficient	
7	Lab059	Insufficient	
7	Lab081	Insufficient	
7	Lab084	Insufficient	
7	Lab089	Insufficient	
7	Lab091	Insufficient	
7	Lab108	Insufficient	
7	Lab109	Insufficient	
6	Lab006	Insufficient	
6	Lab010	Insufficient	
6	Lab051	Insufficient	
6	Lab083	Insufficient	

Num of Reported Pesticides	Lab Code	Scope	Reported False positive
5	Lab055	Insufficient	
5	Lab088	Insufficient	
4	Lab060	Insufficient	
4	Lab120	Insufficient	Yes
3	Lab050	Insufficient	
3	Lab105	Insufficient	Yes
2	Lab052	Insufficient	
0	Lab053	Insufficient	
0	Lab074	No Results	
0	Lab080	No Results	
0	Lab126	No Results	

5. CONCLUSIONS

Twenty-five countries, including Norway and Romania that do not belong to the EU, participated in this 6th European Proficiency Test. Eight countries were new European members.

The majority of the invited laboratories submitted the results of their analyses of the pesticides present in the homogenate tomato test material (126 from 130). Only 48% from 126 laboratories were able to determine 11 or more of the pesticides present in the test material. These laboratories are classified in Category A (sufficient scope) while the rest are in Category B (insufficient scope). A large number also reported results for the contaminant metalaxyl (73 from 126).

The assigned values were calculated using ALL the results submitted.

For each laboratory/pesticide combination, z-Scores based on the FFP and Qn/median were calculated. Only for the FFP for 25%, there is a graphical representation. The z-Scores were used to assign an overall performance using the descriptive classifications 'acceptable', 'questionable' or 'unacceptable' (Eq. 1). Typically acceptable submitted results were around an 85%, 7% of 'questionable' and a 5% of 'unacceptable'. So, it is noticeable that the reported results from laboratories have a high-quality although there are variable results according to the number of pesticides reported. The number of reported results varies from 42 (for oxydemeton-methyl) to 124 (for diazinon).

The classical combined z-Scores were calculated for all the labs and a new criterion was used as this year first approach. It is a very valuable formula that represents very easy and significantly the laboratories performance although as it is new, it could be modified in the future and its acceptance has to be valorised. It was applied for those labs with sufficient scope (Category A). This new classification was used to assign a combined value: 'good', 'satisfactory' or 'insufficient'. In this Category A, there are 60 from 126 laboratories from which 60% are classified as 'good', 18% as 'satisfactory' and 21% as 'Insufficient'. The rest of the labs (66 from 126) have been assigned to Category B, with insufficient scope.

A significant decrease in the number of false positive and negatives reported from previous PTs has been noticed. Some explanation for this may be because more laboratories are now using MS/MS techniques.

6. SUGGESTIONS FOR FUTURE WORK

The Advisory Group appointed for Proficiency Test 6 recommend maintaining the use of the 25% FFP criteria for use in the statistical treatment of future PTs. Thus the results from future PTs will be more comparable.

To give these PTs a greater relevance, it was pointed out the convenience of announcing future actions to be held from the results derived from future reports. The possibility of relating the codes with the names of the labs was considered. The usefulness of including in the Final Report a harmonized list of target pesticides agreed by the Advisory Group for use in the next PT was also considered, so that participant laboratories could obtain standards on time and validate their methods in order to broaden the scope of the pesticides that they look for.

It is also worth reiterating the problems encountered this year with the shipment of samples by asking participating labs to help the organisers by making every effort to ensure that the test material can be put into freezer storage immediately on receipt.

More over, it would be very interesting to recalculate the z-Scores and 'Weighted sum of z-Scores' for previous PTs using the 25% FFP, so that directly comparable data are obtained and clearer information regarding any trends in labs performances can be seen.

To evaluate false negatives, the Protocol should be changed and MRPLs should be applied for cases where the laboratory either gives no Reporting Limit (RL) or gives a RL higher than MRPL (insufficient sensitivity). A RL given by the laboratory, which is below, or equal to the MRPL, should be used for the calculation of z-Scores (as in this PT).

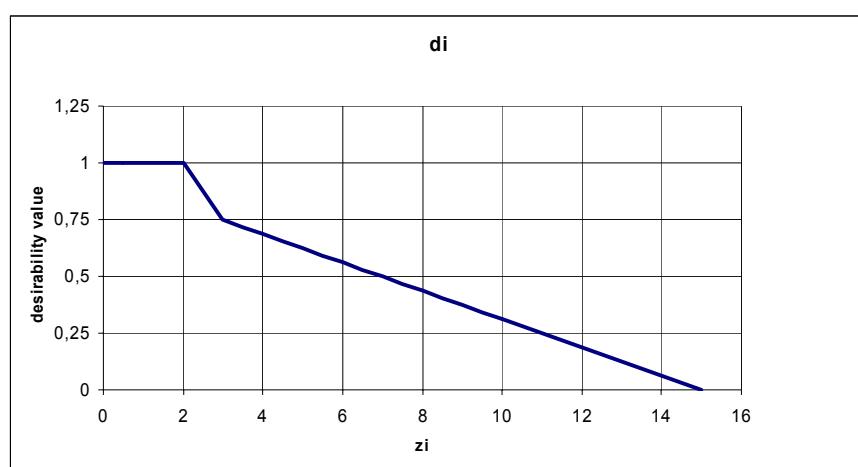
For future work a Desirability Function (D) proposal will be considered defined as the product of individual desirability functions d_i .

$$D = \prod_i d_i$$

D takes value between 0 (the less desirable) and 1 (the most desirable). Each individual d_i is built segment by segment as follows:

- For $z < 2$ $d_i = 1$
- For $2 < z < 3$ d_i decreases linearly from 1 to 0.75
- For $3 < z < 15$ d_i decreases linearly from 0.75 to 0
- For $z > 15$ $d_i = 0$

The figure bellows illustrate this function.



The D function is obtained by multiplying all the individual desirability obtained for all the compounds. The D values will range from 0 to 1. It can be expressed as a percentage since 1 = 100%.

7. REFERENCES

1. Multiresidue method for the analysis of multiclass pesticides in agricultural products by gas chromatography-tandem mass spectrometry. Agüera A., Contreras M., Crespo J., Fernández-Alba A.R., *Analyst*, 2002, 127, 347-354
2. One-year routine application of the new method based on liquid chromatography-tandem mass spectrometry to the analysis of 16 multiclass pesticides in vegetables samples. Agüera A., López S., Fernández-Alba A.R., Contreras M., Crespo J., Piedra L., *JAOAL*, 1045 (2004), 125-135
3. Keppel Method using Winell slight modification. *G.E. Keppel, JAOAC_hem*, 54, 528-532 (1971) and *B. Winell. Var Föda*, 27, 94-102 (1975)
4. Thompson M. and Wood R. International Harmonized Protocol for Proficiency Testing of (Chemical) Analytical Laboratories. *J. AOAC_{Int.}* 1993; 76 (4): 926-940.
5. Fearn T. And Thompson M. A new test for sufficient homogeneity. *Analyst* , 2001, 126
6. Estimation of Variance components with high breakdown point and high efficiency. Muller C. H. and Uhlig S., *Biometrika*, 88 (2001) 353-366,

8. ACKNOWLEDGEMENTS

The organiser is much obliged to the European Commission for funding this 6th European Proficiency Test.

The organiser wishes to thank the members of the Advisory Group for the valuable and knowledgeable advice.

Many thanks too for the cooperation, statistical calculations and assessments.

Our appreciation to the University of Almeria and the organising team.

Special thanks to Carmelo Rodríguez for Qn calculation.

Special mention to the greenhouse holder Cuevas Bio Sat, for letting us used the field.

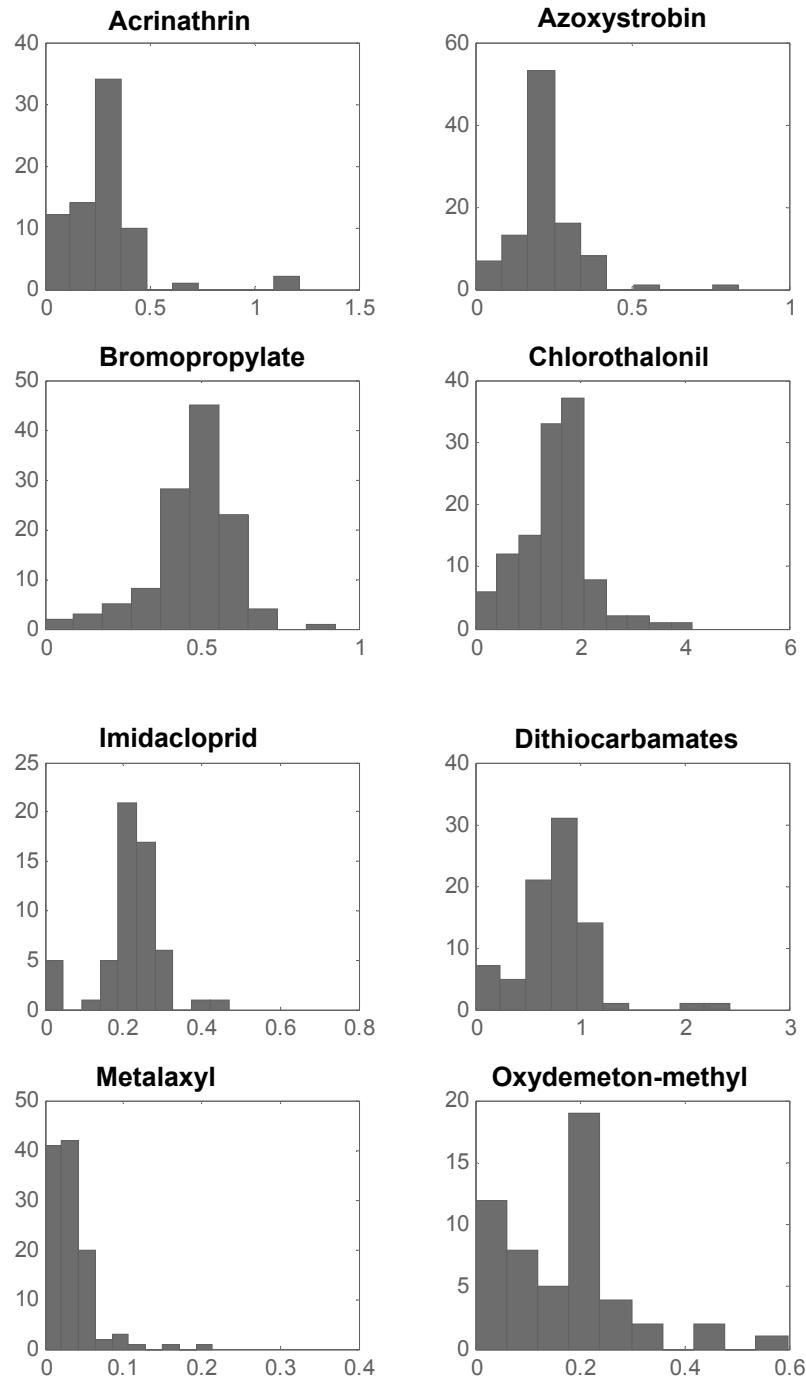
APPENDIX 1. Homogeneity Data

Bottle	Acrinathrin mg/kg		Azoxystrobin mg/kg		Bromopropylate mg/kg		Chlorothalonil mg/kg		Diazinon mg/kg	
1	0.308	0.307	0.250	0.251	0.467	0.466	1.66	1.66	4.002	4.003
2	0.308	0.307	0.249	0.248	0.466	0.468	1.66	1.66	4.002	4.008
3	0.299	0.298	0.235	0.236	0.470	0.470	1.63	1.63	4.004	4.005
4	0.289	0.289	0.237	0.237	0.469	0.469	1.65	1.65	4.015	4.018
5	0.295	0.296	0.238	0.238	0.468	0.468	1.63	1.63	3.995	3.996
6	0.294	0.294	0.240	0.240	0.467	0.468	1.65	1.65	4.004	4.005
7	0.294	0.294	0.245	0.245	0.468	0.468	1.66	1.65	3.670	3.675
8	0.299	0.298	0.242	0.243	0.468	0.469	1.65	1.65	3.998	3.999
9	0.298	0.299	0.239	0.238	0.467	0.468	1.66	1.66	4.009	4.008
10	0.299	0.299	0.238	0.239	0.467	0.467	1.63	1.64	4.002	4.001

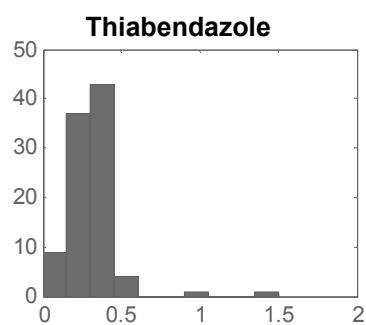
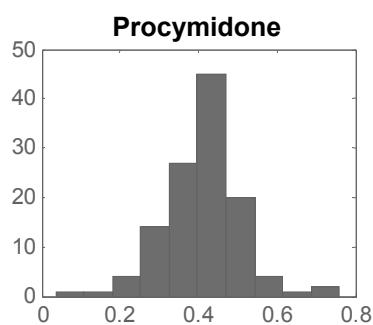
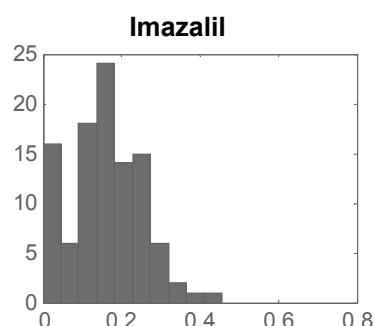
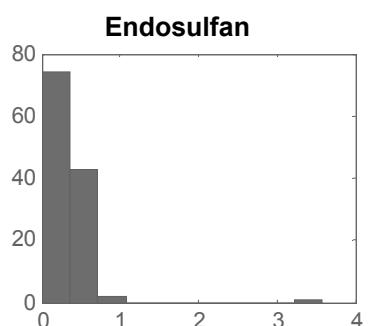
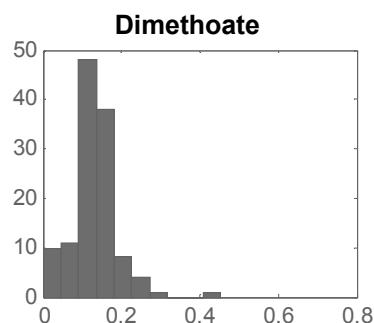
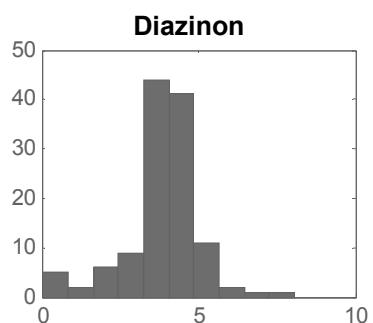
Bottle	Dimethoate mg/kg		Endosulfan mg/kg		Imazalil mg/kg		Imidacloprid mg/kg		Dithiocarabamates mg/kg	
1	0.155	0.156	0.385	0.386	0.194	0.195	0.242	0.240	0.800	0.801
2	0.154	0.155	0.374	0.374	0.192	0.192	0.239	0.238	0.756	0.755
3	0.153	0.154	0.385	0.384	0.193	0.193	0.239	0.239	0.790	0.795
4	0.157	0.157	0.384	0.385	0.190	0.191	0.240	0.240	0.792	0.793
5	0.152	0.153	0.373	0.375	0.187	0.189	0.239	0.239	0.750	0.752
6	0.155	0.155	0.380	0.380	0.190	0.191	0.235	0.235	0.805	0.806
7	0.155	0.154	0.385	0.384	0.185	0.185	0.237	0.237	0.810	0.809
8	0.155	0.154	0.383	0.383	0.186	0.185	0.238	0.238	0.798	0.799
9	0.156	0.155	0.384	0.384	0.170	0.172	0.235	0.235	0.765	0.768
10	0.155	0.155	0.385	0.384	0.189	0.188	0.238	0.238	0.798	0.798

Bottle	Oxydemeton-methyl mg/kg		Metalaxyl mg/kg		Procymidone mg/kg		Thiabendazole mg/kg	
1	0.199	0.200	0.050	0.050	0.419	0.420	0.326	0.327
2	0.198	0.199	0.045	0.046	0.415	0.415	0.320	0.322
3	0.198	0.199	0.045	0.045	0.416	0.416	0.330	0.331
4	0.198	0.199	0.049	0.049	0.416	0.416	0.335	0.332
5	0.199	0.200	0.048	0.049	0.418	0.418	0.330	0.329
6	0.200	0.200	0.049	0.049	0.418	0.418	0.340	0.339
7	0.202	0.201	0.050	0.050	0.417	0.418	0.335	0.334
8	0.200	0.200	0.047	0.048	0.417	0.418	0.335	0.335
9	0.196	0.197	0.048	0.048	0.419	0.419	0.328	0.329
10	0.199	0.199	0.050	0.050	0.423	0.422	0.328	0.328

APPENDIX 2. Histograms of residue data for each pesticide for all the laboratories.



APPENDIX 2. Histograms of residue data for each pesticide for all the laboratories.



APPENDIX 3. Results (mg/kg) and z-Score for FFP and Q_n

Lab Code	MPLR / Median (mg/kg)	Acrinathrin 0.05 / 0.284	z-Score (FFP 25%)	z-Score (Qn 34%)	Azoxystrobin 0.05 / 0.225	z-Score (FFP 25%)	z-Score (Qn 27%)	Bromopropylate 0.05 / 0.490	z-Score (FFP 25%)	z-Score (Qn 21%)	Chlorotalonil 0.01 / 1.626	z-Score (FFP 25%)	z-Score (Qn 33%)	Diazinon 0.02 / 3.955	z-Score (FFP 25%)	z-Score (Qn 21%)
001	0.382	1.4	1.0	0.237	0.2	0.2	0.584	0.8	0.9	1.830	0.5	0.4	4.340	0.4	0.5	
002/3	0.389	1.5	1.1	0.238	0.2	0.2	0.618	1.0	1.3	1.890	0.7	0.5	4.410	0.5	0.6	
004	NA			0.220	-0.1	-0.1	0.520	0.2	0.3	2.040	1.0	0.8	4.620	0.7	0.8	
005	NA			0.242	0.3	0.3	0.584	0.8	0.9	1.620	0.0	0.0	4.440	0.5	0.6	
006	>0.160			0.112	-2.0	-1.9	0.371	-1.0	-1.2	NA			>0.16			
007	0.325	0.6	0.4	0.229	0.1	0.1	0.439	-0.4	-0.5	1.350	-0.7	-0.5	3.520	-0.4	-0.5	
008	NA			0.222	-0.1	-0.1	0.324	-1.4	-1.6	0.950	-1.7	-1.3	3.165	-0.8	-1.0	
009	NA			0.194	-0.6	-0.5	0.511	0.2	0.2	1.390	-0.6	-0.4	4.250	0.3	0.4	
010	NA			NA			0.618	1.0	1.3	1.555	-0.2	-0.1	5.508	1.6	1.9	
011	NA			ND	-3.1	-2.9	ND	-3.6	-4.3	0.850	-1.9	-1.5	4.100	0.1	0.2	
012	0.278	-0.1	-0.1	ND	-3.1	-2.9	0.501	0.1	0.1	0.655	-2.4	-1.8	1.036	-3.0	-3.5	
013	0.240	-0.6	-0.5	0.280	1.0	0.9	0.460	-0.2	-0.3	2.120	1.2	0.9	3.460	-0.5	-0.6	
014	NA			0.090	-2.4	-2.3	0.550	0.5	0.6	0.420	-3.0	-2.3	4.200	0.2	0.3	
015	NA			ND	-3.1	-2.9	0.343	-1.2	-1.4	1.660	0.1	0.1	3.730	-0.2	-0.3	
016	NA			ND	-3.1	-2.9	0.498	0.1	0.1	0.877	-1.8	-1.4	3.800	-0.2	-0.2	
017	ND	-3.3	-2.5	0.173	-0.9	-0.9	0.363	-1.0	-1.2	0.649	-2.4	-1.8	3.500	-0.5	-0.6	
018	0.238	-0.6	-0.5	0.133	-1.6	-1.5	0.402	-0.7	-0.9	1.210	-1.0	-0.8	3.630	-0.3	-0.4	
019	0.200	-1.2	-0.9	0.230	0.1	0.1	0.460	-0.2	-0.3	2.200	1.4	1.1	4.400	0.5	0.5	
020	0.133	-2.1	-1.6	0.208	-0.3	-0.3	0.488	0.0	0.0	1.860	0.6	0.4	3.670	-0.3	-0.3	
021	0.291	0.1	0.1	0.259	0.6	0.6	0.591	0.8	1.0	1.810	0.5	0.3	4.780	0.8	1.0	
022	0.142	-2.0	-1.5	0.225	0.0	0.0	0.550	0.5	0.6	1.690	0.2	0.1	4.140	0.2	0.2	
023	0.393	1.5	1.1	0.266	0.7	0.7	0.613	1.0	1.2	1.740	0.3	0.2	3.460	-0.5	-0.6	
024	ND	-3.3	-2.5	0.290	1.2	1.1	0.497	0.1	0.1	ND	-4.0	-3.0	4.090	0.1	0.2	
025	0.275	-0.1	-0.1	0.228	0.1	0.1	0.480	-0.1	-0.1	1.160	-1.1	-0.9	4.100	0.1	0.2	
026	0.292	0.1	0.1	0.839	10.9	10.3	0.433	-0.5	-0.6	1.631	0.0	0.0	4.829	0.9	1.1	
027	0.293	0.1	0.1	0.251	0.5	0.4	0.626	1.1	1.3	0.951	-1.7	-1.3	5.050	1.1	1.3	
028	0.324	0.6	0.4	0.162	-1.1	-1.1	0.465	-0.2	-0.2	1.850	0.6	0.4	4.640	0.7	0.8	
029	0.016	-3.8	-2.8	0.265	0.7	0.7	0.655	1.3	1.6	1.930	0.7	0.6	5.240	1.3	1.6	
030	0.029	-3.6	-2.7	0.222	-0.1	-0.1	0.560	0.6	0.7	2.230	1.5	1.1	4.460	0.5	0.6	
031	0.113	-2.4	-1.8	0.413	3.3	3.1	0.517	0.2	0.3	2.080	1.1	0.9	4.610	0.7	0.8	
032	0.075	-2.9	-2.2	0.235	0.2	0.2	0.476	-0.1	-0.1	0.136	-3.7	-2.8	3.610	-0.3	-0.4	
033	0.362	1.1	0.8	0.344	2.1	2.0	0.532	0.3	0.4	0.774	-2.1	-1.6	4.820	0.9	1.1	
034	0.014	-3.8	-2.8	0.206	-0.3	-0.3	0.371	-1.0	-1.2	0.817	-2.0	-1.5	4.090	0.1	0.2	
035	ND	-3.3		0.245	0.4	0.3	0.451	-0.3	-0.4	1.010	-1.5	-1.2	3.650	-0.3	-0.4	
036	0.236	-0.7	-0.5	0.086	-2.5	-2.3	0.266	-1.8	-2.2	1.420	-0.5	-0.4	2.680	-1.3	-1.5	
037	0.435	2.1	1.6	NA			0.532	0.3	0.4	0.140	-3.7	-2.8	3.920	0.0	0.0	
038	0.287	0.0	0.0	0.271	0.8	0.8	0.576	0.7	0.8	0.456	-2.9	-2.2	5.090	1.1	1.4	
039	ND	-3.3	-2.5	0.182	-0.8	-0.7	0.252	-1.9	-2.3	1.870	0.6	0.5	8.050	4.1	5.0	
040	0.378	1.3	1.0	NA			0.504	0.1	0.1	1.407	-0.5	-0.4	0.043	-4.0	-4.8	
041	0.077	-2.9	-2.2	0.312	1.5	1.5	0.925	3.6	4.3	1.450	-0.4	-0.3	5.310	1.4	1.6	
042	0.220	-0.9	-0.7	0.260	0.6	0.6	0.490	0.0	0.0	0.950	-1.7	-1.3	5.900	2.0	2.4	
043	0.423	2.0	1.5	0.510	5.1	4.8	0.633	1.2	1.4	2.030	1.0	0.8	4.440	0.5	0.6	
044	1.220	13.2	9.8	0.198	-0.5	-0.5	0.602	0.9	1.1	2.620	2.4	1.9	3.570	-0.4	-0.5	
045	0.318	0.5	0.4	0.236	0.2	0.2	0.581	0.7	0.9	1.860	0.6	0.4	4.930	1.0	1.2	
046	0.294	0.1	0.1	0.255	0.5	0.5	0.501	0.1	0.1	2.030	1.0	0.8	4.210	0.3	0.3	
047	0.185	-1.4	-1.0	0.202	-0.4	-0.4	0.505	0.1	0.1	1.270	-0.9	-0.7	4.540	0.6	0.7	
048	NA			NA			0.445	-0.4	-0.4	1.390	-0.6	-0.4	3.950	0.0	0.0	
049	0.311	0.4	0.3	NA			0.493	0.0	0.0	1.760	0.3	0.3	3.570	-0.4	-0.5	
050	NA			NA			NA			NA			4.890	0.9	1.1	
051	NA			NA			0.533	0.4	0.4	1.260	-0.9	-0.7	3.890	-0.1	-0.1	
052	NA			NA			NA			NA			3.450	-0.5	-0.6	
053	NA			NA			NA			NA			NA			
054	NA			0.169	-1.0	-0.9	0.484	0.0	-0.1	1.650	0.1	0.0	4.030	0.1	0.1	
055	NA			NA			0.504	0.1	0.1	2.540	2.3	1.7	4.620	0.7	0.8	
056	NA			0.220	-0.1	-0.1	0.510	0.2	0.2	2.010	0.9	0.7	4.910	1.0	1.2	
057	0.253	-0.4	-0.3	0.240	0.3	0.3	0.366	-1.0	-1.2	1.500	-0.3	-0.2	4.200	0.2	0.3	
058	NA			ND	-3.1	-2.9	0.225	-2.2	-2.6	0.757	-2.1	-1.6	1.670	-2.3	-2.8	
059	NA			NA			0.280	-1.7	-2.1	0.740	-2.2	-1.7	2.100	-1.9	-2.3	
060	NA			NA			NA			NA			2.900	-1.1	-1.3	
061	0.322	0.5	0.4	0.255	0.5	0.5	0.506	0.1	0.2	1.650	0.1	0.0	4.480	0.5	0.6	
062	NA			NA			0.421	-0.6	-0.7	0.470	-2.8	-2.2	4.570	0.6	0.7	
063	NA			NA			0.361	-1.1	-1.3	0.579	-2.6	-2.0	1.770	-2.2	-2.7	
064	NA			0.248	0.4	0.4	0.483	-0.1	-0.1	1.590	-0.1	-0.1	2.560	-1.4	-1.7	
065	0.245	-0.5	-0.4	0.179	-0.8	-0.8	0.414	-0.6	-0.7	1.575	-0.1	-0.1	1.558	-2.4	-2.9	
066	NA			0.212	-0.2	-0.2	0.313	-1.4	-1.7	1.880	0.6	0.5	2.600	-1.4	-1.6	
067	NA			NA			0.269	-1.8	-2.2	1.340	-0.7	-0.5	2.390	-1.6	-1.9	
068	0.283	0.0	0.0	NA			0.546	0.5	0.5	1.750	0.3	0.2	4.320	0.4	0.4	
069	NA			0.205	-0.4	-0.3	0.480	-0.1	-0.1	1.450	-0.4	-0.3	3.150	-0.8	-1.0	
070	NA			0.251	0.5	0.4	0.477	-0.1	-0.1	1.260	-0.9	-0.7	4.020	0.1	0.1	
071	0.282	0.0	0.0	0.263	0.7	0.6	0.592	0.8	1.0	1.110	-1.3	-1.0	3.560	-0.4	-0.5	

APPENDIX 3. Results (mg/kg) and z-Score for FFP and Q_n

Lab Code	MRL/Median (mg/kg)	Acarathrin 0.05 / 0.284	z-Score (FFP 25%)	Azoxystrobin 0.05 / 0.225	z-Score (FFP 25%)	Bromopropylate 0.05 / 0.490	z-Score (Qn 27%)	Bromopropylate 0.05 / 0.490	z-Score (FFP 25%)	Chlorothalonil 0.01 / 1.626	z-Score (Qn 21%)	Diazinon 0.02 / 3.955	z-Score (FFP 25%)	Diazinon 0.02 / 3.955	z-Score (Qn 21%)	
072	NA			0.194	-0.6	-0.5	0.161	-2.7	-3.2	1.722	0.2	0.2	NA			
073	NA			0.286	1.1	1.0	0.440	-0.4	-0.5	NA			4.060	0.1	0.1	
074										No Reported Results						
075	NA			NA			0.650	1.3	1.6	1.250	-0.9	-0.7	4.480	0.5	0.6	
076	1.160	12.3	9.2	0.394	3.0	2.8	0.409	-0.7	-0.8	1.690	0.2	0.1	3.820	-0.1	-0.2	
077	NA			0.380	2.8	2.6	0.740	2.0	2.5	3.350	4.2	3.2	3.890	-0.1	-0.1	
078	0.189	-1.3	-1.0	0.170	-1.0	-0.9	0.430	-0.5	-0.6	1.700	0.2	0.1	5.000	1.1	1.3	
079	NA			0.295	1.2	1.2	0.552	0.5	0.6	2.990	3.4	2.6	3.080	-0.9	-1.1	
080										No Reported Results						
081	NA			NA			0.590	0.8	1.0	1.680	0.1	0.1	3.960	0.0	0.0	
082	0.257	-0.4	-0.3	ND	-3.1	-2.9	0.459	-0.3	-0.3	1.590	-0.1	-0.1	3.570	-0.4	-0.5	
083	NA			NA			0.700	1.7	2.1	2.940	3.2	2.5	4.360	0.4	0.5	
084	NA			NA			0.383	-0.9	-1.0	0.868	-1.9	-1.4	3.647	-0.3	-0.4	
085	0.258	-0.4	-0.3	0.245	0.4	0.3	0.460	-0.2	-0.3	1.930	0.7	0.6	4.320	0.4	0.4	
086	0.390	1.5	1.1	0.230	0.1	0.1	0.607	1.0	1.1	2.260	1.6	1.2	6.560	2.6	3.2	
087	0.321	0.5	0.4	0.246	0.4	0.4	0.621	1.1	1.3	2.110	1.2	0.9	5.130	1.2	1.4	
088	0.033	-3.5	-2.6	NA			0.047	-3.6	-4.3	0.130	-3.7	-2.8	D			
089	0.302	0.3	0.2	NA			NA			1.980	0.9	0.7	3.580	-0.4	-0.5	
090	0.190	-1.3	-1.0	0.200	-0.4	-0.4	0.380	-0.9	-1.1	0.910	-1.8	-1.3	2.000	-2.0	-2.4	
091	0.264	-0.3	-0.2	NA			0.482	-0.1	-0.1	2.290	1.6	1.2	3.610	-0.3	-0.4	
092	0.367	1.2	0.9	0.171	-1.0	-0.9	0.564	0.6	0.7	1.581	-0.1	-0.1	3.990	0.0	0.0	
093	NA			NA			0.243	-2.0	-2.4	ND	-4.0	-3.0	0.330	-3.7	-4.4	
094	0.388	1.5	1.1	NA			0.464	-0.2	-0.3	1.390	-0.6	-0.4	2.890	-1.1	-1.3	
095	0.295	0.2	0.1	0.207	-0.3	-0.3	0.439	-0.4	-0.5	2.420	2.0	1.5	4.290	0.3	0.4	
096	0.240	-0.6	-0.5	0.225	0.0	0.0	0.398	-0.8	-0.9	1.900	0.7	0.5	4.130	0.2	0.2	
097	0.285	0.0	0.0	0.233	0.1	0.1	0.534	0.4	0.4	1.200	-1.0	-0.8	3.790	-0.2	-0.2	
098	0.309	0.4	0.3	0.174	-0.9	-0.9	0.604	0.9	1.1	1.690	0.2	0.1	4.280	0.3	0.4	
099	NA			0.160	-1.2	-1.1	0.530	0.3	0.4	2.030	1.0	0.8	4.360	0.4	0.5	
100	0.281	0.0	0.0	0.184	-0.7	-0.7	0.514	0.2	0.2	1.590	-0.1	-0.1	3.800	-0.2	-0.2	
101	0.263	-0.3	-0.2	0.225	0.0	0.0	0.369	-1.0	-1.2	0.938	-1.7	-1.3	3.750	-0.2	-0.2	
102	NA			0.208	-0.3	-0.3	0.506	0.1	0.2	1.570	-0.1	-0.1	3.900	-0.1	-0.1	
103	NA			0.196	-0.5	-0.5	0.455	-0.3	-0.3	1.990	0.9	0.7	3.880	-0.1	-0.1	
104	NA			0.161	-1.1	-1.1	0.472	-0.1	-0.2	4.160	6.2	4.8	4.910	1.0	1.2	
105	NA			NA			NA			NA			0.117	-3.9	-4.7	
106	0.310	0.4	0.3	0.221	-0.1	-0.1	0.411	-0.6	-0.8	1.950	0.8	0.6	2.950	-1.0	-1.2	
107	ND	-3.3	-2.5	0.224	0.0	0.0	0.534	0.4	0.4	1.680	0.1	0.1	4.280	0.3	0.4	
108	NA			0.143	-1.5	-1.4	0.551	0.5	0.6	2.140	1.3	1.0	3.520	-0.4	-0.5	
109	NA			0.138	-1.5	-1.5	0.499	0.1	0.1	1.900	0.7	0.5	3.450	-0.5	-0.6	
110	0.642	5.0	3.8	0.198	-0.5	-0.5	0.525	0.3	0.3	1.410	-0.5	-0.4	3.860	-0.1	-0.1	
111	NA			0.272	0.8	0.8	0.456	-0.3	-0.3	1.988	0.9	0.7	3.798	-0.2	-0.2	
112	NA			0.154	-1.3	-1.2	0.574	0.7	0.8	1.927	0.7	0.6	6.050	2.1	2.5	
113	NA			0.368	2.5	2.4	0.595	0.9	1.0	1.780	0.4	0.3	3.670	-0.3	-0.3	
114	NA			0.213	-0.2	-0.2	0.374	-0.9	-1.1	1.810	0.5	0.3	2.370	-1.6	-1.9	
115	NA			ND	-3.1	-2.9	0.133	-2.9	-3.5	ND	-4.0	-3.0	0.524	-3.5	-4.2	
116	NA			0.210	-0.3	-0.3	0.490	0.0	0.0	NA			3.500	-0.5	-0.6	
117	NA			0.190	-0.6	-0.6	0.624	1.1	1.3	NA			4.570	0.6	0.7	
118	0.195	-1.3	-0.9	0.201	-0.4	-0.4	0.490	0.0	0.0	1.830	0.5	0.4	3.700	-0.3	-0.3	
119	0.256	-0.4	-0.3	0.167	-1.0	-1.0	0.417	-0.6	-0.7	1.580	-0.1	-0.1	3.480	-0.5	-0.6	
120	NA			NA			NA			0.750	-2.2	-1.6	3.510	-0.5	-0.5	
121	NA			0.130	-1.7	-1.6	0.413	-0.6	-0.8	1.710	0.2	0.2	3.990	0.0	0.0	
122	0.306	0.3	0.2	0.362	2.4	2.3	0.488	0.0	0.0	1.620	0.0	0.0	4.200	0.2	0.3	
123	0.397	1.6	1.2	0.286	1.1	1.0	0.542	0.4	0.5	1.610	0.0	0.0	3.260	-0.7	-0.8	
124	0.344	0.8	0.6	0.304	1.4	1.3	0.413	-0.6	-0.8	1.600	-0.1	0.0	4.350	0.4	0.5	
125	0.256	-0.4	-0.3	0.389	2.9	2.7	0.462	-0.2	-0.3	1.670	0.1	0.1	4.240	0.3	0.3	
126							No Reported Results									
127	0.208	-1.1	-0.8	0.108	-2.1	-2.0	0.112	-3.1	-3.7	0.811	-2.0	-1.5	0.677	-3.3	-4.0	
128	0.318	0.5	0.4	0.236	0.2	0.2	0.521	0.3	0.3	1.650	0.1	0.0	4.230	0.3	0.3	
129	0.200	-1.2	-0.9	0.345	2.1	2.0	0.465	-0.2	-0.2	1.070	-1.4	-1.0	3.930	0.0	0.0	
130	0.360	1.1	0.8	0.250	0.4	0.4	0.560	0.6	0.7	1.190	-1.1	-0.8	4.640	0.7	0.8	

APPENDIX 3. Results (mg/kg) and z-Score for FFP and Q_n

Lab Code	MRL / Median (mg/kg)	Dimethoate 0.02 / 0.132	z-Score (FFP 25%)	z-Score (Qn 9%)	Endosulfan 0.05 / 0.344	z-Score (FFP 25%)	z-Score (Qn 22%)	Imazalil 0.02 / 0.167	z-Score (FFP 25%)	z-Score (Qn 41%)	Imidacloprid 0.05 / 0.232	z-Score (FFP 25%)	Dithiocarbamates (CSZ) 0.05 / 0.810	z-Score (FFP 25%)	z-Score (Qn 33%)
001	0.142	0.3	0.3	0.400	0.7	0.7	0.194	0.6	0.4	0.214	-0.3	-0.4	1.180	1.8	1.4
002/3	0.131	0.0	0.0	0.470	1.5	1.7	0.215	1.1	0.7	NA			0.580	-1.1	-0.9
004	0.140	0.2	0.2	0.370	0.3	0.4	0.260	2.2	1.4	0.200	-0.5	-0.8	NA		
005	0.111	-0.6	-0.6	0.363	0.2	0.3	ND	-3.5	-2.1	NA			1.090	1.4	1.1
006	NA			0.264	-0.9	-1.1	0.079	-2.1	-1.3	0.159	-1.3	-1.8	NA		
007	0.120	-0.4	-0.3	0.355	0.1	0.2	0.249	2.0	1.2	0.284	0.9	1.3	NA		
008	0.053	-2.4	-2.1	0.299	-0.5	-0.6	0.390	5.3	3.2	NA			NA		
009	0.112	-0.6	-0.5	0.369	0.3	0.3	0.166	0.0	0.0	0.239	0.1	0.2	0.923	0.6	0.4
010	0.151	0.6	0.5	0.307	-0.4	-0.5	ND			NA			NA		
011	0.220	2.7	2.3	0.210	-1.6	-1.8	NA			NA			NA		
012	0.028	-3.2	-2.8	0.283	-0.7	-0.8	0.184	0.4	0.2	NA			1.040	1.1	0.9
013	0.055	-2.3	-2.0	0.250	-1.1	-1.2	0.155	-0.3	-0.2	0.230	0.0	0.0	0.700	-0.5	-0.4
014	0.050	-2.5	-2.2	0.450	1.2	1.4	0.240	1.7	1.1	NA			0.730	-0.4	-0.3
015	0.169	1.1	1.0	0.199	-1.7	-1.9	0.287	2.9	1.7	NA			0.780	-0.1	-0.1
016	0.113	-0.6	-0.5	0.305	-0.4	-0.5	ND	-3.5	-2.1	ND	-3.1	-4.5	NA		
017	0.084	-1.4	-1.3	0.300	-0.5	-0.6	NA			ND	-3.1	-4.5	0.918	0.5	0.4
018	0.164	1.0	0.8	0.270	-0.9	-1.0	0.131	-0.9	-0.5	0.244	0.2	0.3	0.781	-0.1	-0.1
019	0.130	-0.1	-0.1	0.310	-0.4	-0.4	0.160	-0.2	-0.1	0.230	0.0	0.0	0.860	0.2	0.2
020	0.121	-0.3	-0.3	0.347	0.0	0.0	0.132	-0.8	-0.5	0.244	0.2	0.3	0.997	0.9	0.7
021	0.167	1.1	0.9	0.368	0.3	0.3	0.135	-0.8	-0.5	0.243	0.2	0.3	0.622	-0.9	-0.7
022	0.118	-0.4	-0.4	0.396	0.6	0.7	0.294	3.0	1.8	0.217	-0.3	-0.4	NA		
023	0.139	0.2	0.2	0.354	0.1	0.1	0.195	0.7	0.4	0.285	0.9	1.3	1.040	1.1	0.9
024	ND	-3.4	-3.0	0.340	0.0	0.0	0.043	-3.0	-1.8	0.211	-0.4	-0.5	0.890	0.4	0.3
025	0.122	-0.3	-0.3	0.313	-0.4	-0.4	0.137	-0.7	-0.4	0.240	0.1	0.2	0.984	0.9	0.7
026	0.198	2.0	1.7	0.439	1.1	1.3	0.458	7.0	4.2	NA			NA		
027	0.123	-0.3	-0.2	0.278	-0.8	-0.9	0.237	1.7	1.0	0.265	0.6	0.8	0.648	-0.8	-0.6
028	0.164	1.0	0.8	0.305	-0.4	-0.5	0.115	-1.2	-0.8	0.260	0.5	0.7	0.840	0.1	0.1
029	0.132	0.0	0.0	0.476	1.5	1.8	0.209	1.0	0.6	0.241	0.2	0.2	NA		
030	0.144	0.4	0.3	0.430	1.0	1.1	0.216	1.2	0.7	0.227	-0.1	-0.1	0.950	0.7	0.5
031	0.151	0.6	0.5	0.413	0.8	0.9	0.163	-0.1	-0.1	0.241	0.2	0.2	0.558	-1.2	-1.0
032	0.117	-0.5	-0.4	0.289	-0.6	-0.7	0.115	-1.2	-0.8	0.220	-0.2	-0.3	0.805	0.0	0.0
033	0.198	2.0	1.7	0.370	0.3	0.4	ND	-3.5	-2.1	NA			0.914	0.5	0.4
034	0.114	-0.5	-0.5	0.315	-0.3	-0.4	0.150	-0.4	-0.2	0.240	0.1	0.2	NA		
035	0.120	-0.4	-0.3	0.340	0.0	0.0	0.165	0.0	0.0	0.230	0.0	0.0	NA		
036	0.121	-0.3	-0.3	0.294	-0.6	-0.7	ND	-3.5	-2.1	NA			NA		
037	0.151	0.6	0.5	0.433	1.0	1.2	0.264	2.3	1.4	NA			0.796	-0.1	-0.1
038	0.140	0.2	0.2	0.364	0.2	0.3	0.182	0.4	0.2	0.150	-1.4	-2.0	ND	-3.8	-2.9
039	0.115	-0.5	-0.5	0.345	0.0	0.0	0.121	-1.1	-0.7	0.207	-0.4	-0.6	0.471	-1.7	-1.3
040	0.099	-1.0	-0.9	0.246	-1.1	-1.3	NA			NA			NA		
041	0.110	-0.7	-0.6	0.339	-0.1	-0.1	0.163	-0.1	-0.1	0.393	2.8	4.0	0.875	0.3	0.2
042	0.150	0.5	0.5	0.350	0.1	0.1	0.120	-1.1	-0.7	NA			0.950	0.7	0.5
043	0.233	3.1	2.7	0.390	0.5	0.6	0.242	1.8	1.1	0.468	4.1	5.9	0.266	-2.7	-2.1
044	0.130	-0.1	-0.1	0.373	0.3	0.4	0.095	-1.7	-1.0	0.285	0.9	1.3	0.590	-1.1	-0.8
045	0.178	1.4	1.2	0.336	-0.1	-0.1	0.234	1.6	1.0	0.193	-0.7	-1.0	0.640	-0.8	-0.6
046	0.144	0.4	0.3	0.330	-0.2	-0.2	0.162	-0.1	-0.1	NA			0.900	0.4	0.3
047	0.152	0.6	0.5	0.339	-0.1	-0.1	0.143	-0.6	-0.3	NA			2.070	6.2	4.8
048	0.155	0.7	0.6	0.440	1.1	1.3	NA			NA			NA		
049	0.120	-0.4	-0.3	0.331	-0.1	-0.2	ND			NA			NA		
050	0.090	-1.3	-1.1	NA			NA			NA			NA		
051	0.114	-0.5	-0.5	0.298	-0.5	-0.6	NA			NA			NA		
052	0.101	-0.9	-0.8	NA			NA			NA			NA		
053	NA			NA			NA			NA			NA		
054	0.135	0.1	0.1	0.319	-0.3	-0.3	NA			NA			NA		
055	NA			0.305	-0.4	-0.5	NA			NA			NA		
056	0.130	-0.1	-0.1	0.340	0.0	0.0	NA			NA			NA		
057	0.191	1.8	1.6	0.310	-0.4	-0.4	0.149	-0.4	-0.3	0.238	0.1	0.2	0.601	-1.0	-0.8
058	0.115	-0.5	-0.5	0.854	5.9	6.8	ND	-3.5	-2.1	ND	-3.1	-4.5	NA		
059	ND	-3.4	-3.0	0.230	-1.3	-1.5	NA			NA			ND	-3.8	-2.9
060	NA			NA			ND	-3.5	-2.1	NA			0.780	-0.1	-0.1
061	0.241	3.3	2.9	0.386	0.5	0.6	0.225	1.4	0.8	NA			0.841	0.2	0.1
062	ND	-3.4	-3.0	0.589	2.9	3.2	ND			0.212	-0.3	-4.5	0.490	-1.6	-1.2
063	0.155	0.7	0.6	0.242	-1.2	-1.3	0.118	-1.2	-0.7	NA			NA		
064	0.240	3.3	2.9	0.381	0.4	0.5	0.154	-0.3	-0.2	NA			0.945	0.7	0.5
065	0.139	0.2	0.2	0.272	-0.8	-0.9	0.133	-0.8	-0.5	ND	-3.1	-4.5	0.660	-0.7	-0.6
066	0.112	-0.6	-0.5	0.187	-1.8	-2.1	0.100	-1.6	-1.0	NA			0.650	-0.8	-0.6

APPENDIX 3. Results (mg/kg) and z-Score for FFP and Q_n

Lab Code MPRL / Median (mg/kg)	Dimethoate 0.02 / 0.132	z-Score (FFP 25%)	z-Score (Qn 9%)	Endosulfan 0.05 / 0.344	z-Score (FFP 25%)	z-Score (Qn 22%)	Imazalil 0.02 / 0.167	z-Score (FFP 25%)	z-Score (Qn 41%)	Imidacloprid 0.05 / 0.232	z-Score (FFP 25%)	z-Score (Qn 17.3%)	Dithiocarbamates (CS ₂) 0.05 / 0.810	z-Score (FFP 25%)	z-Score (Qn 33%)
067	0.452	9.7	8.5	0.274	-0.8	-0.9	0.240	1.7	1.1	NA			0.852	0.2	0.2
068	0.164	1.0	0.8	0.373	0.3	0.4	0.210	1.0	0.6	NA			NA		
069	0.260	3.9	3.4	0.330	-0.2	-0.2	0.130	-0.9	-0.5	NA			1.200	1.9	1.5
070	0.168	1.1	1.0	0.406	0.7	0.8	0.284	2.8	1.7	NA			1.160	1.7	1.3
071	0.165	1.0	0.9	0.421	0.9	1.0	0.155	-0.3	-0.2	0.215	-0.3	-0.4	1.110	1.5	1.1
072	0.132	0.0	0.0	0.272	-0.8	-0.9	0.182	0.4	0.2	NA			1.020	1.0	0.8
073	0.175	1.3	1.1	0.451	1.3	1.4	0.257	2.2	1.3	NA			0.852	0.2	0.2
074	No Reported Results														
075	ND	-3.4	-3.0	0.160	-2.1	-2.4	0.230	1.5	0.9	NA			0.500	-1.5	-1.2
076	0.284	4.6	4.0	0.300	-0.5	-0.6	0.052	-2.8	-1.7	NA			1.260	2.2	1.7
077	0.089	-1.3	-1.1	3.580	37.7	42.8	ND	-3.5	-2.1	NA			NA		
078	0.133	0.0	0.0	0.294	-0.6	-0.7	0.238	1.7	1.0	0.200	-0.5	-0.8	0.741	-0.3	-0.3
079	0.124	-0.2	-0.2	0.357	0.2	0.2	0.091	-1.8	-1.1	NA			0.080	-3.6	-2.8
080	No Reported Results														
081	0.199	2.0	1.8	0.397	0.6	0.7	0.157	-0.2	-0.1	NA			NA		
082	0.180	1.5	1.3	0.418	0.9	1.0	0.221	1.3	0.8	NA			1.080	1.3	1.0
083	0.180	1.5	1.3	0.990	7.5	8.6	NA			NA			NA		
084	0.090	-1.3	-1.1	0.346	0.0	0.0	0.134	-0.8	-0.5	NA			NA		
085	0.089	-1.3	-1.1	0.292	-0.6	-0.7	0.194	0.6	0.4	0.280	0.8	1.2	0.623	-0.9	-0.7
086	0.122	-0.3	-0.3	0.468	1.4	1.6	0.150	-0.4	-0.2	0.209	-0.4	-0.6	1.030	1.1	0.8
087	0.119	-0.4	-0.3	0.404	0.7	0.8	0.208	1.0	0.6	NA			0.950	0.7	0.5
088	D		D				NA			0.237	0.1	0.1	NA		
089	0.141	0.3	0.2	0.353	0.1	0.1	NA			NA			0.426	-1.9	-1.5
090	ND	-3.4	-3.0	0.290	-0.6	-0.7	NA			0.180	-0.9	-1.3	ND	-3.8	-2.9
091	0.117	-0.5	-0.4	0.334	-0.1	-0.1	NA			NA			NA		
092	0.205	2.2	1.9	0.438	1.1	1.3	0.091	-1.8	-1.1	NA			0.590	-1.1	-0.8
093	0.204	2.2	1.9	0.258	-1.0	-1.1	NA			NA			0.130	-3.4	-2.6
094	0.088	-1.3	-1.2	0.268	-0.9	-1.0	0.084	-2.0	-1.2	NA			2.440	8.0	6.2
095	0.126	-0.2	-0.2	0.320	-0.3	-0.3	0.264	2.3	1.4	0.283	0.9	1.3	0.733	-0.4	-0.3
096	0.137	0.2	0.1	0.371	0.3	0.4	0.121	-1.1	-0.7	0.257	0.4	0.6	0.585	-1.1	-0.9
097	0.125	-0.2	-0.2	0.351	0.1	0.1	0.168	0.0	0.0	0.251	0.3	0.5	0.720	-0.4	-0.3
098	0.107	-0.8	-0.7	0.368	0.3	0.3	0.182	0.4	0.2	0.248	0.3	0.4	0.856	0.2	0.2
099	0.150	0.5	0.5	0.370	0.3	0.4	0.110	-1.4	-0.8	NA			0.220	-2.9	-2.2
100	0.119	-0.4	-0.3	0.393	0.6	0.7	0.153	-0.3	-0.2	0.196	-0.6	-0.9	1.120	1.5	1.2
101	0.125	-0.2	-0.2	0.547	2.4	2.7	ND	-3.5	-2.1	NA			NA		
102	0.147	0.5	0.4	0.338	-0.1	-0.1	ND	-3.5	-2.1	NA			NA		
103	0.107	-0.8	-0.7	0.325	-0.2	-0.2	0.127	-1.0	-0.6	NA			NA		
104	0.037	-2.9	-2.5	0.348	0.1	0.1	0.081	-2.1	-1.2	NA			NA		
105	ND	-3.4	-3.0	0.203	-1.6	-1.9	NA			NA			NA		
106	0.130	-0.1	-0.1	ND	-3.4	-3.9	0.161	-0.1	-0.1	NA			0.480	-1.6	-1.2
107	0.124	-0.2	-0.2	0.358	0.2	0.2	0.179	0.3	0.2	0.210	-0.4	-0.5	0.936	0.6	0.5
108	0.139	0.2	0.2	0.342	0.0	0.0	NA			NA			NA		
109	0.156	0.7	0.6	0.280	-0.7	-0.8	NA			NA			NA		
110	0.101	-0.9	-0.8	0.354	0.1	0.1	0.154	-0.3	-0.2	NA			0.710	-0.5	-0.4
111	0.160	0.8	0.7	0.357	0.2	0.2	NA			NA			0.780	-0.1	-0.1
112	0.139	0.2	0.2	0.402	0.7	0.8	NA			NA			0.746	-0.3	-0.2
113	0.178	1.4	1.2	0.369	0.3	0.3	0.308	3.4	2.0	NA			NA		
114	0.115	-0.5	-0.5	0.221	-1.4	-1.6	0.205	0.9	0.6	NA			NA		
115	ND	-3.4	-3.0	ND	-3.4	-3.9	ND	-3.5	-2.1	ND	-3.1	-4.5	NA		
116	0.080	-1.6	-1.4	0.280	-0.7	-0.8	0.220	1.3	0.8	NA			NA		
117	0.086	-1.4	-1.2	NA			ND	-3.5	-2.1	NA			0.893	0.4	0.3
118	0.109	-0.7	-0.6	0.294	-0.6	-0.7	0.131	-0.9	-0.5	NA			NA		
119	0.094	-1.2	-1.0	0.249	-1.1	-1.3	0.138	-0.7	-0.4	0.220	-0.2	-0.3	0.399	-2.0	-1.6
120	ND	-3.4	-3.0	0.120	-2.6	-3.0	NA			NA			NA		
121	0.107	-0.8	-0.7	0.400	0.7	0.7	0.147	-0.5	-0.3	0.248	0.3	0.4	0.242	-2.8	-2.2
122	0.154	0.7	0.6	0.398	0.6	0.7	0.245	1.9	1.1	0.312	1.4	2.0	0.864	0.3	0.2
123	0.174	1.3	1.1	0.455	1.3	1.5	0.305	3.3	2.0	0.186	-0.8	-1.1	0.573	-1.2	-0.9
124	0.140	0.2	0.2	0.309	-0.4	-0.5	0.325	3.8	2.3	0.232	0.0	0.0	1.000	0.9	0.7
125	0.135	0.1	0.1	0.325	-0.2	-0.2	0.197	0.7	0.4	0.177	-0.9	-1.4	0.820	0.0	0.0
126	No Reported Results														
127	0.096	-1.1	-1.0	0.311	-0.4	-0.4	0.273	2.5	1.5	0.231	0.0	0.0	0.495	-1.6	-1.2
128	0.158	0.8	0.7	0.465	1.4	1.6	0.350	4.4	2.7	0.306	1.3	1.9	0.815	0.0	0.0
129	0.130	-0.1	-0.1	0.500	1.8	2.1	0.271	2.5	1.5	0.140	-1.6	-2.3	0.900	0.4	0.3
130	0.190	1.8	1.5	0.470	1.5	1.7	0.280	2.7	1.6	0.200	-0.5	-0.8	0.700	-0.5	-0.4

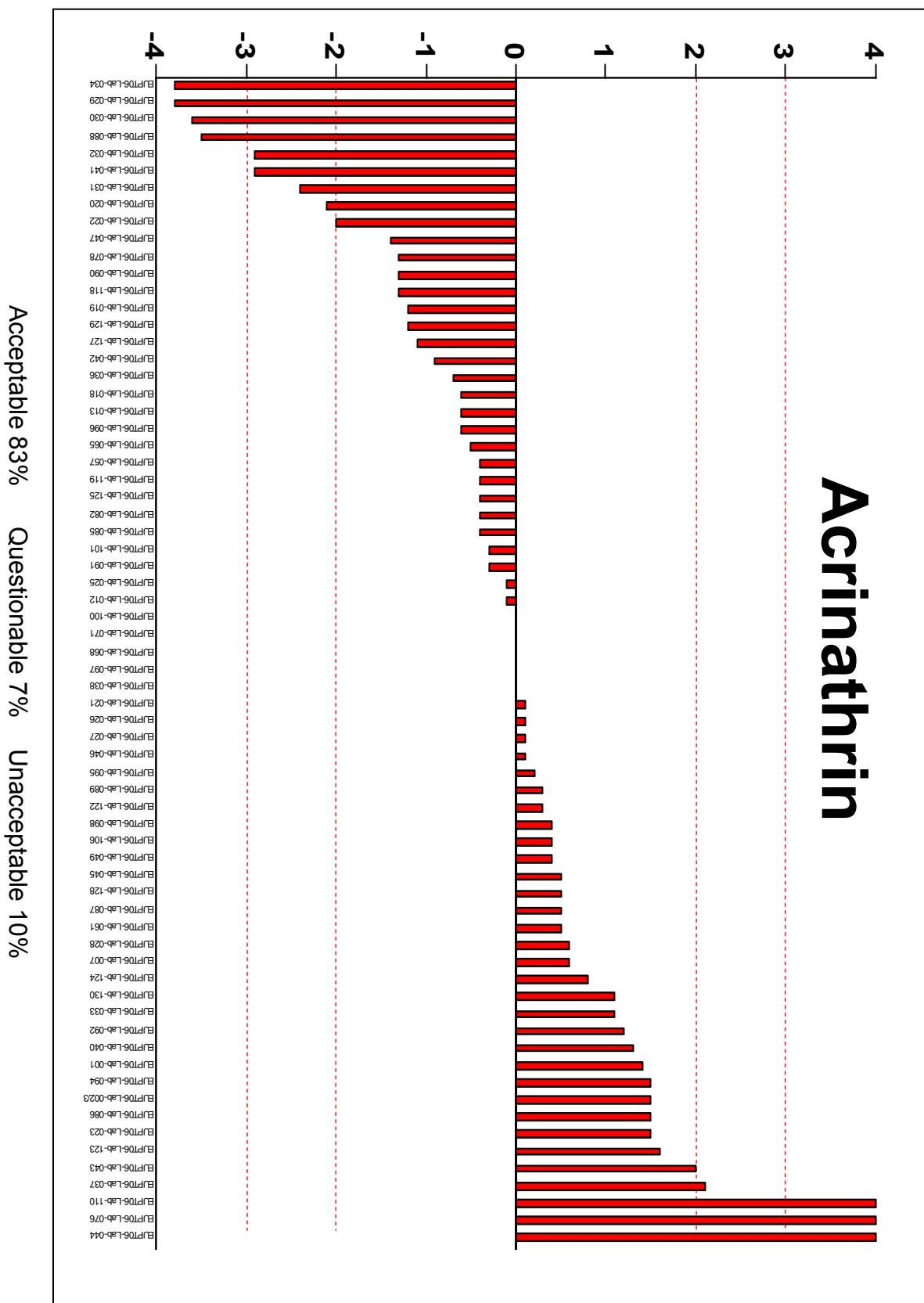
APPENDIX 3. Results (mg/kg) and z-Score for FFP and Q_n

Lab Code MPRL Median (mg/kg)	Metaxyl 0.05	Oxydemeton-methyl 0.02 / 0.199	z-Score (FFP 25%)	z-Score (Qn 35%)	Procymidone 0.02 / 0.412	z-Score (FFP 25%)	z-Score (Qn 21%)	Thiabendazole 0.05 / 0.314	z-Score (FFP 25%)	z-Score (Qn 27%)
001	ND	0.202	0.1	0.1	0.444	0.3	0.4	0.404	1.1	1.1
002/3	ND	NA			0.473	0.6	0.7	0.430	1.5	1.4
004	D	0.240	0.8	0.6	0.310	-1.0	-1.2	0.360	0.6	0.5
005	ND	NA			0.467	0.5	0.6	0.313	0.0	0.0
006	<0.05	NA			NA			0.215	-1.3	-1.2
007	0.034	0.137	-1.2	-0.9	0.416	0.0	0.0	0.312	0.0	0.0
008	ND	ND	-3.6	-2.6	0.333	-0.8	-0.9	0.258	-0.7	-0.7
009	ND	0.194	-0.1	-0.1	0.521	1.1	1.3	0.249	-0.8	-0.8
010	ND	NA			0.050	-3.5	-4.2	ND	-3.4	-3.1
011	NA	NA			0.370	-0.4	-0.5	0.292	-0.3	-0.3
012	0.016	NA			0.423	0.1	0.1	0.345	0.4	0.4
013	0.055	NA			0.420	0.1	0.1	0.290	-0.3	-0.3
014	0.035	NA			0.420	0.1	0.1	0.240	-0.9	-0.9
015	ND	NA			0.309	-1.0	-1.2	0.338	0.3	0.3
016	ND	ND	-3.6	-2.6	0.303	-1.1	-1.3	0.285	-0.4	-0.3
017	0.027	ND	-3.6	-2.6	0.327	-0.8	-1.0	0.190	-1.6	-1.5
018	0.044	0.205	0.1	0.1	0.313	-1.0	-1.1	0.217	-1.2	-1.1
019	0.028	0.240	0.8	0.6	0.410	0.0	0.0	0.330	0.2	0.2
020	0.024	0.205	0.1	0.1	0.393	-0.2	-0.2	0.302	-0.2	-0.1
021	0.045	0.233	0.7	0.5	0.448	0.3	0.4	0.313	0.0	0.0
022	0.033	0.191	-0.2	-0.1	0.436	0.2	0.3	0.370	0.7	0.7
023	0.034	0.274	1.5	1.1	0.495	0.8	1.0	0.377	0.8	0.7
024	0.032	0.223	0.5	0.4	0.412	0.0	0.0	0.335	0.3	0.2
025	0.030	0.198	0.0	0.0	0.423	0.1	0.1	0.355	0.5	0.5
026	0.050	NA			0.518	1.0	1.2	NA		
027	0.030	NA			0.423	0.1	0.1	0.471	2.0	1.9
028	0.031	0.190	-0.2	-0.1	0.352	-0.6	-0.7	0.350	0.5	0.4
029	0.037	0.213	0.3	0.2	0.482	0.7	0.8	0.348	0.4	0.4
030	ND	0.299	2.0	1.5	0.516	1.0	1.2	0.430	1.5	1.4
031	0.050	0.088	-2.2	-1.6	0.445	0.3	0.4	0.376	0.8	0.7
032	0.025	0.199	0.0	0.0	0.412	0.0	0.0	0.164	-1.9	-1.8
033	0.048	NA			0.445	0.3	0.4	0.235	-1.0	-0.9
034	0.025	NA			0.395	-0.2	-0.2	0.286	-0.4	-0.3
035	0.040	0.100	-2.0	-1.4	0.406	-0.1	-0.1	0.490	2.2	2.1
036	0.151	NA			0.311	-1.0	-1.2	NA		
037	ND	NA			0.437	0.2	0.3	0.390	1.0	0.9
038	0.038	0.100	-2.0	-1.4	0.474	0.6	0.7	0.193	-1.5	-1.4
039	ND	0.231	0.7	0.5	0.395	-0.2	-0.2	0.297	-0.2	-0.2
040	ND	ND	-3.6	-2.6	0.525	1.1	1.3	NA		
041	0.037	0.423	4.5	3.3	0.507	0.9	1.1	0.467	1.9	1.8
042	0.030	0.200	0.0	0.0	0.490	0.8	0.9	0.420	1.4	1.3
043	0.092	0.597	8.0	5.8	0.418	0.1	0.1	1.503	15.1	14.1
044	0.034	0.067	-2.6	-1.9	0.398	-0.1	-0.2	0.286	-0.4	-0.3
045	0.040	0.050	-3.0	-2.2	0.462	0.5	0.6	0.357	0.5	0.5
046	ND	0.424	4.5	3.3	0.424	0.1	0.1	0.285	-0.4	-0.3
047	0.053	0.302	2.1	1.5	0.448	0.3	0.4	0.249	-0.8	-0.8
048	NA	ND	-3.6	-2.6	0.507	0.9	1.1	NA		
049	ND	NA			0.408	0.0	0.0	NA		
050	NA	NA			NA			ND	-3.4	-3.1
051	NA	NA			0.362	-0.5	-0.6	NA		
052	NA	NA			NA			NA		
053	NA	NA			NA			NA		
054	ND	NA			0.319	-0.9	-1.1	ND	-3.4	-3.1
055	ND	NA			0.429	0.2	0.2	NA		
056	0.040	NA			0.400	-0.1	-0.1	0.390	1.0	0.9
057	0.051	NA			0.352	-0.6	-0.7	0.258	-0.7	-0.7
058	ND	NA			0.480	0.7	0.8	ND	-3.4	-3.1
059	NA	NA			0.220	-1.9	-2.2	NA		
060	0.060	NA			0.320	-0.9	-1.1	NA		
061	0.058	ND	-3.6	-2.6	0.415	0.0	0.0	0.299	-0.2	-0.2
062	ND	NA			0.760	3.4	4.0	0.218	-1.2	-1.1
063	ND	NA			0.269	-1.4	-1.7	0.338	0.3	0.3
064	0.047	NA			0.451	0.4	0.5	0.218	-1.2	-1.1
065	0.038	ND	-3.6	-2.6	0.342	-0.7	-0.8	0.319	0.1	0.1
066	ND	NA			0.359	-0.5	-0.6	0.140	-2.2	-2.1
067	0.110	NA			0.270	-1.4	-1.6	0.275	-0.5	-0.5
068	0.051	NA			0.452	0.4	0.5	0.368	0.7	0.6
069	ND	NA			0.375	-0.4	-0.4	NA		
070	0.101	NA			0.416	0.0	0.0	0.205	-1.4	-1.3
071	0.050	NA			0.556	1.4	1.7	0.190	-1.6	-1.5
072	ND	NA			0.470	0.6	0.7	0.553	3.0	2.8

APPENDIX 3. Results (mg/kg) and z-Score for FFP and Q_n

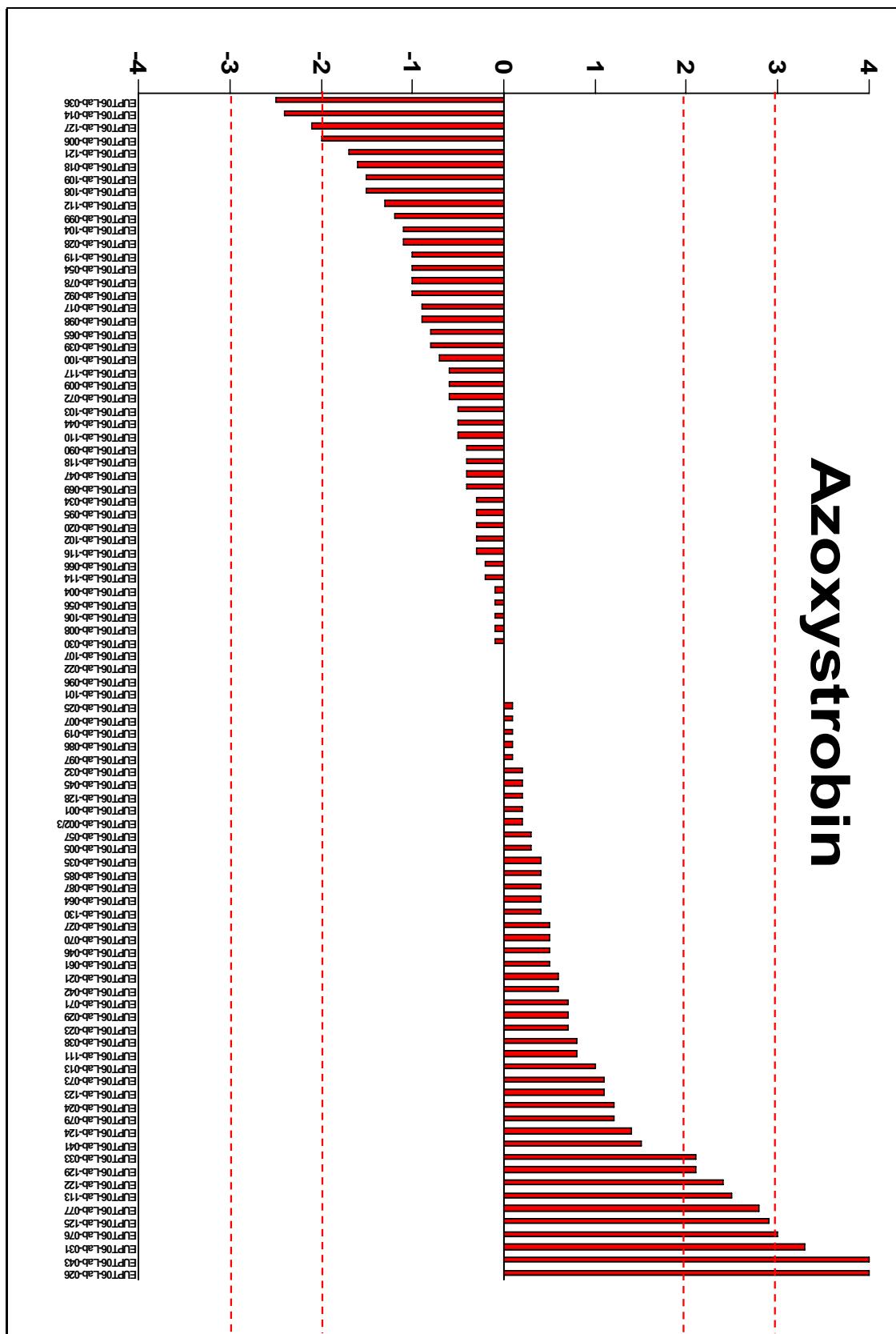
Lab Code MPRL Median (mg/kg)	Metalaxyl 0.05	Oxydemeton-methyl 0.02 / 0.199	z-Score (FFP 25%)	z-Score (Qn 35%)	Procymidone 0.02 / 0.412	z-Score (FFP 25%)	z-Score (Qn 21%)	Thiabendazole 0.05 / 0.314	z-Score (FFP 25%)	z-Score (Qn 27%)
073	0.087	NA			0.372	-0.4	-0.5	NA		
No Reported Results										
074	ND	NA			0.710	2.9	3.4	0.400	1.1	1.0
075	0.051	ND	-3.6	-2.6	0.341	-0.7	-0.8	0.980	8.5	7.9
076	0.060	ND	-3.6	-2.6	0.420	0.1	0.1	0.360	0.6	0.5
077	ND	0.112	-1.7	-1.3	0.338	-0.7	-0.9	0.264	-0.6	-0.6
079	ND	NA			0.339	-0.7	-0.8	NA		
No Reported Results										
080	0.041	NA			0.543	1.3	1.5	NA		
081	0.214	NA			0.266	-1.4	-1.7	NA		
082	0.023	NA			0.400	-0.1	-0.1	NA		
083	0.030	NA			0.286	-1.2	-1.5	NA		
084	0.031	0.083	-2.3	-1.7	0.315	-0.9	-1.1	0.291	-0.3	-0.3
086	0.038	0.073	-2.5	-1.8	0.574	1.6	1.9	0.365	0.6	0.6
087	0.036	0.094	-2.1	-1.5	0.539	1.2	1.5	0.271	-0.5	-0.5
088	D	NA			0.035	-3.7	-4.4	NA		
089	NA	NA			0.553	1.4	1.6	NA		
090	NA	ND	-3.6	-2.6	0.350	-0.6	-0.7	0.230	-1.1	-1.0
091	ND	NA			0.622	2.0	2.4	NA		
092	0.050	NA			0.466	0.5	0.6	NA		
093	NA	NA			0.174	-2.3	-2.8	0.315	0.0	0.0
094	NA	NA			0.244	-1.6	-1.9	NA		
095	ND	0.211	0.3	0.2	0.369	-0.4	-0.5	0.404	1.1	1.1
096	0.031	0.159	-0.8	-0.6	0.332	-0.8	-0.9	0.341	0.3	0.3
097	0.031	0.190	-0.2	-0.1	0.416	0.0	0.0	0.300	-0.2	-0.2
098	0.031	0.249	1.0	0.7	0.470	0.6	0.7	0.356	0.5	0.5
099	0.040	0.160	-0.8	-0.6	0.390	-0.2	-0.3	0.340	0.3	0.3
100	ND	0.165	-0.7	-0.5	0.392	-0.2	-0.2	0.277	-0.5	-0.4
101	ND	NA			0.405	-0.1	-0.1	ND	-3.4	-3.1
102	0.031	NA			0.396	-0.2	-0.2	NA		
103	0.029	ND	-3.6	-2.6	0.456	0.4	0.5	0.275	-0.5	-0.5
104	0.026	NA			0.474	0.6	0.7	0.029	-3.6	-3.4
105	NA	NA			NA			NA		
106	ND	NA			0.250	-1.6	-1.9	0.449	1.7	1.6
107	ND	NA			0.433	0.2	0.2	0.292	-0.3	-0.3
108	ND	NA			0.351	-0.6	-0.7	NA		
109	ND	NA			0.341	-0.7	-0.8	NA		
110	ND	NA			0.421	0.1	0.1	NA		
111	ND	NA			0.285	-1.2	-1.5	NA		
112	ND	NA			0.526	1.1	1.3	NA		
113	0.038	NA			0.392	-0.2	-0.2	0.300	-0.2	-0.2
114	ND	NA			0.247	-1.6	-1.9	0.263	-0.6	-0.6
115	ND	NA			D			ND	-3.4	-3.1
116	0.030	NA			0.410	0.0	0.0	0.230	-1.1	-1.0
117	0.027	NA			0.535	1.2	1.4	ND	-3.4	-3.1
118	0.025	NA			0.403	-0.1	-0.1	0.351	0.5	0.4
119	0.028	NA			0.347	-0.6	-0.8	0.379	0.8	0.8
120	NA	NA			NA			NA		
121	0.031	NA			0.358	-0.5	-0.6	0.398	1.1	1.0
122	ND	NA			0.414	0.0	0.0	0.335	0.3	0.2
123	ND	0.205	0.1	0.1	0.471	0.6	0.7	0.422	1.4	1.3
124	0.050	0.188	-0.2	-0.2	0.406	-0.1	-0.1	0.310	-0.1	0.0
125	0.073	0.196	-0.1	0.0	0.431	0.2	0.2	0.349	0.4	0.4
No Reported Results										
126	0.050	NA			0.483	0.7	0.8	0.298	-0.2	-0.2
127	0.067	0.202	0.1	0.1	0.471	0.6	0.7	0.355	0.5	0.5
128	0.050	NA			0.320	-0.9	-1.1	0.300	-0.2	-0.2
129	0.050	0.120	-1.6	-1.1	0.420	0.1	0.1	0.390	1.0	0.9

APPENDIX 4.- Graphical representation of z-Scores for FFP (25%).

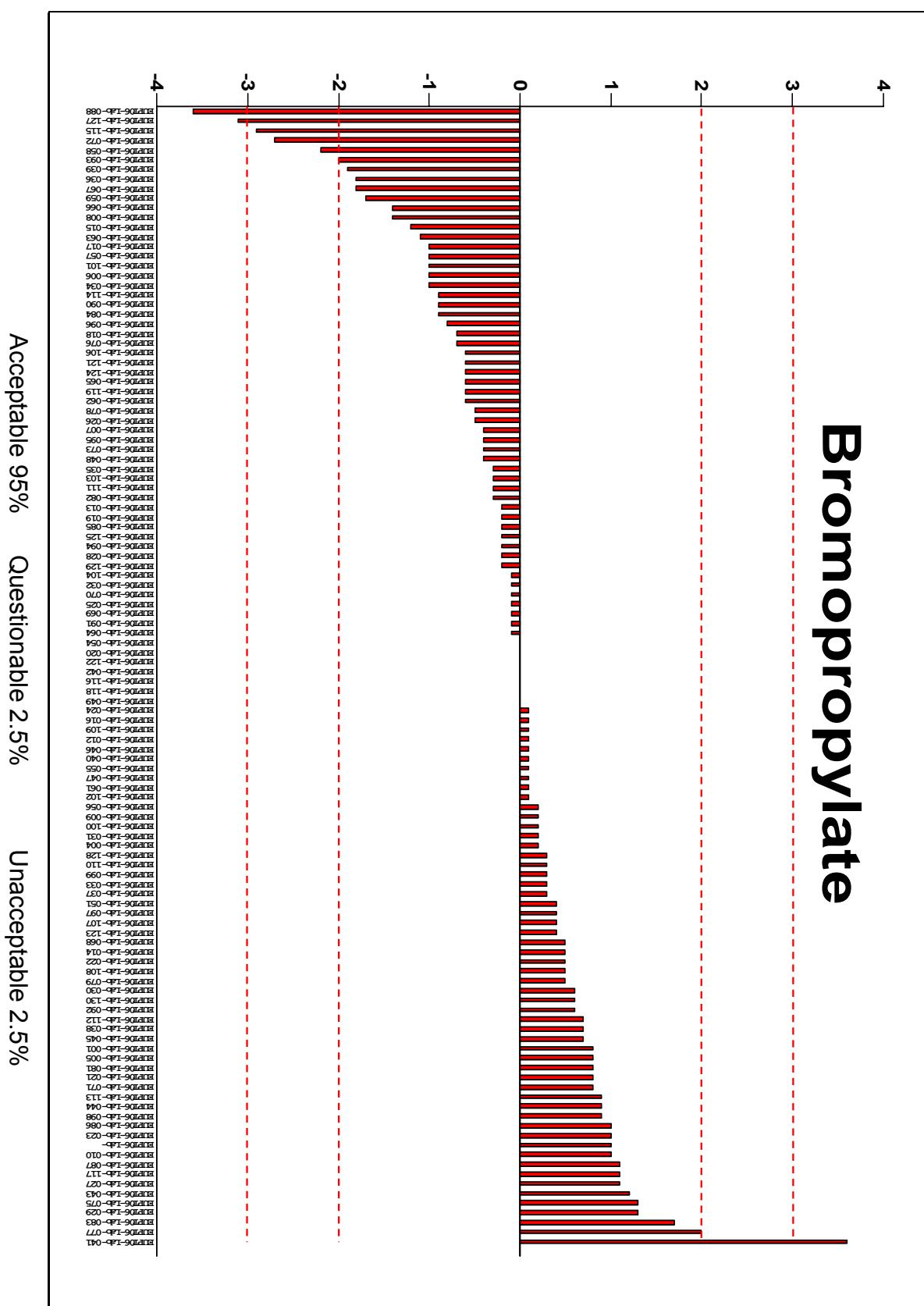


APPENDIX 4.- Graphical representation of z-Scores for FFP (25%).

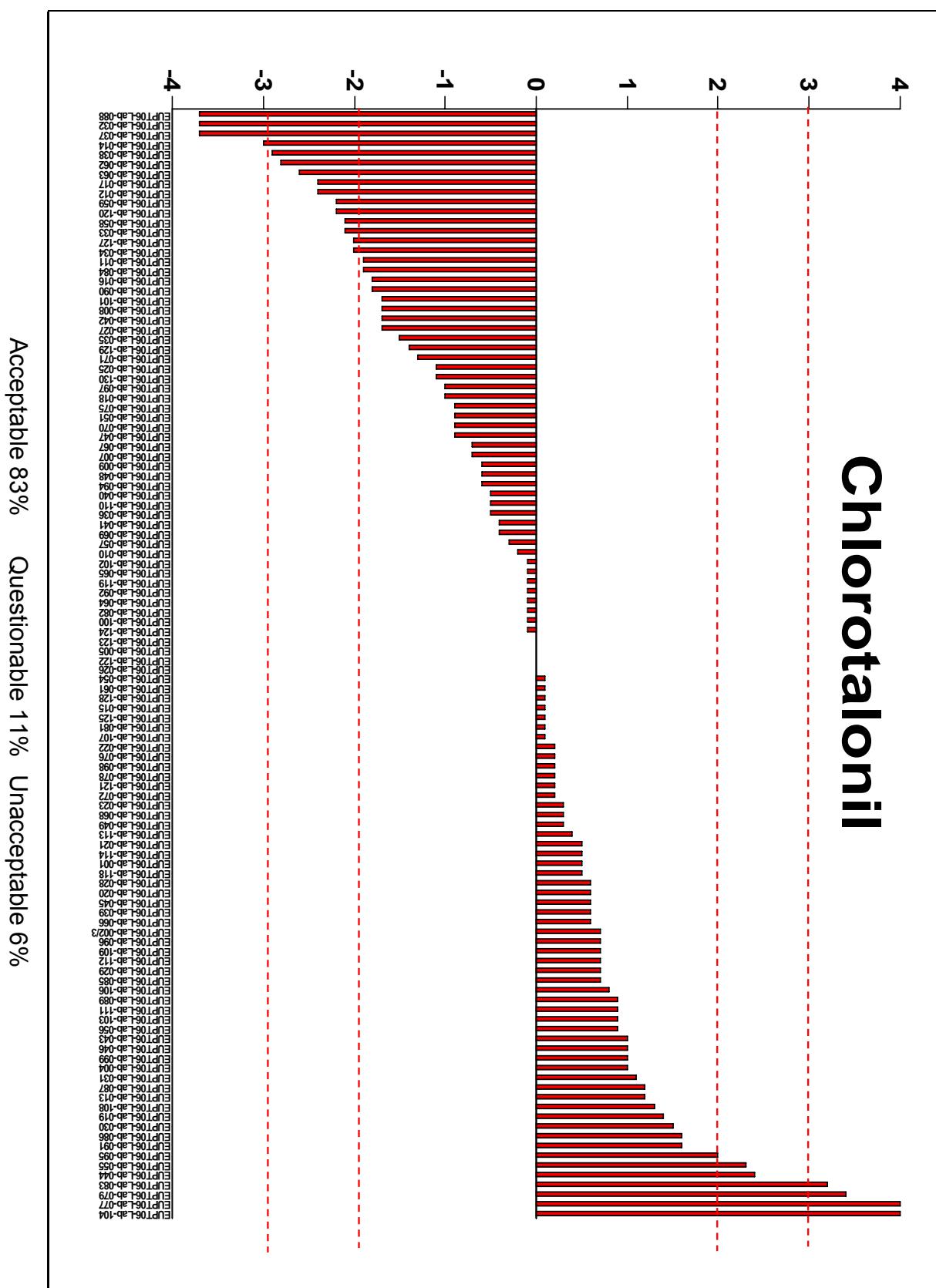
Acceptable 86% Questionable 11% Unacceptable 3%



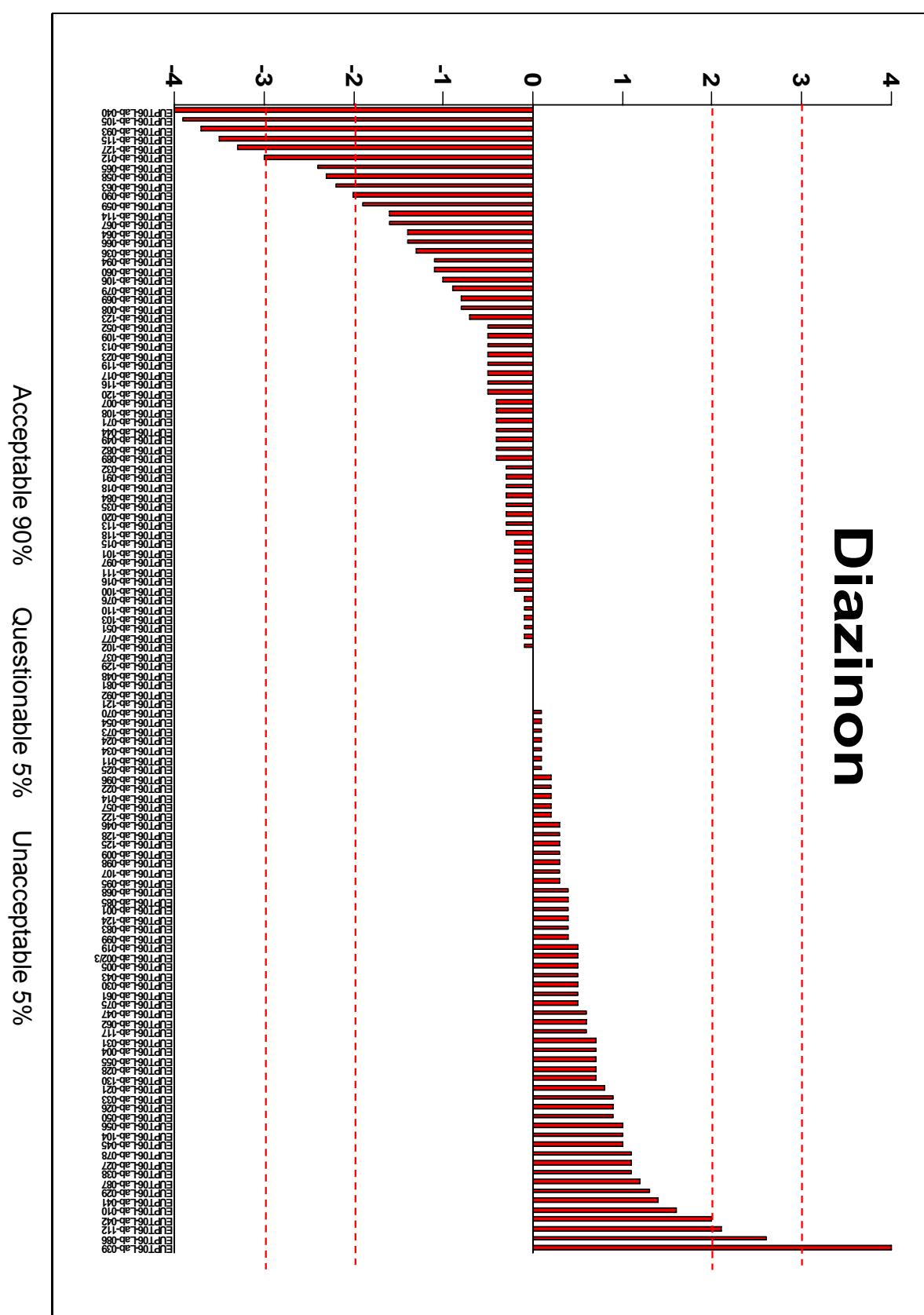
APPENDIX 4.- Graphical representation of z-Scores for FFP (25%).



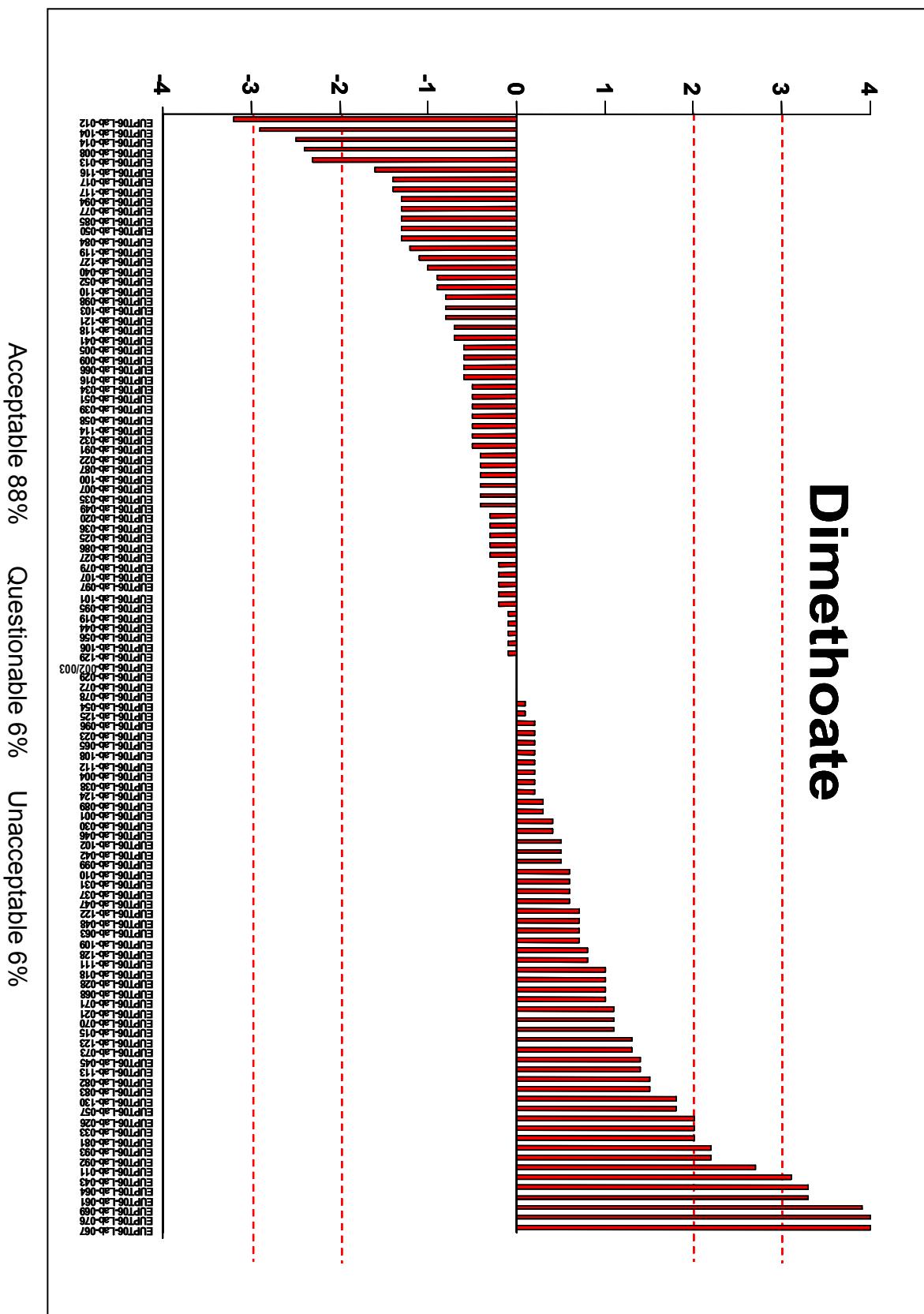
APPENDIX 4.- Graphical representation of z-Scores for FFP (25%).



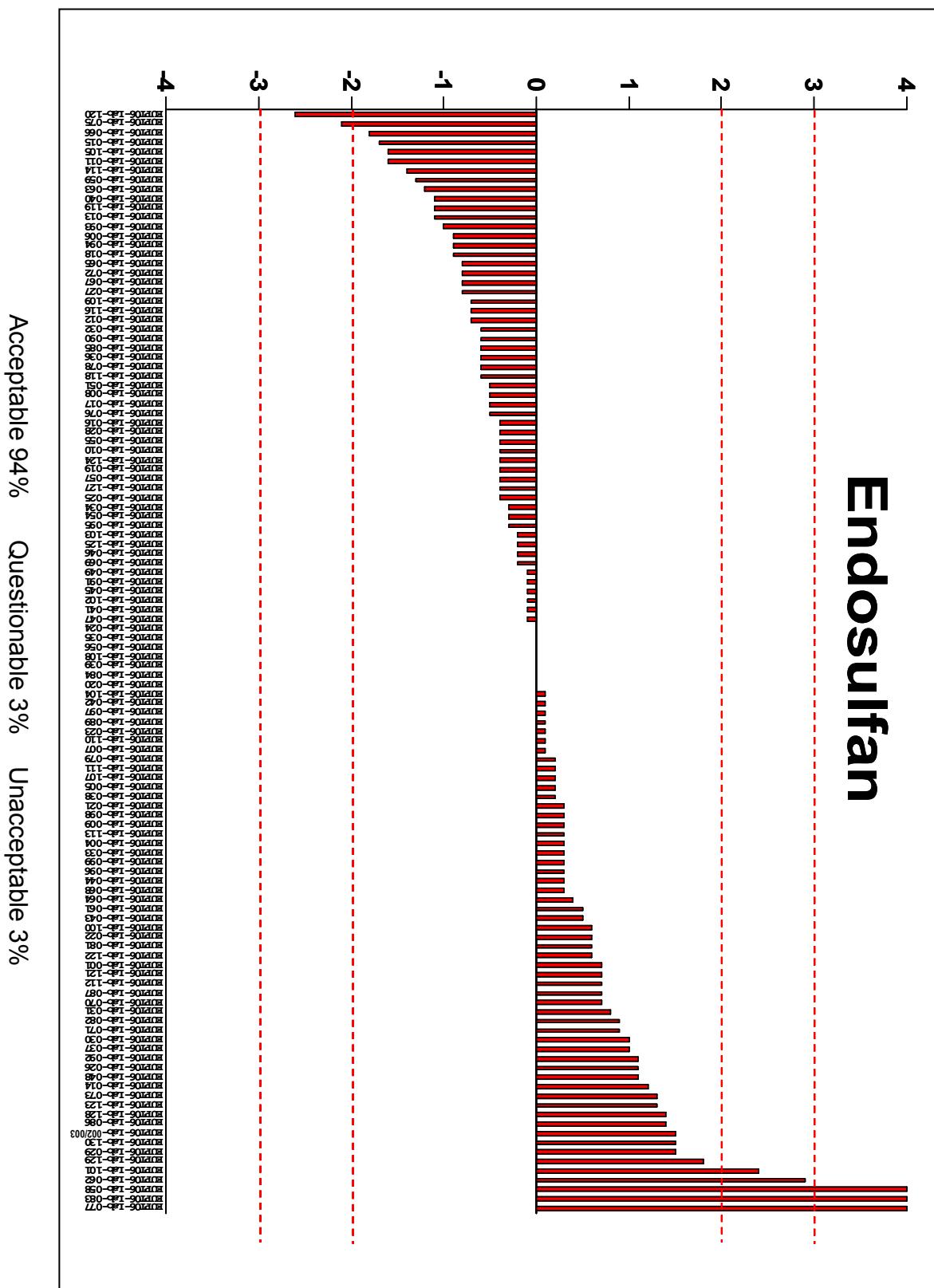
APPENDIX 4.- Graphical representation of z-Scores for FFP (25%).



APPENDIX 4.- Graphical representation of z-Scores for FFP (25%).

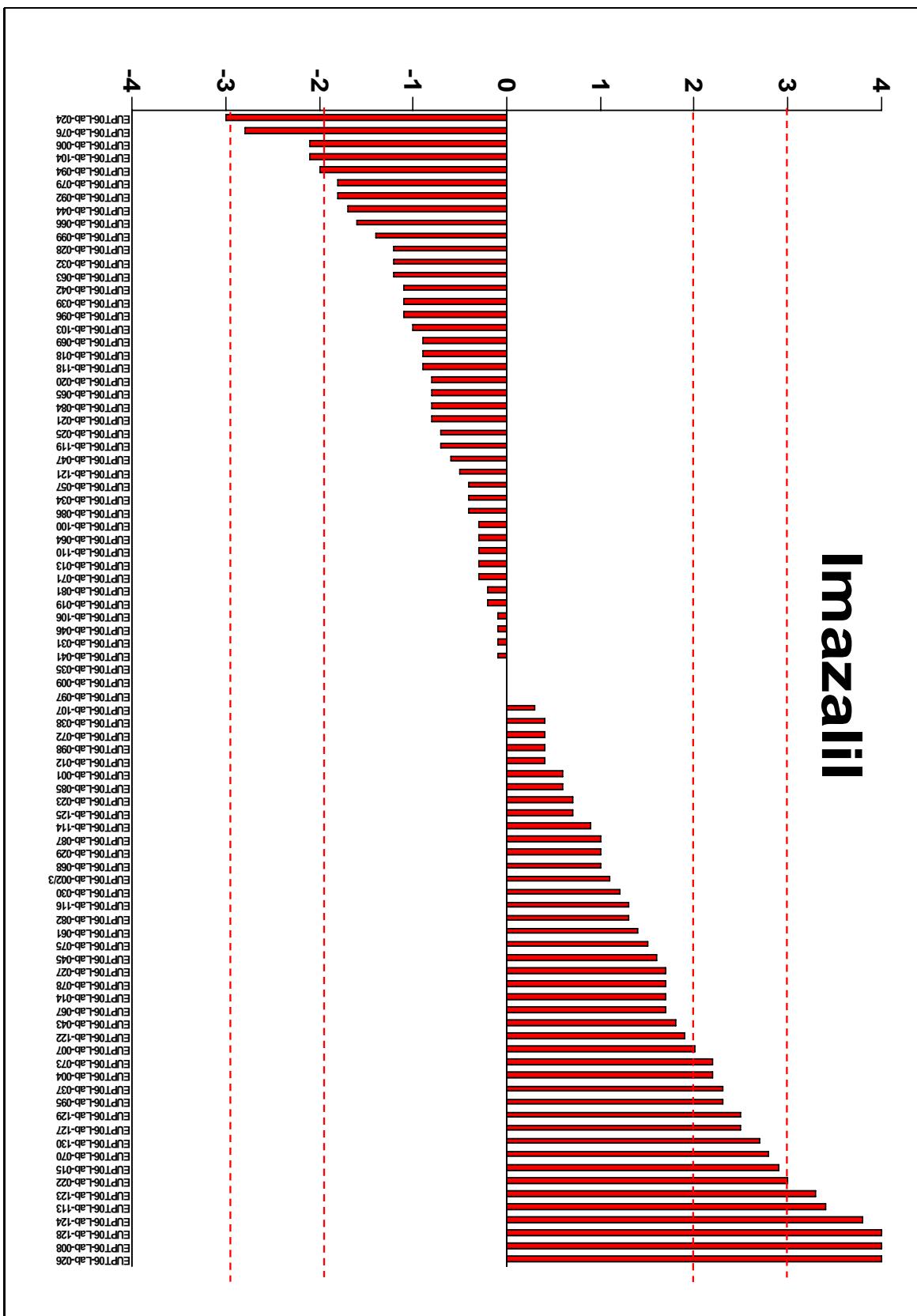


APPENDIX 4.- Graphical representation of z-Scores for FFP (25%).

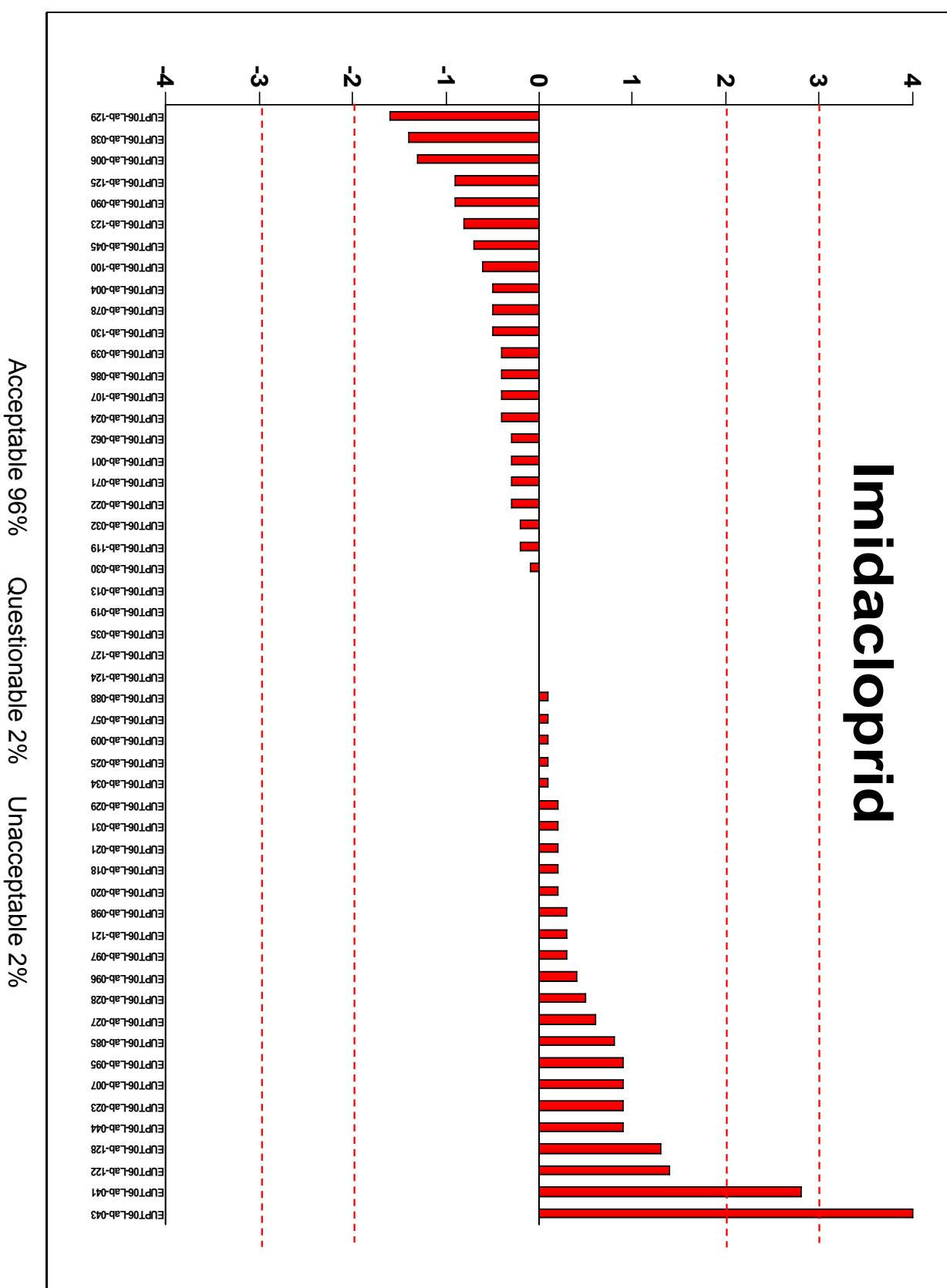


APPENDIX 4.- Graphical representation of z-Scores for FFP (25%).

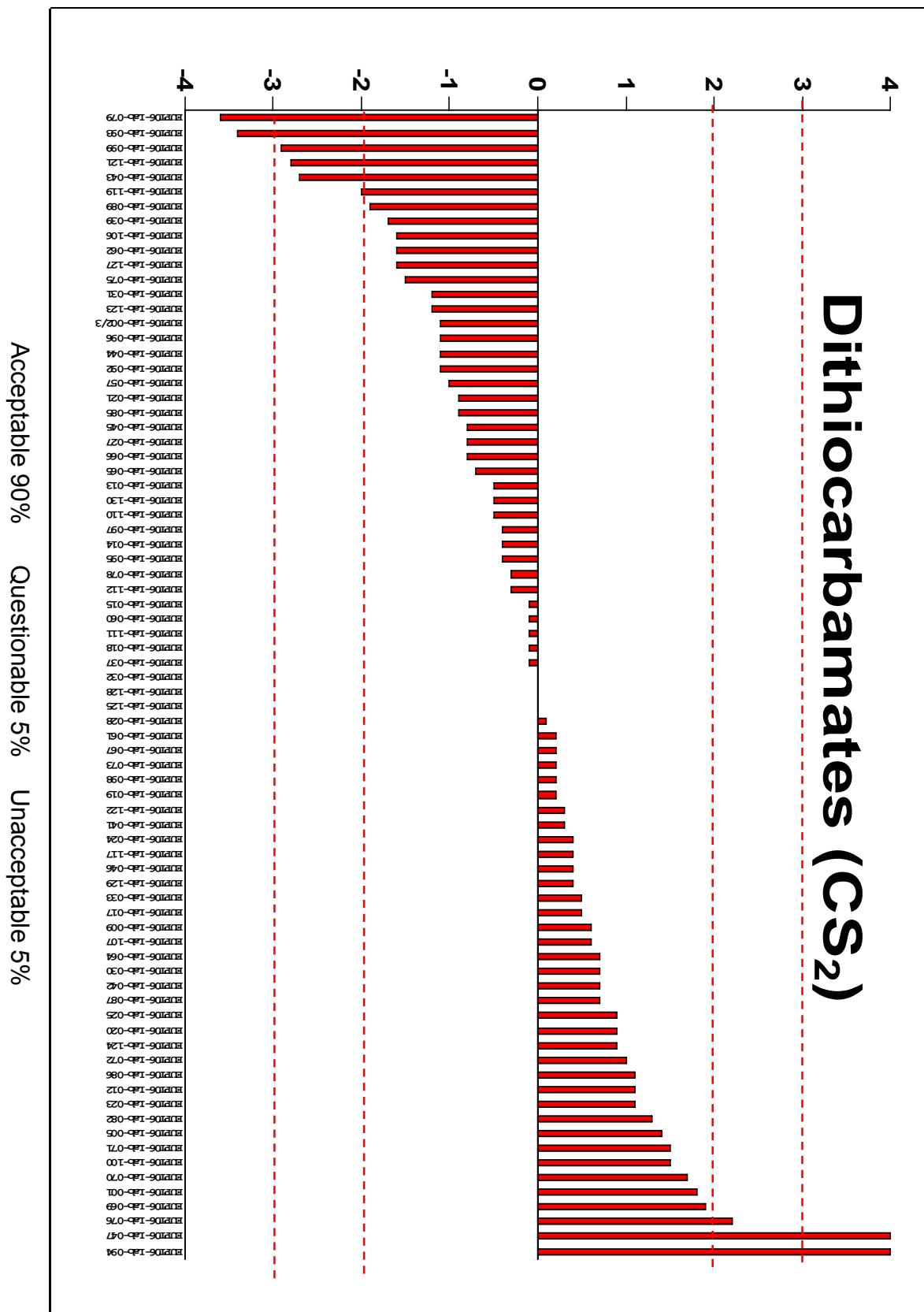
Acceptable 77% Questionable 16% Unacceptable 7%



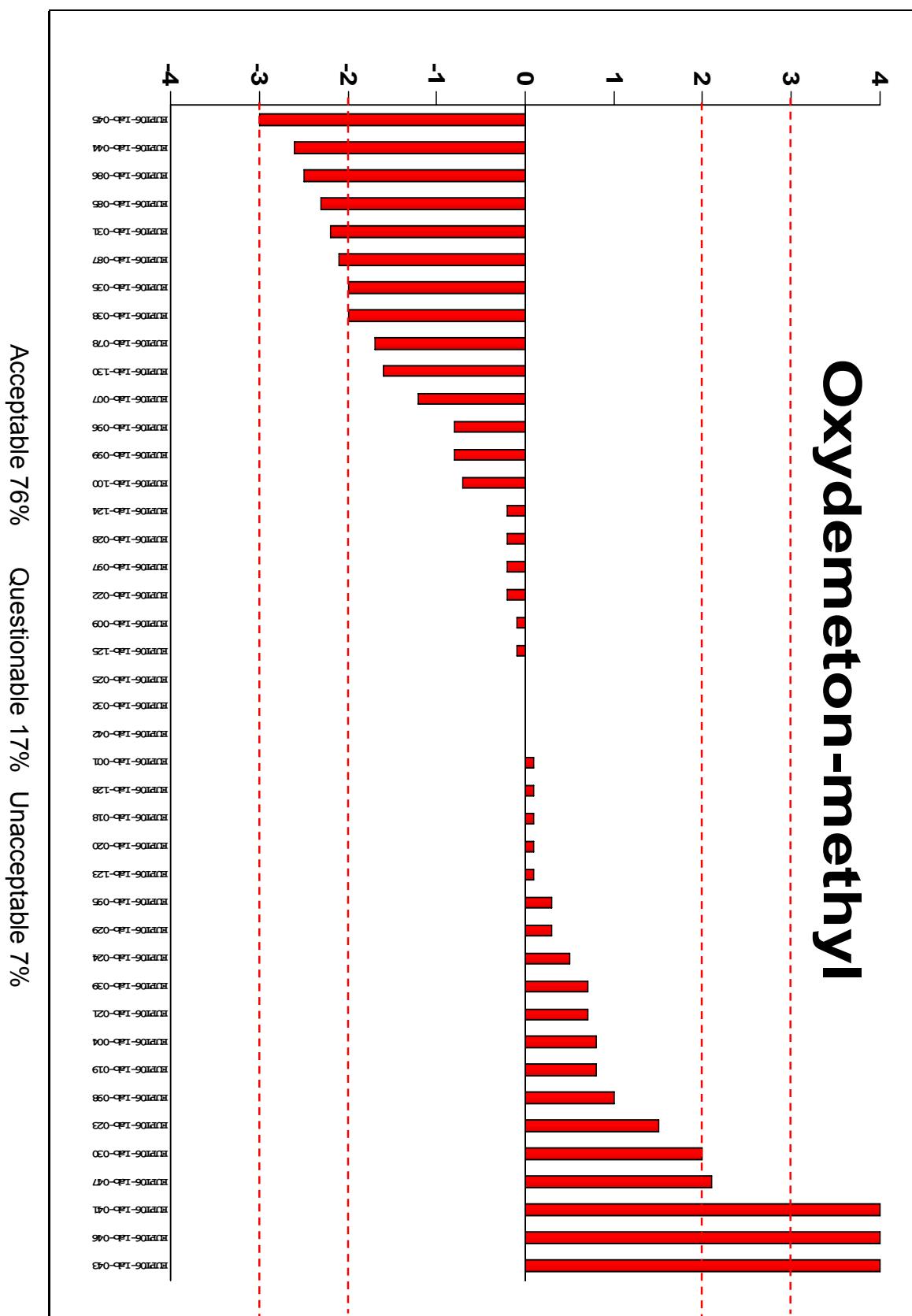
APPENDIX 4.- Graphical representation of z-Scores for FFP (25%).



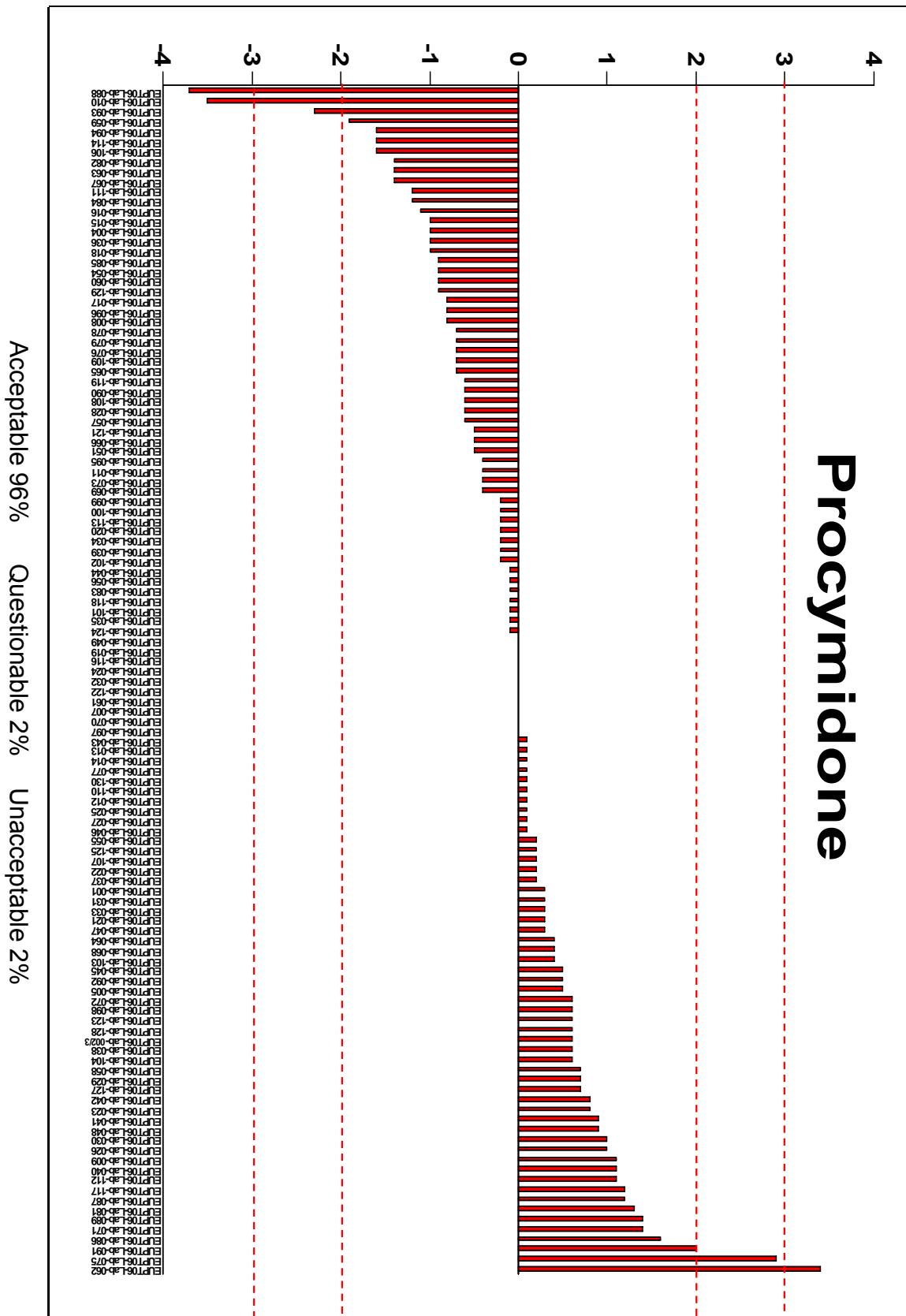
APPENDIX 4.- Graphical representation of z-Scores for FFP (25%).



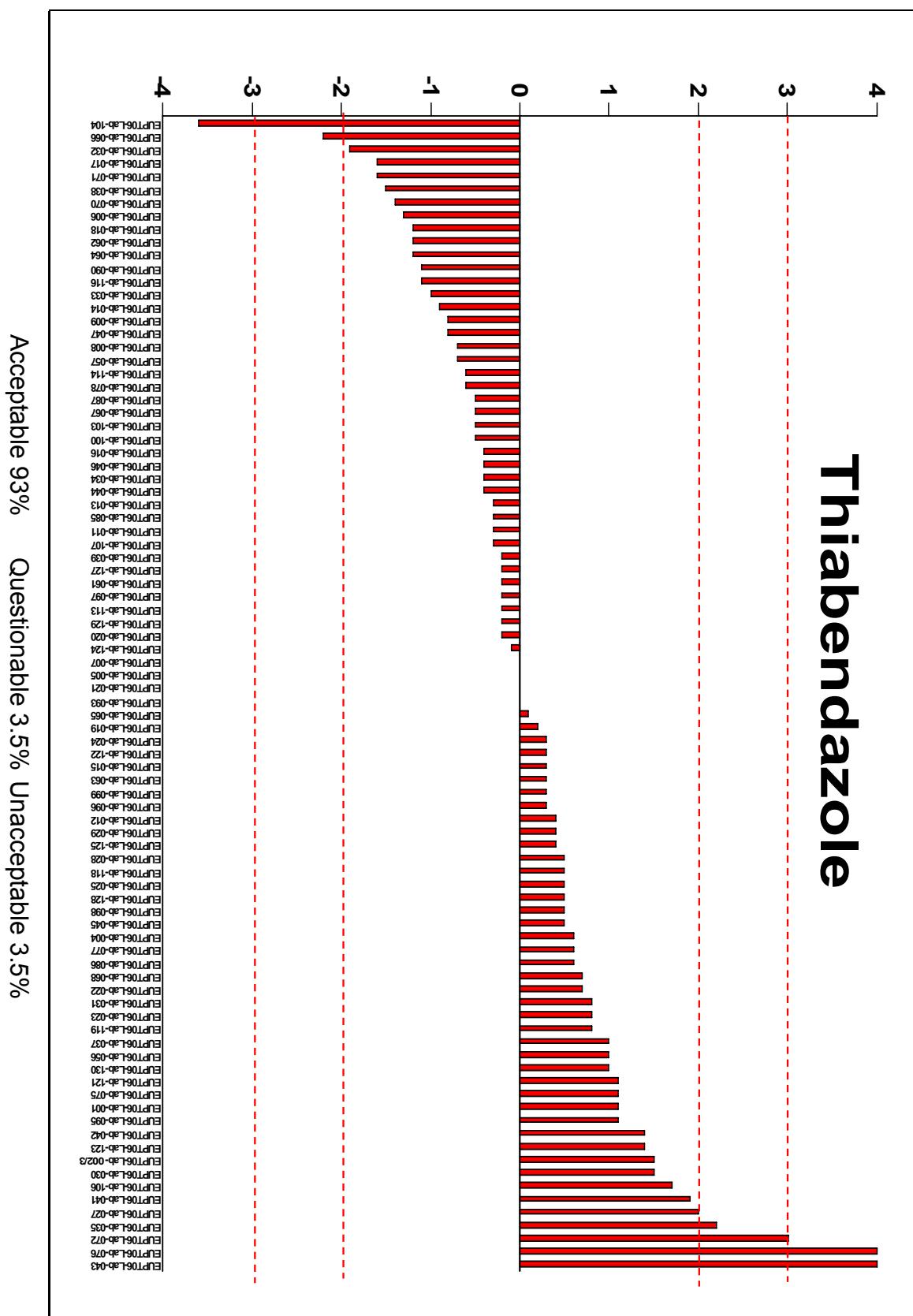
APPENDIX 4.- Graphical representation of z-Scores for FFP (25%).



APPENDIX 4.- Graphical representation of z-Scores for FFP (25%).



APPENDIX 4.- Graphical representation of z-Scores for FFP (25%).



APPENDIX 5. Combined scores RSZ and SSZ for FFP (25%)

Lab Code	n	SSZ	RSZ
Lab-001	13	8.73	2.19
Lab-002/3	11	11.28	2.23
Lab-004	11	9.00	1.36
Lab-005	10	15.82	-0.15
Lab-006	6	13.44	-3.48
Lab-007	12	7.53	0.15
Lab-008	10	53.83	-2.06
Lab-009	12	3.36	-0.05
Lab-010	6	16.45	-0.38
Lab-011	8	36.01	-2.84
Lab-012	11	36.10	-3.09
Lab-013	12	10.24	-1.06
Lab-014	10	26.68	-1.70
Lab-015	10	24.57	-0.94
Lab-016	11	49.64	-5.33
Lab-017	12	47.18	-5.38
Lab-018	13	9.72	-1.97
Lab-019	13	4.66	0.32
Lab-020	13	6.82	-0.65
Lab-021	13	5.26	1.03
Lab-022	12	14.76	0.75
Lab-023	13	10.49	2.56
Lab-024	13	49.23	-3.21
Lab-025	13	3.14	-0.25
Lab-026	9	175.04	7.52
Lab-027	12	13.90	1.06
Lab-028	13	5.93	0.03
Lab-029	12	22.97	1.28
Lab-030	13	26.02	1.83
Lab-031	13	26.97	0.56
Lab-032	13	28.12	-3.15
Lab-033	11	28.68	0.29
Lab-034	11	20.25	-2.60
Lab-035	12	22.61	-1.55
Lab-036	9	25.48	-4.05
Lab-037	10	25.82	1.22
Lab-038	13	34.11	-2.07
Lab-039	13	37.73	-1.30
Lab-040	8	34.16	-2.72
Lab-041	13	59.03	3.55
Lab-042	12	12.28	0.68
Lab-043	13	362.81	11.01
Lab-044	13	193.30	3.03
Lab-045	13	17.29	0.67
Lab-046	12	22.54	1.97
Lab-047	12	47.82	1.69
Lab-048	7	16.03	-0.68
Lab-049	7	0.56	-0.08
Lab-050	3	14.07	-2.15
Lab-051	6	1.75	-0.89
Lab-052	2	1.14	-1.03
Lab-053	0		
Lab-054	8	13.47	-1.91
Lab-055	5	5.76	1.23
Lab-056	8	2.82	0.97
Lab-057	12	6.90	-0.72
Lab-058	10	93.66	-4.31
Lab-059	7	42.43	-6.11
Lab-060	4	14.21	-2.80
Lab-061	12	26.98	0.97
Lab-062	9	44.04	-1.03
Lab-063	8	17.91	-3.03
Lab-064	10	15.25	0.66
Lab-065	13	32.23	-3.89
Lab-066	10	16.48	-3.15
Lab-067	9	106.19	1.63
Lab-068	9	3.18	1.51
Lab-069	9	20.68	0.91

APPENDIX 5. Combined scores RSZ and SSZ for FFP (25%)

Lab Code	n	SSZ	RSZ
Lab-070	10	15.53	1.43
Lab-071	12	11.54	0.70
Lab-072	9	19.06	0.39
Lab-073	8	9.46	1.88
Lab-074		No results	
Lab-075	9	33.16	-0.22
Lab-076	12	281.15	6.49
Lab-077	10	1477.45	12.31
Lab-078	13	11.73	-1.22
Lab-079	9	30.76	-0.67
Lab-080		No results	
Lab-081	7	6.87	1.75
Lab-082	10	18.31	-0.21
Lab-083	6	72.38	5.81
Lab-084	7	8.08	-2.38
Lab-085	13	11.44	-1.12
Lab-086	13	25.51	2.18
Lab-087	12	12.74	1.42
Lab-088	5	52.52	-6.44
Lab-089	7	6.53	0.23
Lab-090	11	38.45	-5.06
Lab-091	7	7.25	0.91
Lab-092	10	13.54	0.53
Lab-093	8	55.87	-5.00
Lab-094	9	77.64	0.60
Lab-095	13	12.23	1.41
Lab-096	13	5.56	-0.85
Lab-097	13	1.70	-0.34
Lab-098	13	4.50	0.94
Lab-099	11	14.10	-1.06
Lab-100	13	4.64	-0.39
Lab-101	10	33.44	-2.53
Lab-102	8	12.61	-1.28
Lab-103	10	16.09	-1.76
Lab-104	9	67.22	-0.67
Lab-105	3	29.30	-5.15
Lab-106	11	21.90	-1.70
Lab-107	12	11.96	-0.61
Lab-108	7	4.57	-0.20
Lab-109	7	4.67	-0.76
Lab-110	10	27.27	0.85
Lab-111	8	3.88	0.32
Lab-112	8	8.94	1.40
Lab-113	9	20.93	2.73
Lab-114	9	9.84	-1.86
Lab-115	9	102.68	-10.09
Lab-116	8	6.07	-1.01
Lab-117	8	29.32	-1.98
Lab-118	10	3.86	-1.01
Lab-119	12	10.08	-2.19
Lab-120	4	23.18	-4.30
Lab-121	11	13.90	-1.40
Lab-122	12	12.51	2.33
Lab-123	13	22.84	2.32
Lab-124	13	18.72	1.72
Lab-125	13	10.50	0.82
Lab-126		No results	
Lab-127	12	40.72	-3.34
Lab-128	13	24.51	2.86
Lab-129	12	20.92	0.40
Lab-130	13	19.92	1.67

APPENDIX 6. z-Score, ‘Weighted Sum of z-Score’ and Graphical Representation for laboratories classified in Category A

z-Score

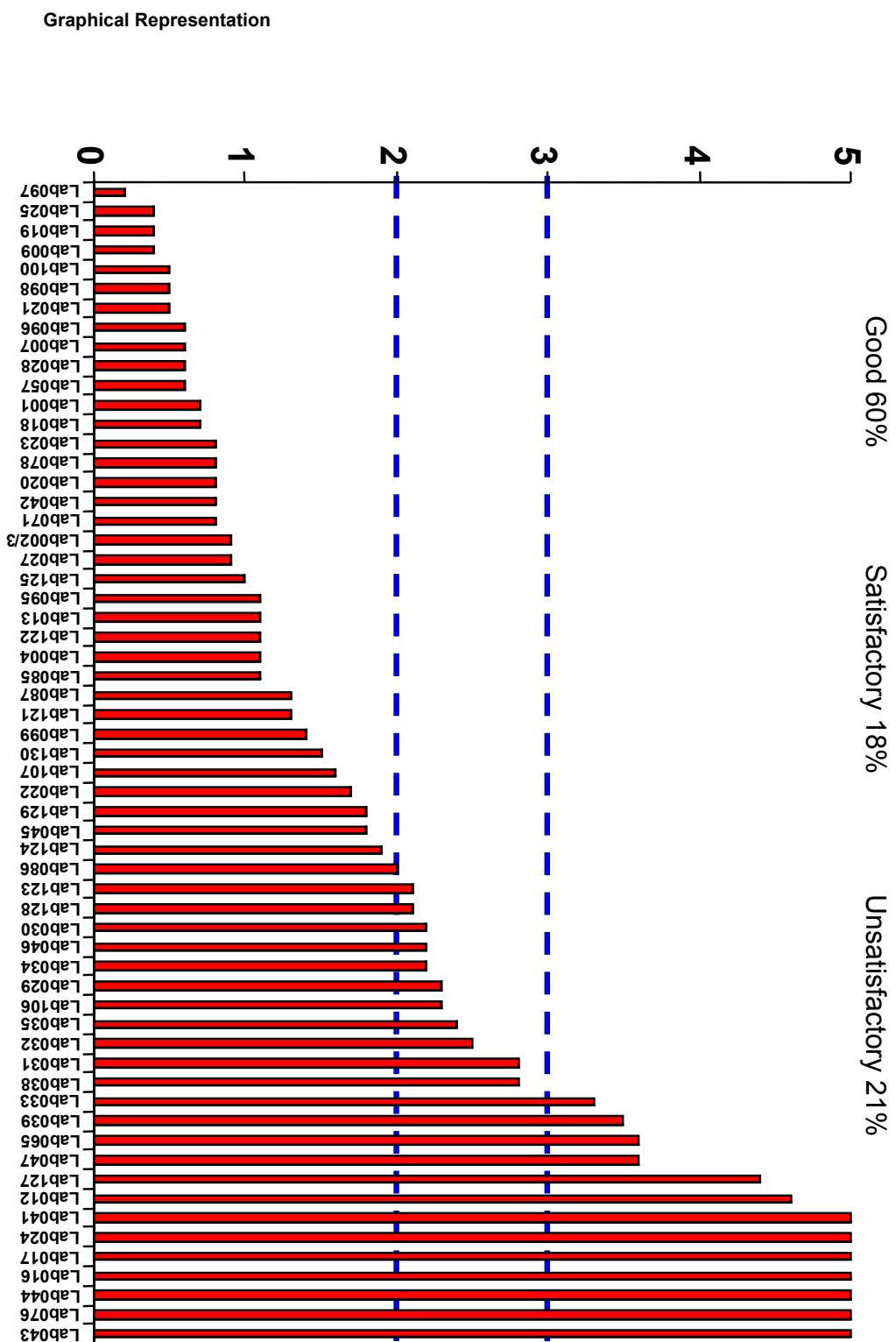
Lab Code	Acrinathrin	Azoxystrobin	Bromopropylate	Chlorothalonil	Diazinon	Dimethoate	Endosulfan	Imazalil	Imidacloprid	Dithiocarbamates (CS ₂)	Oxydemeton-methyl	Procymidone	Thiabendazole
001	1.4	0.2	0.8	0.5	0.4	0.3	0.7	0.6	0.3	1.8	0.1	0.3	1,1
002/3	1.5	0.2	1.0	0.7	0.5	0.0	1.5	1.1		1.1		0.6	1,5
004		0.1	0.2	1.0	0.7	0.2	0.3	2.2	0.5		0.8	1.0	0,6
007	0.6	0.1	0.4	0.7	0.4	0.4	0.1	2.0	0.9		1.2	0.0	0,0
009		0.6	0.2	0.6	0.3	0.6	0.3	0.0	0.1	0.6	0.1	1.1	0,8
012	0.1	3.1	0.1	2.4	3.0	3.2	0.7	0.4		1.1		0.1	0,4
013	0.6	1.0	0.2	1.2	0.5	2.3	1.1	0.3	0.0	0.5		0.1	0,3
016		3.1	0.1	1.8	0.2	0.6	0.4	3.5	3.1		3.6	1.1	0,4
017	3.3	0.9	1.0	2.4	0.5	1.4	0.5		3.1	0.5	3.6	0.8	1,6
018	0.6	1.6	0.7	1.0	0.3	1.0	0.9	0.9	0.2	0.1	0.1	1.0	1,2
019	1.2	0.1	0.2	1.4	0.5	0.1	0.4	0.2	0.0	0.2	0.8	0.0	0,2
020	2.1	0.3	0.0	0.6	0.3	0.3	0.0	0.8	0.2	0.9	0.1	0.2	0,2
021	0.1	0.6	0.8	0.5	0.8	1.1	0.3	0.8	0.2	0.9	0.7	0.3	0,0
022	2.0	0.0	0.5	0.2	0.2	0.4	0.6	3.0	0.3		0.2	0.2	0,7
023	1.5	0.7	1.0	0.3	0.5	0.2	0.1	0.7	0.9	1.1	1.5	0.8	0,8
024	3.3	1.2	0.1	4.0	0.1	3.4	0.0	3.0	0.4	0.4	0.5	0.0	0,3
025	0.1	0.1	0.1	1.1	0.1	0.3	0.4	0.7	0.1	0.9	0.0	0.1	0,5
027	0.1	0.5	1.1	1.7	1.1	0.3	0.8	1.7	0.6	0.8		0.1	2,0
028	0.6	1.1	0.2	0.6	0.7	1.0	0.4	1.2	0.5	0.1	0.2	0.6	0,5
029	3.8	0.7	1.3	0.7	1.3	0.0	1.5	1.0	0.2		0.3	0.7	0,4
030	3.6	0.1	0.6	1.5	0.5	0.4	1.0	1.2	0.1	0.7	2.0	1.0	1,5
031	2.4	3.3	0.2	1.1	0.7	0.6	0.8	0.1	0.2	1.2	2.2	0.3	0,8
032	2.9	0.2	0.1	3.7	0.3	0.5	0.6	1.2	0.2	0.0	0.0	0.0	1,9
033	1.1	2.1	0.3	2.1	0.9	2.0	0.3	3.5		0.5		0.3	1,0
034	3.8	0.3	1.0	2.0	0.1	0.5	0.3	0.4	0.1			0.2	0,4
035	3.3	0.4	0.3	1.5	0.3	0.4	0.0	0.0	0.0		2.0	0.1	2,2
038	0.0	0.8	0.7	2.9	1.1	0.2	0.2	0.4	1.4	3.8	2.0	0.6	1,5
039	3.3	0.8	1.9	0.6	4.1	0.5	0.0	1.1	0.4	1.7	0.7	0.2	0,2
041	2.9	1.5	3.6	0.4	1.4	0.7	0.1	0.1	2.8	0.3	4.5	0.9	1,9
042	0.9	0.6	0.0	1.7	2.0	0.5	0.1	1.1		0.7	0.0	0.8	1,4
043	2.0	5.1	1.2	1.0	0.5	3.1	0.5	1.8	4.1	2.7	8.0	0.1	15,1
044	13.2	0.5	0.9	2.4	0.4	0.1	0.3	1.7	0.9	1.1	2.6	0.1	0,4
045	0.5	0.2	0.7	0.6	1.0	1.4	0.1	1.6	0.7	0.8	3.0	0.5	0,5
046	0.1	0.5	0.1	1.0	0.3	0.4	0.2	0.1		0.4	4.5	0.1	0,4
047	1.4	0.4	0.1	0.9	0.6	0.6	0.1	0.6		6.2	2.1	0.3	0,8
057	0.4	0.3	1.0	0.3	0.2	1.8	0.4	0.4	0.1	1.0		0.6	0,7
065	0.5	0.8	0.6	0.1	2.4	0.2	0.8	0.8	3.1	0.7	3.6	0.7	0,1
071	0.0	0.7	0.8	1.3	0.4	1.0	0.9	0.3	0.3	1.5		1.4	1,6
076	12.3	3.0	0.7	0.2	0.1	4.6	0.5	2.8		2.2	3.6	0.7	8,5
078	1.3	1.0	0.5	0.2	1.1	0.0	0.6	1.7	0.5	0.3	1.7	0.7	0,6
085	0.4	0.4	0.2	0.7	0.4	1.3	0.6	0.6	0.8	0.9	2.3	0.9	0,3
086	1.5	0.1	1.0	1.6	2.6	0.3	1.4	0.4	0.4	1.1	2.5	1.6	0,6
087	0.5	0.4	1.1	1.2	1.2	0.4	0.7	1.0		0.7	2.1	1.2	0,5
095	0.2	0.3	0.4	2.0	0.3	0.2	0.3	2.3	0.9	0.4	0.3	0.4	1,1
096	0.6	0.0	0.8	0.7	0.2	0.2	0.3	1.1	0.4	1.1	0.8	0.8	0,3
097	0.0	0.1	0.4	1.0	0.2	0.2	0.1	0.0	0.3	0.4	0.2	0.0	0,2
098	0.4	0.9	0.9	0.2	0.3	0.8	0.3	0.4	0.3	0.2	1.0	0.6	0,5
099	1.2	0.3	1.0	0.4	0.5	0.5	0.3	1.4		2.9	0.8	0.2	0,3
100	0.0	0.7	0.2	0.1	0.2	0.4	0.6	0.3	0.6	1.5	0.7	0.2	0,5
106	0.4	0.1	0.6	0.8	1.0	0.1	3.4	0.1		1.6		1.6	1,7
107	3.3	0.0	0.4	0.1	0.3	0.2	0.2	0.3	0.4	0.6		0.2	0,3
121		1.7	0.6	0.2	0.0	0.8	0.7	0.5	0.3	2.8		0.5	1,1
122	0.3	2.4	0.0	0.0	0.2	0.7	0.6	1.9	1.4	0.3		0.0	0,3
123	1.6	1.1	0.4	0.0	0.7	1.3	1.3	3.3	0.8	1.2	0.1	0.6	1,4
124	0.8	1.4	0.6	0.1	0.4	0.2	0.4	3.8	0.0	0.9	0.2	0.1	0,1
125	0.4	2.9	0.2	0.1	0.3	0.1	0.2	0.7	0.9	0.0	0.1	0.2	0,4
127	1.1	2.1	3.1	2.0	3.3	1.1	0.4	2.5	0.0	1.6		0.7	0,2
128	0.5	0.2	0.3	0.1	0.3	0.8	1.4	4.4	1.3	0.0	0.1	0.6	0,5
129	1.2	2.1	0.2	1.4	0.0	0.1	1.8	2.5	1.6	0.4		0.9	0,2
130	1.1	0.4	0.6	1.1	0.7	1.8	1.5	2.7	0.5	0.5	1.6	0.1	1,0

APPENDIX 6. z-Score, ‘Weighted Sum of z-Score’ and Graphical Representation for laboratories classified in Category A

‘Weighted Sum of z-Score’

Lab Code	n	Weighted Sum
Lab001	13	0.7
Lab002/3	11	0.9
Lab004	11	1.1
Lab007	12	0.6
Lab009	12	0.4
Lab012	11	4.6
Lab013	12	1.1
Lab016	11	6.5
Lab017	12	5.4
Lab018	13	0.7
Lab019	13	0.4
Lab020	13	0.8
Lab021	13	0.5
Lab022	12	1.7
Lab023	13	0.8
Lab024	13	5.0
Lab025	13	0.4
Lab027	12	0.9
Lab028	13	0.6
Lab029	12	2.3
Lab030	13	2.2
Lab031	13	2.8
Lab032	13	2.5
Lab033	11	3.3
Lab034	11	2.2
Lab035	12	2.4
Lab038	13	2.8
Lab039	13	3.5
Lab041	13	5.0
Lab042	12	0.8
Lab043	13	14.8
Lab044	13	6.7
Lab045	13	1.8
Lab046	12	2.2
Lab047	12	3.6
Lab057	12	0.6
Lab065	13	3.6
Lab071	12	0.8
Lab076	12	14.3
Lab078	13	0.8
Lab085	13	1.1
Lab086	13	2.0
Lab087	12	1.3
Lab095	13	1.1
Lab096	13	0.6
Lab097	13	0.2
Lab098	13	0.5
Lab099	11	1.4
Lab100	13	0.5
Lab106	11	2.3
Lab107	12	1.6
Lab121	11	1.3
Lab122	12	1.1
Lab123	13	2.1
Lab124	13	1.9
Lab125	13	1.0
Lab127	12	4.4
Lab128	13	2.1
Lab129	12	1.8
Lab130	13	1.5

APPENDIX 6. z-Score, 'Weighted Sum of z-Score' and Graphical Representation for laboratories classified in Category A



ACRINATHRIN

APPENDIX 7. Methods used by participants for determining pesticides

Lab Code									
Quantification Method									
Quantification using Standards in solvents or matrix									
Confirmation method									
RL (mg/kg)									
Recovery									
Sample Weight (g)									
Extraction Solvent									
Partition Step (if any)									
Clean up step									
Internal Standard (if any)									
Single level calibration									
Multi-level calibration									
Nº of points									
Column type									
Stationary phase									
Determination									
001	D	S	GC-MS	0.02	110	15	4		
002/3	D	M	GC-MS	0.01	102	15	4	PCB97	Yes
004	NA								3
005	NA								60x0.25x0.15
006	D	M	GC-MS	0.02	94	10	1	O	Yes
007	D	M	GC-ECD	0.03	87	25	2	GPC Dithalimphos	5
008	NA								J&W 0.32/0.25mmID
009	NA								DB5, DB1701
010	NA								GC-ECD, GC-NPD, GC-ITD
011	NA								
012	D	M	GC-MS	0.01	70-110	50	3		Yes 8 HP5MS
									GC-MS

ACRINATHRIN

APPENDIX 7. Methods used by participants for determining pesticides

Determination									
Stationary phase									
Column type									
Lab Code	Partion Step (if any)								
Quantification Method	Clean up step								
Quantification using Standards in solvents or matrix	Internal Standard (if any)								
Confirmation method	Single level calibration								
RL (mg/kg)	Multi-level calibration								
Recovery	Nº of points								
Sample Weight (g)	Column type								
Extraction Solvent	Stationary phase								
Partion Step (if any)	Determination								
Clean up step	Determination								
Internal Standard (if any)	Determination								
Single level calibration	Determination								
Multi-level calibration	Determination								
Nº of points	Determination								
Column type	Determination								
Stationary phase	Determination								
Determination	Determination								
013	D	S	GC-ECD	0.01	85	25	Ethyl Acetate Dichloromethane	SPE	GC-ECD/ELCD
014	NA								
015	NA								
016	NA								
017	ND								
018	D	S	GC-MS	0.01	100	2	GPC	Yes	Yes
019	D	M	GC-TOF	0.02	102	10	PCB 138	Yes	4
020	D	M	GC-MS	0.010	25	2	GPC	Yes	3
021	D	S	GC-MS	0.01	98.1	50	GPC	Yes	BD 1/5/35/1301. HP50/170/5MS
022	D	S	0.01	47	50	6; Acetone	Cyclohexane Ethyl acetate	Trans-HCEO	Capillary
023	D	M	GC-MS	0.05	50	2	GPC	Yes	DB5/210
024	ND		0.05				GC-MS		GC-MS

ACRINATHRIN

APPENDIX 7. Methods used by participants for determining pesticides

Determination									
Stationary phase									
Column type									
Partion Step (if any)	Clean up step	Internal Standard (if any)	Single level calibration	Multi-level calibration	Nº of points	Column type	Stationary phase	Determination	
Extraction Solvent									
Sample Weight (g)	Recovery								
RL (mg/kg)									
Confirmation method									
Quantification using Standards in solvents or matrix	Quantification Method								
Lab Code									
025	D	M	GC-MS	0.02	50	3	GPC	Yes	3
026	D	S	GC-MS	0.05	100	2	GPC	Yes	3
027	D	M	GC-MS	0.02	100	2	GPC	EPSILON-HCH	Yes
028	D	M	GC-MSD	0.05	97.4	10	others	Triphenyl-phosphat	Yes
029	D	M	GC-MS	0.01	100	2	GPC	Yes	3-5
030	D	M	GC-ECD	0.02	50	6	GPC	Bromophos-ethyl	Yes
031	D	S	GC-MS	0.01	10	5	SPE	Yes	2
032	D	M	GC-MS	0.01	70.3	30	GPC	Ethyl acetate Ciclohexane (2:1)	Yes
033	D	M	GC-NPD	0.02	100	2	GPC		Yes
034	D	M	GC-MS	0.01	100	2	PCB 153		Yes
035	ND			0.05	50	3	GPC		Yes

ACRINATHRIN

Determination													
Stationary phase													
Column type													
Partion Step (if any)	Clean up step	Internal Standard (if any)	Single level calibration	Multi-level calibration	Nº of points								
Extraction Solvent	Partion	Parathion	Yes	Yes	3								
Sample Weight (g)	Recovery												
RL (mg/kg)	Confirmation method												
Quantification Method	Quantification using Standards in solvents or matrix												
Lab Code													
036	D	S	GC-MS	0.05	101	3	GPC	Yes	GC-ECD/NPD/MS				
037	D	M	GC-MS	0.03	100	2	GPC	Yes	GC-MS				
038	D	M	GC-MS	0.01	92	20	Triphenyl-phosphat	Yes	Optima5, DB17-MS				
039	ND	S		0.01	45	3	GPC	Aldrin	GC-ECD, GC-NPD.				
040	D	S	GC-ECD	0.05	340	2	O ₂ GP C	Parathion	GC-MS				
041	D	S	GC-MS	0.02	20	75	Acetone+ Cyclohexan+ Ethyl Acetate	Yes	Restek 30x0.25x0.25				
042	D	M	GC-MS	0.02	100	2	GPC	EPN	RTX-CLPesticide I+II				
043	D	S	GC-MS	0.08	50	3	GPC	Parathion	GC-ECD				
044	D	M		0.02	100	2	GPC		GC-ECD				
045	D	M	GC-ECD, GC-NPD, GC-MS	0.02	100	2	GPC	Yes	GC-ECD, GC-NPD, GC-FPD, GC-MS (single-quad)				
046	D	M	GC-ECD	0.01	100	10	O(PSA)	MIREX	DB5, DB1701				
									GC-ECD				

APPENDIX 7. Methods used by participants for determining pesticides

Determination									
Stationary phase									
Column type									
Lab Code	D	M	GC-ECD	0.01	100	3	GPC	Yes	4
Quantification Method									
047	D	M	GC-ECD	0.01	100	3	GPC	Yes	4
048	NA								
049	D	M	GC-MS	0.02	92.4	15	4	2	DB5MS
050	NA								GC-ECD/GC-T
051	NA								
052	NA								
053	NA								
054	NA								
055	NA								
056	NA								
057	D	S	GC-MS	0.05	20	6	GPC-SPE	Yes	3
058	NA								DB5
059	NA								Me Polisiloxan
060	NA								GC-MS
061	D	M	GC-MS	0.02	87.2	15	6 (Dichloromethane)	Yes	5
							5%Phenyl 95%Dimethyl- Polysiloxane		

ACRINATHRIN

APPENDIX 7. Methods used by participants for determining pesticides

		Lab Code			
		Quantification Method			
		Quantification using Standards in solvents or matrix			
		Confirmation method			
		RL (mg/kg)			
		Recovery			
		Sample Weight (g)			
		Extraction Solvent			
		Partition Step (if any)			
		Clean up step			
		Internal Standard (if any)			
		Single level calibration			
		Multi-level calibration			
		Nº of points			
		Column type			
		Stationary phase			
		Determination			
062	NA				
063	NA				
064	NA				
065	D	S	GC-MS	0.02	84.3
066	NA				
067	NA				
068	D	M	GC-MS	0.05	82.8
069	NA				
070	NA				
071	D	M	GC-MS	0.01	15
072	NA				

APPENDIX 7. Methods used by participants for determining pesticides

ACRINATHRIN

	Lab Code	
	Quantification Method	
073	Quantification using Standards in solvents or matrix	
074	Confirmation method	
	RL (mg/kg)	
	Recovery	
	Sample Weight (g)	
	Extraction Solvent	
	Partition Step (if any)	
	Clean up step	
	Internal Standard (if any)	
	Single level calibration	
	Multi-level calibration	
	Nº of points	
	Column type	
	Stationary phase	
	Determination	
080	NO RESULTS	
081	NA	
082	D	M
083	NA	
084	NA	
085	D	M
086	D	M

ACRINATHRIN

APPENDIX 7. Methods used by participants for determining pesticides

Determination									
Stationary phase									
Column type									
Partion Step (if any)	Clean up step	Internal Standard (if any)	Single level calibration	Multi-level calibration	Nº of points				
Extraction Solvent	Partion Step (if any)	Internal Standard (if any)	Single level calibration	Multi-level calibration	Nº of points				
Sample Weight (g)	Extraction Solvent	Internal Standard (if any)	Single level calibration	Multi-level calibration	Nº of points				
Recovery	Sample Weight (g)	Internal Standard (if any)	Single level calibration	Multi-level calibration	Nº of points				
RL (mg/kg)	Recovery	Internal Standard (if any)	Single level calibration	Multi-level calibration	Nº of points				
Confirmation method	RL (mg/kg)	Internal Standard (if any)	Single level calibration	Multi-level calibration	Nº of points				
Quantification using Standards in solvents or matrix	Confirmation method	Internal Standard (if any)	Single level calibration	Multi-level calibration	Nº of points				
Quantification Method	Quantification using Standards in solvents or matrix	Internal Standard (if any)	Single level calibration	Multi-level calibration	Nº of points				
Lab Code	Quantification Method	Internal Standard (if any)	Single level calibration	Multi-level calibration	Nº of points				
087 D M GC-MS 0.02	10 4	Yes	3	HP-5					
088 D S MS 0.03	15 4	Yes	5	NarrowBore	HP5-MS				
089 D M GC-MS/MS 0.02	15	Yes	4	Capillary	BP5				
090 D M GC-MS/MS 0.02	92.10 15	TPP(QC)	Yes	3	Capillary	DB-5ms	GC-MS/MS		
091 D M GC-MS/MS 0.02	15 1	TPP, Anthracene	Yes	3	HP-5MS		GC-ITD		
092 D M GC-MS/MS 0.05	5.2 4		Yes	6	Factor Four		GC-MS/MS-ECD		
093 NA									
094 D M MS/MS	15 4		Yes	4	30mx0.25x0.2	CPSIL8	GC-ECVD-GC-NPD,GC-PFPD		
095 D M GC-MS 0.05	86 75 1	GPC	Yes	Yes	3	CPSIL, 5CB,CPSIL, 8CB	GC-ECVD,GC-FPD,GC-ITD		
096 D M GC-ITD 0.05	108 15 4		Yes	4	Capillary, 25mx0.22mmID	VF-5MS, 0.25μm (Varian)	GC-ITD		
097 D M GC-MS 0.02	95 30 1	GPC	Yes	5	Capillary	DB-5	GC-MS		

ACRINATHRIN

APPENDIX 7. Methods used by participants for determining pesticides

Lab Code	Quantification Method					
	Quantification using Standards in solvents or matrix					
Confirmation method						
RL (mg/kg)						
Recovery						
Sample Weight (g)						
Extraction Solvent						
Partition Step (if any)						
Clean up step						
Internal Standard (if any)						
Single level calibration						
Multi-level calibration						
Nº of points						
Column type						
Stationary phase						
Determination						
098	D	M	GC-MS	0.05	10	1
099	NA				SPE	Yes
100	D	M	GC-MS	0.02	88	10
101	D	M	GC-MS	0.05	90	50
102	NA				GPC	Yes
103	NA				SPE	Yes
104	NA				HP 5/HP 50	See column type
105	NA					GC-ECD.GC-NPD
106	D	M	GC-MS	7.5	1	EthyLParathion
107	ND			0.02	20	4
108	NA				DB-1	0.25mm
109	NA				Capillary 30 m	HP-5
110	D	M	GC-MS	0.02	88.80	5
111	NA				DB-5	GC-ECD/GC-NPD
112	NA					GC-ECD/GC-NPD
113	NA					

ACRINATHRIN

Determination									
Stationary phase									
Column type									
124	D	S	0.05	30	1	GPC	Yes	3	Supelco
123	D	S	GC-MS	0.05	30	1	GPC	Yes	30mx0.32mm
122	D	S	GC-MS	0.05	97.80	30	1	Yes	Chromopack 50mx0.25mmx0.25mm
121	NA						NO	3	CPSIL-8CB
120	NA						Yes		GC-ECD
119	D	M	GC-MS	0.05	10	1	NO		DB-5MS
118	D	M	GC-MS	0.02	105	10	6	Yes	35%Phenyl GC-MS
117	NA						SPE	3	DB-35MS
116	NA								
115	NA								
114	NA								

ACRINATHRIN

APPENDIX 7. Methods used by participants for determining pesticides

Determination												
Stationary phase												
Column type												
Lab Code	Clean up step											
Quantification Method	Internal Standard (if any)											
Quantification using Standards in solvents or matrix	Single level calibration											
Confirmation method	Multi-level calibration											
RL (mg/kg)	Nº of points											
Recovery	Column type											
Sample Weight (g)	Stationary phase											
Extraction Solvent	Determination											
Partition Step (if any)	Determination											
Clean up step	Determination											
Internal Standard (if any)	Determination											
Single level calibration	Determination											
Multi-level calibration	Determination											
Nº of points	Determination											
Column type	Determination											
Stationary phase	Determination											
126	NO RESULTS											
127	D		0.002	30	1	GPC		Yes	3	50mx0.32mmx0.25um	CP-SII 8 CB	GC-ECD
128	D	S	0.05	95	30	1		Yes	3	30mx0.25mmx0.25um	RTX-CLP	GC-NPD
129	D	S	0.05	30	1	GPC		Yes	3	Restek	Stx-Cl-Pest.	GC-ECD
130	D	M	0.05	30	1	GPC		Yes	3	Restek	RTX-5MS	GC-ECD

AZOXYSTROBIN

APPENDIX 7. Methods used by participants for determining pesticides

	Lab Code											
	Quantification Method		Quantification using Standards in solvents or matrix		Confirmation method		RL (mg/kg)		Recovery		Sample Weight (g)	
001	D	S	GC-MS	0.05	103	15	4		Yes	3-4	DB5 DB 1701	ECD NPD MS
002	D	M	GC-MS	0.05	99	15	4		PCB97	Yes	3	60x0.25x0.1 60x0.25x0.15
003												DB5 DB1701
004	D	S	GC-MS/LC-MS	0.05	118	20	6		Nitrofen+ Mirex	Yes	5	CAP
005	D	M	GC-MS	0.15	100	50	6	O+LL		Yes	3	15x0.32x0.25
006	D	M	LC-MS-MS	0.05	94	10	5			Yes	5	R-P
007	D	M	GC-ECD	0.01	83	25	2	GPC	Dithalimphos	Yes	5	J&W 0.32x0.25mmID
008	D	M		0.01	63	25	2					DB5, DB1701
009	D	M	GC-MS	0.03	112.0	50	4	Y _{es}	L _l Aldrin Ditalimphos	Yes	5	DB
010	NA											1701/05/01/35
011	ND	S	GC-MS	0.05	25	1		SPE		Yes	4	30mx0.53mmx1.5μm
												DB5, DB608
												GC-ECD, GC-MS

AZOXYSTROBIN

APPENDIX 7. Methods used by participants for determining pesticides

Determination									
Stationary phase									
Column type									
Partion Step (if any)	Clean up step	Internal Standard (if any)	Single level calibration	Multi-level calibration	Nº of points				
Extraction Solvent									
Sample Weight (g)	Recovery	RL (mg/kg)	Confirmation method	Quantification using Standards in solvents or matrix	Quantification Method				
Lab Code									
012	ND	M	GC-MS	0.05	60-70	50	3		GC-MS
013	D	S	LC-MS/MS	0.01	76	15	4		GC-MS
014	D	S	GC-MS	0.05	40	3		Yes	8
015	ND			0.1	50	1/DCM (80/20)	LL	Yes	3
016	ND	S		0.02	50	3	Y _{es}	Yes	3
017	D	S	GC-MS	0.01	40	2		Yes	4
018	D	S	GC-MS	0.013	100	2		Yes	5
019	D	M	GC-TOF	0.005	94	10	5	Yes	5
020	D	M	GC-MS	0.010	25	2		Yes	4
021	D	S	GC-MS	0.03	132.6	50	2	Yes	3
022	D	M		0.01	92	10	6 _i (Methanol)	Yes	7
023	D	M	LC-MS/MS	0.01	10	10	Methanol	Yes	5

AZOXYSTROBIN

APPENDIX 7. Methods used by participants for determining pesticides

Determination											
Stationary phase											
Column type											
Internal Standard (if any)	Single level calibration	Multi-level calibration	Nº of points	Partion Step (if any)	Clean up step	Extraction Solvent	Sample Weight (g)	Recovery	RL (mg/kg)	Confirmation method	Lab Code
Determination	GC-ECD	GC-NPD	GC-MS	Extraction Solvent	Clean up step	Internal Standard (if any)	Sample Weight (g)	Recovery	RL (mg/kg)	Confirmation method	Lab Code
024 D M GC-ECD 0.01	Acetonitril	PCB 209	Yes	9	ZP 5	Dimethyl-polysiloxan	GC-MS/ECD				024
025 D M LC-MS/MS 0.006		20	Yes	2x4	HPLC(RP)	RPC(18)	LC-MS/MS				025
026 D S GC-ECD 0.05	100	2	Yes	3	Capillary	Varian VF-5MS	GC-ECD GC-NPD GC-MS				026
027 D M GC-MS	100	2	GPC	EPSILON-HCH	30mx0.25x0.25	DB-5, Factor four	GC-ECD				027
028 D M GC-MSD 0.02	120	10	Others	Triphenyl-phosphat	Yes	7	KS	HPMS	GC-MS		028
029 D M GC-MS	0.02	100	2	GPC SPE	Yes	3-5	GC	DB5 HP5-MS DB17	GC-ECD GC-NPD GC-MS (single-quad)		029
030 D M GC-NPD 0.03	50	6	GPC	Bromophos-ethyl	Yes		Capillary-GC	ZB5	GC-NPD		030
031 D S GC-MS 0.02	50	2	GPC		Yes	2	Capillary	HP 5	GC-ECD GC-NPD GC-MS		031
032 D M GC-MS 0.01	95.8	30	Ethyl Acetate/ Cyclohexane (2:1)	GPC	Yes	4	DB5		GC-ECD, GC-NPD, GC-MS		032
033 D M GC-ECD 0.03	100	2	GPC		Yes	3	25m x 0.32mmx0.52μm	Ultra2	GC-ECD/NPD		033

AZOXYSTROBIN

APPENDIX 7. Methods used by participants for determining pesticides

Determination											
Stationary phase											
Column type											
Lab Code	Quantification Method	Quantification using Standards in solvents or matrix	Confirmation method	RL (mg/kg)	Recovery	Sample Weight (g)	Extraction Solvent	Partition Step (if any)	Clean up step	Internal Standard (if any)	Single level calibration
034 D M GC-MS 0.01				100 2			PCB 153	Yes 2	30mx0.3mmx0.25µm	DB5 OÜ1701	GC-ECD
035 D M LC-MS/MS 0.01				10 5	O			Yes 5	Aqua5µ. 50x2mm	RP18	LC-MS/MS
036 D M GC-MS 0.05				101 3	GPC	Yes					GC-ECD/NPD/MS
037 NA											
038 D M LC-MS/MS 0.05				88 20	2		Triphenyl-phosphat	Yes 3	Capillary-GC	Optima5 Optima 17. DB17-MS. C18. Aqua5µ	GC-ECD, GC-NPD, GC-MS, LC-MS/MS
039 D S GC-MS 0.05				45 3	LL	Yes 3	Restek 30x0.25x0.25	RTX-CLPesticide I+II	GC-ECD		
040 NA											
041 D S GC-MS 0.05		Acetone Followed By Cyclohexan And Ethyl Acetate 101 75		GPC		Yes 3	HP5			GC-ECD	
042 D M GC-MS 0.02		100 2		GPC EPN		Yes 3	Capillary	DB5, DB1701. DB35	GC-ECD, GC-NPD, GC-FPD, GC-MS (single-quad)		
043 D M LC-MS/MS 0.006		10 6 (Methanol)		SPE		Yes 2		C18	LC-MS/MS		

AZOXYSTROBIN

APPENDIX 7. Methods used by participants for determining pesticides

Determination									
Stationary phase									
Column type									
Sample Weight (g)	Extraction Solvent	Partion Step (if any)	Clean up step	Internal Standard (if any)	Single level calibration	Multi-level calibration	Nº of points	Column type	Stationary phase
Recovery	RL (mg/kg)	Confirmation method	Quantification using Standards in solvents or matrix	Quantification Method	Lab Code	Lab Code	RP8	LC-MS/MS	Determination
044 D M 0.01 100 2	GPC		GC-ECD, GC-NPD, GC-MS	0.03	045 D M 0.03 100 2	Yes	4	RP8	LC-MS/MS
045 D M 0.03 100 2	GPC		GC-ECD, GC-NPD, GC-MS	0.03	046 D M 0.01 109 50 3	Yes	5	DB5, DB1701	GC-ECD, GC-NPD, GC-MS
046 D M 0.01 109 50 3	GPC	Y _{es}	GC-ECD, GC-NPD, GC-MS	0.01	047 D M 0.01 100 2	Capillary	DB17, HP5	GC	
047 D M 0.01 100 2	GPC	Y _{es}	GC-ECD, GC-NPD, GC-MS	0.01	048 NA	Capillary	DB5, DB1701	GC-ECD, GC-NPD, GC-FPD	
048 NA					049 NA				
049 NA					050 NA				
050 NA					051 NA				
051 NA					052 NA				
052 NA					053 NA				
053 NA					054 D M GC-ECD 0.03 90.5 25 6 Y _{es}	O	Yes	3	HP5MS
054 D M GC-ECD 0.03 90.5 25 6 Y _{es}	GC-ECD	O	GC-ECD	0.03	055 NA		Yes	3	5% Phenyl
055 NA					056 D M 0.02 90.35 15 4 GPC		Yes	5	HP5MS
056 D M 0.02 90.35 15 4 GPC					057 D S GC-MS 0.05 20 6 GPC-SPE	GC-MS	Yes	5	GC-MS
057 D S GC-MS 0.05 20 6 GPC-SPE		Y _{es}	Me Polisiloxan	0.05		DB5	3	GC-MS	

AZOXYSTROBIN

APPENDIX 7. Methods used by participants for determining pesticides

Determination									
Stationary phase									
Column type									
Nº of points									
Single level calibration									
Multi-level calibration									
Internal Standard (if any)									
Clean up step									
Partition Step (if any)									
Extraction Solvent									
Sample Weight (g)									
Recovery									
RL (mg/kg)									
Confirmation method									
Quantification using Standards in solvents or matrix									
Quantification Method									
Lab Code									
058	ND	M	GC-MS	0.1	15	1	GPC	✓Yes	✓Yes
059	NA								
060	NA								
061	D	M	GC-MS	0.05	75.1	15	6-Dichloro-methane	✓Yes	5 DB5
062	NA								
063	NA								
064	D	S	GC-MS	111.3	10	6	O	✓Yes	✓Yes
065	D	S	GC-MS	0.03	75	10	Dichloro-methane	✓Yes	HP5
066	D	S	GC-MS	0.05	95	25	✓Yes	✓Yes	POLYSILOXANE
067	NA								GC-ECD
068	NA								GC-ECD/NPD

AZOXYSTROBIN

APPENDIX 7. Methods used by participants for determining pesticides

Lab Code	Quantification Method									
	Quantification using Standards in solvents or matrix									
Confirmation method										
RL (mg/kg)	Recovery									
	Sample Weight (g)									
Extraction Solvent										
Partion Step (if any)	Clean up step									
	Internal Standard (if any)									
Single level calibration	Determination									
	Multi-level calibration									
Nº of points	Column type									
	Stationary phase									
069										
D	S	GC-MS	81	5	1	AS-E (Dionex)	GPC	Ronnel+Azo benzene+ Triphenyl Phosphate	Yes	MDN-5S
070										
D	S	GC-ECD	0.01	15	6		GPC	M-series	Yes	5%PHENYL SILICONE
071										
D	M	GC-MS	0.01	15	6		GPC		Yes	25m ID 0.32 mm 0.25um
072										
D	S		0.01	73	10		GPC	Dichloro- fertition	Yes	SE 54; OV1701
073										
D	S	GC-ECD	0.02	10	Dichloro- methane				Yes	HP1. HP5
074										
NO RESULTS										
075										
NA										
076										
D	M	GC-MS	0.05	10	5	Yes	O		Yes	3
077										
D	S	GC-MS	0.05	43	25	2	GPC		Yes	4
078										
D	M	GC-MS	0.05	64	50	1	GPC		Yes	4
079										
D	M	GC-ECD	0.05	15	4			Narrowbore	1	HP-1
080										
NO RESULTS										

AZOXYSTROBIN

APPENDIX 7. Methods used by participants for determining pesticides

		Lab Code	
081	NA	Quantification Method	
082	ND	Quantification using Standards in solvents or matrix	
083	NA	Confirmation method	
084	NA	RL (mg/kg)	
085	D	Recovery	
086	D	Sample Weight (g)	
087	D	Extraction Solvent	
088	NA	Partition Step (if any)	
089	NA	Clean up step	
090	M	Internal Standard (if any)	
091	NA	Single level calibration	
092	D	Multi-level calibration	
		Nº of points	
		Column type	
		Stationary phase	
		Determination	
		TPP(QC)	
		Yes	
		6	Factor Four
		GC-MS/MS-ECD	

AZOXYSTROBIN

APPENDIX 7. Methods used by participants for determining pesticides

Determination									
Stationary phase									
Column type									
093	NA								
094	NA								
095	D	M	GC-MS	0.1	81	75	1		
096	D	S	GC-ITD	0.01	103	7.5	4(+Na ₂ SO ₄)		
097	D	M	GC-MS	0.02	86	30	1		
098	D	M	LC-MS/MS	0.05		10	1		
099	D	M	GC-MS	0.01	80	30	1	O	
100	D	M	LC-MS/MS	0.05	93	10	1		
101	D	M	GC-MS	0.05	90	50	5	SPE	
102	D	M	GC-ITD	0.05	85	50	1	GPC	
103	D	S	GC-MS	0.02	83	20	3	LL	
104	D	M	GC-MS	0.02	88	75	1	1/20	
105	NA								
106	D	M	GC-MS	0.05	7.5	1		Ethyl-parathion	Y
								DB-1	
								0.25mm	GC-ECD/MS

AZOXYSTROBIN

APPENDIX 7. Methods used by participants for determining pesticides

Determination												
Stationary phase												
Column type												
Nº of points	Single level calibration	Multi-level calibration	Internal Standard (if any)	Clean up step	Partion Step (if any)	Extraction Solvent	Sample Weight (g)	Recovery	RL (mg/kg)			
Determination	GC-ECD/NPD	GC-ECD	5%Phenyl	HP-5	Capillary 60 m	HP-5	GC-MS	GC-MS	Lab Code			
107 D M GC-MS 0.05 117 20 4 Yes 3 Capillary 60 m HP-5	108 D S GC-ECD 0.05 93 5 3 Yes 3 HP-5	109 D S GC-EC 0 91 100 3 Yes 3 HP-1	110 D M GC-MS 0.02 92.80 20 3 SPE Yes 5 DB-5	111 D S 0.05 98.90 10 4 SPE (Silica) Yes 5 HP-5MS	112 D S GC-NPD 0.05 95 100 3 O Yes 5 HP1	113 D M GC-TD 0.026 103 15 4 NO Hexachlorobenzene Yes 5 DB-5MS	114 D S GC-ECD 0.09 18 1 GPC Yes 4 Capillary HP5	115 ND GC/MS 0.05 50 4 Dichloromethane LL Yes HP(30x0.25x0.25)	116 D S GC/MS 0.01 92 50 2 SPE Yes HP5-MS	117 D M GC-MS 0.05 102.20 20 4 Yes GPC Yes 5 HP-5MS	118 D M GC-MS 0.02 101 10 6 SPE Yes 3 DB-35MS 35%Phenyl	119 D M GC-MS 0.05 10 1 NO Yes Capillary DB-5MS

AZOXYSTROBIN

APPENDIX 7. Methods used by participants for determining pesticides

Determination										
Stationary phase										
Column type										
Nº of points										
Single level calibration										
Multi-level calibration										
Internal Standard (if any)										
Clean up step										
Partion Step (if any)										
Extraction Solvent										
Sample Weight (g)										
Recovery										
RL (mg/kg)										
Quantification Method										
Quantification using Standards in solvents or matrix										
Confirmation method										
Lab Code										
120	NA									
121	D	M	GC-MS	0.03	25	1	GPC	Yes	5	Capillary
122	D	S	GC-ECD	0.05	106	30	1	GPC	Yes	4 Agilent Technologies
123	D	S	HPLC	0.05	30	1	GPC	Yes	30mx0.32mm	CP-Sil 8 CB
124	D	S	TLC	0.05	30	1	GPC	Yes	3	Supelco
125	D	S	GC-MS	0.05	87	25	1	GPC	Yes	5 Capillary
126										
NO RESULTS										
127	D	S		0.03	101.5	30	1	O	Yes	4 Nucleosil 100
										C-18
										HPLC-UV
128	D	S	GC-ECD	0.05	104	30	1	GPC	Yes	3 30mx0.25mm x0.25um
129	D	S	HPLC-UV	0.05	30	1	GPC	Yes	3 Restek	RTX-CLP
130	D	M	GC-ECD	0.05	30	1	GPC	Yes	3 Waters	Six-Ch-Pest.
										GC-ECD
										Nova-Pack C-18
										HPLC-UV

BROMOPROPYLATE

APPENDIX 7. Methods used by participants for determining pesticides

Lab Code	Quantification Method								
	Quantification using Standards in solvents or matrix								
Confirmation method									
001	D	S	GC-ECD	0.01	101	15	4	RL (mg/kg)	
	D	M	GC-MS	0.02	107	15	4	Recovery	
002/3	D	M	GC-MS	0.02	107	15	4	Sample Weight (g)	
	D	M	GC-MS/LC-MS	0.05	92	5	Acetone/EtOAc/Hexane	Extraction Solvent	
004	D	M	GC-MS	0.06	95	50	6	Partition Step (if any)	
	D	M	GC-MS	0.06	95	50	O+LL	Clean up step	
005	D	M	GC-MS	0.06	95	50	6	Internal Standard (if any)	
	D	M	GC-MS	0.06	95	50	6	Single level calibration	
006	D	M	GC-MS	92	10	1	O	Multi-level calibration	
	D	M	GC-ITD	0.02	119	25	2	Nº of points	
007	D	M	GC-ITD	0.02	119	25	2	Column type	
	D	M	GC-ITD	0.02	119	25	2	Stationary phase	
008	D	M	GPC	0.25	77	25	2	Determination	
	D	M	GPC	0.25	77	25	2	Determination	
009	D	M	GC-MS	0.02	117	50	4	Determination	
	D	M	GC-EC	0.02	100	3	LL	Determination	
010	D	M	GC-EC	0.02	100	3	LL	Determination	
	ND	S	GC-MS	0.02	25	1	SPE	Determination	
011	ND	S	GC-MS	0.02	25	1	SPE	Determination	

BROMOPROPYLATE

APPENDIX 7. Methods used by participants for determining pesticides

Lab Code		Quantification Method		Confirmation method		RL (mg/kg)		Recovery		Sample Weight (g)		Extraction Solvent		Partition Step (if any)		Clean up step		Internal Standard (if any)		Single level calibration		Multi-level calibration		Nº of points		Column type		Stationary phase		Determination	
012	D	M	GC-MS	0.01	70-110	50	3												Yes	8	HP5MS									GC-MS	
013	D	S	GC-ELCD	0.01	82	25	Ethyl Acetate-Dichloromethane					SPE																	GC-ECD/ELCD		
014	D	S	GC-MS	0.05		40	3		LL										Yes	3	GC-Capillary	DB5	GC-ECD/NPD-MS								
015	D	S	GC-MS	0.01		50	¹ /DCM (80/20)					SPE Florisil							Yes	3	Capillary 0.32	DB5, DB35	GC-ECD								
016	D	S		0.02		50	3	Yes				SPE						Yes	4	A11701/AT1/DB05-MS		GC-ECD/PFPD/NPD/MS									
017	D	S	GC-MS	0.01		40	2					GPC						Yes	5	DB5/DB1701		GC-ECD/PND									
018	D	S	GC-MS	0.005		100	2					GPC	Yes					Yes	5	DB-5/1/1701		GC-ECD/NPD/FPD									
019	D	M	GC-TOF	0.01	97	10	5					PCB 138						Yes	4	DB5		GC-MS									
020	D	M	GC-MS	0.010		25	2					GPC						Yes	3	FSOT	HP-5-MS	GC-MS									
021	D	S	GC-MS	0.01	112.1	50	2					GPC																GC-ECD			
022	D	S		0.01	101	50	6; Acetone	Cyclohexane/Ethyl acetate	GPC	Trans-HCEO	Yes																	GC-ECD			

BROMOPROPYLATE

Lab Code	Quantification Method		Confirmation method		RL (mg/kg)	Recovery	Sample Weight (g)	Extraction Solvent	Partition Step (if any)	Clean up step	Internal Standard (if any)	Single level calibration	Multi-level calibration	Nº of points	Column type	Stationary phase	Determination		
	Quantification using Standards in solvents or matrix																		
023	D	M	GC-MS	0.05	50	2		GPC			Yes	5	30mmx0.25mm	HPS/MS				GC-MS	
024	D	M	GC-MS/ECD	0.02	10		Acetonitril			PCB 209	yes	9	ZP 5	Dimethyl-polysiloxan				GC-MS/ECD	
025	D	M	GC-MS	0.01	50	3		GPC			Yes	3	Capillary-GC	DB1701 (HPG-MS)				GC-ECD/PND/FPD	
026	D	S	GC-MS	0.01	10	5		SPE			Yes	3	Capillary	Varien VF-5MS				GC-ECD GC-NPD GC-MS	
027	D	M	GC-MS		100	2		GPC	EPSILON-HCH				30mmx0.25mmx0.25	DB-5. Factor four				GC-ECD	
028	D	M	GC-MSD	0.01	103	10	5	others	Triphenyl-phosphat		Yes	7	KS	HPS/MS				GC-MS	
029	D	M	GC-MS	0.005	100	2	1	GPC SPE			Yes	3-5	GC	DB5-MS DB17	GC-ECD GC-NPD GC-MS (single-quad)				
030	D	M	GC-ECD	0.02	50	6		GPC	Bromophos-ethyl	Yes			Capillary-GC	DB1				GC-ECD	
031	D	S	GC-MS	0.02	50	2		GPC			Yes	2	Capillary	HP 5				GC-ECD GC-NPD GC-MS	
032	D	M	GC-MS	0.01	95.9	30	Ethyl acetate/Cyclohexane (2:1)	GPC			Yes	4	DB5					GC-ECD, GC-NPD, GC-MS	
033	D	M	GC-ECD	0.02	100	2		GPC			Yes	3	25m x 0.32mmx0.52μm	Ultra2				GC-ECD/NPD	

BROMOPROPYLATE

APPENDIX 7. Methods used by participants for determining pesticides

Lab Code	Quantification Method		Confirmation method		RL (mg/kg)	Recovery	Sample Weight (g)	Extraction Solvent	Partition Step (if any)	Clean up step	Internal Standard (if any)	Single level calibration	Multi-level calibration	Nº of points	Column type	Stationary phase	Determination	
	Quantification using Standards in solvents or matrix																	
034	D	M	GC-MS	0.01	100	2		PCB 153		Yes	2	30mx0.3mmx0.25µm	DB5	DB1701	GC-ECD			
035	D	M	GC-MS	0.02	50	3		GPC		Yes	4	HP-5MS			GC-MS/ECD/FPD			
036	D	S	GC-MS	0.05	101	3		GPC		Yes					GC-ECD/NPD/MS			
037	D	M	GC-MS	0.05	100	2		GPC	TPP/Nitrofen	Yes	4	HP5/DB1701			GC-MS/ECD			
038	D	M	GC-MS	0.01	90	20	2	Triphenyl-phosphat		Yes	3	Capillary-GC	Optimas, DB17-MS		GC-ECD, GC-NPD, GC-MS			
039	D	S	GC-MS	0.05	45	3	LL	GPC	Aldrin	Yes	3	Restek 30x0.25x0.25	RTX-CLPesticide I+II		GC-ECD			
040	D	S	GC-ECD	0.05	340	2		O+GPC	Parathion	Yes		Capillary			GC-ECD			
041	D	S	GC-MS	0.05	128	75	Acetone Followed By Cyclohexan And Ethyl Acetate	GPC		Yes	3	HP5			GC-ECD			
042	D	M	GC-MS	0.01	100	2		GPC	EPN	Yes	3	Capillary	DB5, DB1701, DB35	GC-FCD, GC-NPD, GC-FPD, GC-MS (single-quadr)				
043	D	S	GC-MS	0.01	50	3		LL		Yes		Capillary	DB5		GC-ECD			

BROMOPROPYLATE

Determination									
Stationary phase									
Column type									
Lab Code	Quantification Method	Quantification using Standards in solvents or matrix	Confirmation method	RL (mg/kg)	Recovery	Sample Weight (g)	Extraction Solvent	Partion Step (if any)	Clean up step
044 D M	0.01	100 2	GPC	Yes 5		DB5	GC-MS		
045 D M	GC-ECD. GC-MS	0.02	100 2	GPC	Yes 5		DB5, DB1701	GC-ECD, GC-NPD,GC-MS	
046 D M	GC-ECD	0.005	93 50 3	GPC	Yes		Capillary	DB17, HP5	GC
047 D M	GC-ECD	0.01	100 2	GPC	Yes 3. 5	Capillary	DB5, DB1701	GC-ECD, GC-NPD,GC-FPD	
048 D S	GC-ECD. MS	0.01	7 4	Yes	Yes 3	Capillary	DB5	GC-ECD, GC-MS	
049 D M	GC-MS	0.05	99.5 15 4		2	DB5MS	GC-ECD, GC-IT		
050 NA									
051 D M	GC-ECD	0.01	104 25 6		Yes 5	HP608	GC-ECD		
052 NA									
053 NA									
054 D M	GC-ECD	0.02	110 25 6	Yes O	Yes 3	HP5MS	5%phenyl	GC-ECD	
055 D M	GC-ECD	0.02	121 15 4		Yes 5	HP1701	Me Siloxane	GC-ECD	
056 D M	GC-MS	0.04	93.88 15 4	GPC	Yes 3	DB5/1701		GC-ECD	
057 D S	GC-MS	0.02	20 6	GPC-SPE Yes	Yes 3	DB5	Me Polisiloxan	GC-MS	

BROMOPROPYLATE

APPENDIX 7. Methods used by participants for determining pesticides

Determination									
Stationary phase									
Column type									
Lab Code	Quantification Method	Quantification using Standards in solvents or matrix	Confirmation method	RL (mg/kg)	Recovery	Sample Weight (g)	Extraction Solvent	Partion Step (if any)	Clean up step
058 D M GC-MS 0.05	GPC	Yes	Yes	3	HP5MS	5%PHSi	GC-MS		
059 D S GC-MS 0.05	SPE	Yes	2	Capillary	ZB5				GC-ECD
060 NA									
061 D M GC-MS 0.02	6(Dichloro-methane)	Yes	5	DB5	5%Phenyl-Polysioxane	GC-ECD/NPDMS			
062 D S GC-ECD 0.005	Acetone+ Methanol (1:1)	EMPORE DISK C8 Fendorfors	Yes	3		GC-ECD/NPD			
063 D S GC-ECD 0.01	88.2 25	EXTRELUT 20	Yes		Capillary	DB-608. SPB1	GC-ECD		
064 D S GC-MS	113.8 10	6 O	Yes	Yes	Capillary	HP-5	GC-ECD		
065 D S GC-MS 0.01	Dichloro-methane	Yes		HP5	POLYSILOXANE	GC-ECD			
066 D S GC-MS 0.05	100 25	5 Yes	SPE Yes	2	Capillary	5%PbSil	GC-ECD/NPD		
067 D M GC-MS	10 1	EXTRRELUT	GPC Yes	4					GC-MS
068 D M GC-MS 0.05	91 5	6 1	TFF	5	CIP SIL 8CB	FUSED SILICA	GC-MS/HPLC-FL		

BROMOPROPYLATE

APPENDIX 7. Methods used by participants for determining pesticides

Lab Code	Quantification Method					
	Quantification using Standards in solvents or matrix					
Confirmation method						
RL (mg/kg)						
Recovery						
Sample Weight (g)						
Extraction Solvent						
Partition Step (if any)						
Clean up step						
Internal Standard (if any)						
Single level calibration						
Multi-level calibration						
Nº of points						
Column type						
Stationary phase						
Determination						
069	D	S	GC-MS	83	5	1 ASE (Dionex)
070	D	S	GC-ECD	0.01	15	6 GPC
071	D	M	GC-MS	0.01	15	6 GPC
072	D	S		0.01	70	1 Dciofenten
073	D	S	GC-ECD	0.01	10	Dichloro-methane Dichloro-methane
074	NO RESULTS					
075	D	S	GC-MS	0.085	10	3 SPE Fenchlorphos
076	D	M	GC-MS	0.01	10	5 Yes O
077	D	S	GC-MS	0.02	90	25 GPC
078	D	M	GC-MS	0.05	89	50 GPC
079	D	M	GC-MS	0.05	15	4 Yes 1 HP-1
080	NO RESULTS					
GC-ECD; GC-NPD. GC-MS (SINGLE-QUAD)						DB5
GC-MS						GC-MS
5%Phenyl-Silicone						SE 54; OV1701
GC-ECD/NPD						GC-ECD
5%Phenyl-Siloxane						GC-ECD/NPD
Methyl/Siloxane						DB 5
GC-ECD						GC-ECD

APPENDIX 7. Methods used by participants for determining pesticides

BROMOPROPYLATE

Lab Code	Quantification Method			Confirmation method			RL (mg/kg)	Recovery	Sample Weight (g)	Extraction Solvent	Partition Step (if any)	Clean up step	Internal Standard (if any)	Single level calibration	Multi-level calibration	Nº of points	Column type	Stationary phase	Determination		
	D	M	GC-MS	0.01	20	1															
081	D	M	GC-MS	0.01	20	1															
082	D	M	GC-MS	0.05	15	4															
083	D	M	GC-MS	0.01	15	4	LL phase separation														
084	D	M	GC-MSMS	0.01	10	1															
085	D	M	GC-MS	0.02	102	15															
086	D	M	GC-MSMS	0.05	84	15															
087	D	M	GC-MS	0.03	10	4															
088	D	S	MS	0.03	15	4															
089	NA																				
090	D	M	GC-MSMS	0.05	82.20	15	1	TPP(QC)		Yes	3	Capillary	DB-5ms	GC-MS/MS							
091	D	M	GC-MSMS	0.05	15	1	TIP ⁺ Anthracene		Yes	3	HP-5MS			GC-ITID							
092	D	M	GC-MSMS	0.05	5.2	4			Yes		Factor Four			GC-MS/MS-ECD							

BROMOPROPYLATE

APPENDIX 7. Methods used by participants for determining pesticides

Determination									
Stationary phase									
Column type									
Internal Standard (if any)	Clean up step	Partion Step (if any)	Extraction Solvent	Sample Weight (g)	Recovery	RL (mg/kg)	Quantification using Standards in solvents or matrix	Quantification Method	Lab Code
									093
								Yes	D
							30m/25x25	30m/25x25	S
							CPSIL8CB	CPSIL8CB	GC-MS
									094
								Yes	M
							30m/25x0.2	30m/25x0.2	MS/MS
							CPSIL8	CPSIL8	GC-ECD/GC-NPD/GC-FPD
									095
							5CB.CPSIL19CB.CPSIL8CB	5CB.CPSIL19CB.CPSIL8CB	GC-ECD/GC-FPD/GC-ITD
									096
							25m/0.22mmID	25m/0.22mmID	GC-ITD
							VF-5MS/0.25µm (Varian)	VF-5MS/0.25µm (Varian)	GC-MS
							Capillary	Capillary	097
							DB-5	DB-5	M
							Triphenyl Phosphate	Triphenyl Phosphate	GC-MS
							Capillary	Capillary	098
							DB-5	DB-5	M
							CP-SIL13CB	CP-SIL13CB	GC-MS
									099
							30%Phenyl	30%Phenyl	D
							GC-MS	GC-MS	GC-MS
									100
							DB5	DB5	D
							GC-MS	GC-MS	M
									101
							GC-MS	GC-MS	GC-MS
									102
							At-1701	At-1701	M
							GC-FPD	GC-FPD	GC-ITD
									103
							Attm-1701	Attm-1701	S
									104
							GC-ECD/NPD	GC-ECD/NPD	D
									105
							Narrowbore	Narrowbore	M
							A-T-5MS	A-T-5MS	NA
									106
							GC-MS	GC-MS	D
							0.25mm	0.25mm	GC-MS
							DB-1	DB-1	GC-MS
							Ethyl-parathion	Yes	GC-MS

BROMOPROPYLATE

APPENDIX 7. Methods used by participants for determining pesticides

Lab Code	Quantification Method		Confirmation method		RL (mg/kg)	Recovery	Sample Weight (g)	Extraction Solvent	Partition Step (if any)	Clean up step	Internal Standard (if any)	Single level calibration	Multi-level calibration	Nº of points	Column type	Stationary phase	Determination		
	Quantification using Standards in solvents or matrix																		
107 D M GC-MS 0.05 90.30 20 4																			
108 D S GC-ECD 0.05 92 5 3																			
109 D S GC-EC 0 89 100 3																			
110 D M GC-MS 0.02 89.60 20 3																			
111 D S 0.05 82.90 10 4																			
112 D S 0.05 102 100 3																			
113 D M GC-ITD 0.05 88 15 4																			
114 D S GC-ECD 0.03 18 1																			
115 D S GC/MS 0.05 50 4	Dichloromethane	LL		Yes															
116 D S GC/MS 0.01 99 50 2		SPE		Yes															
117 D M GC-MS 0.05 108 20 4	Yes	GPC		Yes															
118 D M GC-MS 0.02 93 10 6		SPE	Yes	Yes															
119 D M GC-MS 0.01 10 1		NO	Yes	Yes															

BROMOPROPYLATE

Determination									
Stationary phase									
Column type									
Nº of points	Single level calibration	Internal Standard (if any)	Clean up step	Partion Step (if any)	Extraction Solvent	Sample Weight (g)	Recovery	RL (mg/kg)	Quantification using Standards in solvents or matrix
Determination	Stationary phase	Column type	Internal Standard (if any)	Clean up step	Partion Step (if any)	Extraction Solvent	Sample Weight (g)	Recovery	Quantification Method
Lab Code	Lab Code	Lab Code	Lab Code	Lab Code	Lab Code	Lab Code	Lab Code	Lab Code	Lab Code
120	NA								
121	D	M	GC-MS	0.005	25	1	GPC	Yes	5
122	D	S	GC-MS	0.01	83.40	30	1	GPC	Yes
123	D	S	GC-MS	0.1		30	1	GPC	Yes
124	D	S		0.05		30	1	GPC	Yes
125	D	S	GC-MS	0.05	84	25	1	GPC	Yes
126	NO RESULTS								
127	D	S	GC-MS	0.001	30	1	GPC	Yes	3
128	D	S	GC-ECD	0.02	116	30	1	GPC	Yes
129	D	S		0.05		30	1	GPC	Yes
130	D	M		0.05		30	1	GPC	Yes

CHLOROTHALONIL

APPENDIX 7. Methods used by participants for determining pesticides

Lab Code	Quantification Method													
	Confirmation method													
RL (mg/kg)	Recovery													
	Sample Weight (g)													
Extraction Solvent	Partion Step (if any)													
	Clean up step													
Internal Standard (if any)	Single level calibration													
	Multi-level calibration													
Nº of points	Column type													
	Stationary phase													
Determination	Participation													
	001	D	S	GC-MS	0.01	90	15	4	Yes	3-4	DB5 DB 1701	ECD NPD MS		
002/3	D	M	GC-MS	0.01	105	15	4		PCB97	Yes	3 60x0.25x0.1 60x0.25x0.15	DB5 DB1701	GC-ECD	
004	D	S	GC-MS	0.05	100	20	6		Nitrofen +Mifen	Yes	5 CAP	Resitele CLP	GC-ECD	
005	D	M	GC-MS	0.03	100	50	6	O+LL		Yes	3 15x0.32x0.25	DB170	GC-ECD GC-NPD	
006	NA													
007	D	M	GC-TID	0.02	107	25	2		GPC	Dithalimphos	Yes	5 J&W 0.32x0.25mmID	DB5, DB1701	GC-ECD, GC-NPD, GC-ITD
008	D	M		0.01	65	25	2		GPC		Yes	5 DB	1701/05/01/35	GC-ECD/NPD/ITD
009	D	M	GC-MS	0.02	93.0	50	4	Yes	LL	Aldrin Dithalimphos	Yes		HP-5 DB-1701P	NPD ECD
010	D	M	GC-EC	0.02	100	3	LL	No	Aldrin		Yes	3 Capillary	DB5 DB1701	GC-ECD GC-NPD
011	D	S	GC-MS	0.01	60	25	1		SPE		Yes	4 30mx0.53mmx1.5µM	DB5, DB608	GC-ECD, GC-MS
012	D	M	GC-MS	0.02	70-110	50	3			Yes	8 HP5MS			GC-MS

CHLOROTHALONIL

APPENDIX 7. Methods used by participants for determining pesticides

Determination									
Stationary phase									
Column type									
Partion Step (if any)	Clean up step	Internal Standard (if any)	Single level calibration	Multi-level calibration	Nº of points	Column type	Stationary phase	Determination	
Extraction Solvent	Sample Weight (g)	Recovery	RL (mg/kg)						
Quantification using Standards in solvents or matrix	Confirmation method								
Lab Code	Quantification Method								
013 D S GC-ELCD 0.01 80 25 ethyl acetate-dichloromethane	SPE								GC-ECD/ELCD
014 D S GC-MS 0.02 40 3	LL		Yes	3	GC-Capillary				GC-ECD/NPD-MS
015 D S GC-MS 0.01 50 1,DCM (80/20)	SPE Florisil		Yes	3	Capillary 0.32	DB5, DB35			GC-ECD
016 D S 0.02 50 3 Yes	SPE		Yes	4	AT1701/AT1/DB05-MS				GC-ECD/PFPD/NPD/MS
017 D S GC-MS 0.01 40 2	GPC		Yes	5	DB5/DB1701				GC-ECD/PND
018 D S GC-MS 0.002 100 2	GPC Yes		Yes	5	DB-5/1/1701				GC-ECD/NPD/FPD
019 D M GC-TOF 0.02 70 10 5	PCB138	Yes	5	DB5					GC-MS
020 D M GC-MS 0.010 25 2	GPC	Yes	3	FSOT		HP-5-MS			GC-MS
021 D S GC-MS 0.01 97.5 50 2	GPC	Yes			BD15/35/1301. HP5/01/1701/5MS				GC-ECD
022 D S 0.01 98 50 6; Acetone	Ciclohexane/Ethyl acetate	Trans-HCEO	Yes		Capillary	DB5/210			GC-ECD
023 D S GC-MS 0.01 50 2	GPC	Yes	5	30mx0.25mm	HP5MS				GC-MS
024 ND	0.01								

CHLOROTHALONIL

APPENDIX 7. Methods used by participants for determining pesticides

Lab Code									
Quantification Method									
Quantification using Standards in solvents or matrix									
Confirmation method									
RL (mg/kg)									
Recovery									
Sample Weight (g)									
Extraction Solvent									
Partition Step (if any)									
Clean up step									
Internal Standard (if any)									
Single level calibration									
Multi-level calibration									
Nº of points									
Column type									
Stationary phase									
Determination									
025	D	M	GC-MS	0.01	50	3	GPC	yes	3
026	D	S	GC-MS	0.05	100	2	GPC	yes	3
027	D	M	GC-MS		100	2	GPC	EPSILON-HCH	30mx0.25x0.25
028	D	M	GC-ECD	0.01	65.6	10	others	yes	7
029	D	M	GC-MS	0.005	100	2	Triphenyl-phosphat	yes	3-5
030	D	M	GC-ECD	0.01	50	6	GC SPE	yes	GC
031	D	S	GC-MS	0.01	50	2	GPC	Bromophos-ethyl	Yes
032	D	M	GC-MS	0.01	90.4	30	Ethyl acetate/Cyclohexane (2:1)	GPC	Capillary-GC
033	D	M	GC-ECD	0.01	100	2	GPC	yes	DB1
034	D	M	GC-MS	0.01	100	2	GPC	Paraffin-methyl	yes
035	D	M	GC-MS	0.01	50	3	GPC	yes	4

CHLOROTHALONIL

APPENDIX 7. Methods used by participants for determining pesticides

Determination										Lab Code	
Quantification Method					Confirmation method					Lab Code	
Quantification using Standards in solvents or matrix					RL (mg/kg)						
Confirmation method					Recovery					Lab Code	
RL (mg/kg)					Sample Weight (g)						
Sample Weight (g)					Extraction Solvent						
Extraction Solvent					Partion Step (if any)					Lab Code	
Partion Step (if any)					Clean up step						
Clean up step					Internal Standard (if any)					Lab Code	
Internal Standard (if any)					Single level calibration						
Single level calibration					Multi-level calibration					Lab Code	
Multi-level calibration					Nº of points						
Nº of points					Column type					Lab Code	
Column type					Stationary phase						
Stationary phase					Determination					Lab Code	
Determination					GC-ECD/NPD/MS						
036	D	S	GC-MS	0.01	101	3	GPC	Yes		GC-ECD/NPD/MS	
037	D	M	GC-MS	0.01	100	2	GPC	TPP	Yes	GC-MS	
038	D	M	GC-MS	0.01	84	20	Triphenyl-Phosphat	Yes	4	HP5	
039	D	S	GC-MS	0.01	45	3	LL	Aldrin	yes	Capillary-GC	
040	D	S	GC-ECD	0.01	340	2	O+GPC	Parathion	Yes	Optima5, Optima 17, DB17-MS	
041	D	S	GC-MS	0.01	75	75	Acetone Followed by Cyclohexan and Ethyl Acetate	GPC	Yes	Restek 30x0.25x0.25	
042	D	M	GC-MS	0.01	100	2	GPC	EPN	Yes	RTX-CLPesticide I+II	
043	D	S	GC-MS	0.004	50	3	GPC	Capillary	3	GC-ECD	
044	D	M		0.03	100	2	GPC	Capillary	5	DB5, DB1701, DB35	
045	D	M	GC-ECD, GC-NPD, GC-MS	0.01	100	2	GPC	Capillary	5	GC-ECD, GC-FPD, GC-MS (single-quadrupole)	
046	D	M	GC-ECD	0.005	101	50	GPC	Capillary	5	DB5, DB1701	
							GC	GC-MS		GC-MS	

CHLOROTHALONIL

Determination									
Stationary phase									
Column type									
Partion Step (if any)	Clean up step	Internal Standard (if any)	Single level calibration	Multi-level calibration	Nº of points	Sample Weight (g)	Extraction Solvent	Recovery	RL (mg/kg)
Lab Code	Quantification Method	Quantification using Standards in solvents or matrix	Confirmation method						
047 D M GC-ECD 0.005	100	3	GPC	Yes	4	Capillary	DB5, DB1701	GC-ECD, GC-NPD	
048 D S GC-ECD. MS 0.01	7	4	Yes	Yes	3	Capillary	DB5	GC-ECD, GC-MS	
049 D M GC-MS 0.01	96	15	4		2		DB5MS	GC-ECD, GC-IT	
050 NA									
051 D M GC-ECD 0.01	64	25	6		Yes	5	HP608	GC-ECD	
052 NA									
053 NA									
054 D M GC-ECD 0.02	25	25	6	Yes	○	Yes	HP5MS	5%phenyl	GC-ECD
055 D M GC-ECD 0.2	132	15	4			Yes	5	HP1701	Me Siloxane
056 D M GC-MS 0.02	103.82	15	4			Yes	3	DB5/1701	GC-ECD
057 D S GC-MS 0.02	20	6		GPC-SPE Yes		Yes	3		GC-MS
058 D M GC-MS 0.03	15	1		GPC Yes		Yes	3	DB5	Me Polisioxan
059 D S GC-MS 0.01	5	1		SPE		Yes	3	HP5MS	GC-MS
060 NA									
061 D M GC-MS 0.02	98.8	15	6(Dichloro-methane)		Yes	5	DB5	5%Phenyl-Dimethyl-Polysioxane	GC-ECD/NPD/MS

CHLOROTHALONIL

APPENDIX 7. Methods used by participants for determining pesticides

Determination										Lab Code		
Quantification Method					Confirmation method							
Quantification using Standards in solvents or matrix					RL (mg/kg)					Determination		
Confirmation method					Recovery							
RL (mg/kg)					Sample Weight (g)							
Sample Weight (g)					Extraction Solvent							
Extraction Solvent					Partion Step (if any)							
Partion Step (if any)					Clean up step							
Clean up step					Internal Standard (if any)							
Internal Standard (if any)					Single level calibration							
Single level calibration					Multi-level calibration							
Multi-level calibration					Nº of points							
Nº of points					Column type							
Column type					Stationary phase							
Stationary phase					Determination							
062	D	S	GC-ECD	0.005	72	50	Acetone+ Methanol (1:1)	Empore Disk C8	Fenclofors	Yes	3	GC-ECD/NPD
063	D	S	GC-ECD	0.01	95	25	3	EXTRELUT 20		Yes		GC-ECD
064	D	S	GC-MS		105.2	10	6	O	Yes	Yes		GC-ECD
065	D	S	GC-MS	0.005	98.6	10	Dichloromethane			Yes		GC-ECD
066	D	S	GC-MS	0.2	60	25	5	Yes	SPE	Yes	2	HP5
067	D	M	GC-MS			10	1	EXTRELUT	GPC	Yes	4	POLYSILOXANE
068	D	M	GC-MS	0.02	89.2	5	6	1	TFF	Yes	5	5%PbSII
069	D	S	GC-MS	80	5	1	ASE(Dionex)	GPC	Ronnell+ Azobencene+ Triphenyl phosphate	MDN-5S	GC-ECD/NPD/FPD	
070	D	S	GC-ECD	0.01	15	6		GPC	M-series	Yes	25m ID 0.32 mm 0.25um	SE 54; OV1701
071	D	M	GC-MS	0.01	15	6		GPC		Yes	2	SE54. OV1701
072	D	S		0.01	75	10	1	GPC	Diclofenit	Yes		GC-ECD/NPD

APPENDIX 7. Methods used by participants for determining pesticides

CHLOROTHALONIL

Lab Code	Quantification Method														
	Quantification using Standards in solvents or matrix					Confirmation method									
074	RL (mg/kg)														
073	Recovery														
074	Sample Weight (g)														
074	Extraction Solvent														
074	Partition Step (if any)														
074	Clean up step														
074	Internal Standard (if any)														
074	Single level calibration														
074	Multi-level calibration														
080	Nº of points														
080	Column type														
080	Stationary phase														
080	Determination														
081	D	M	GC-MS	0.01	20	1			Yes	3-6					
082	D	M	GC-MS	0.01	15	4			Yes	HP-5MS					
083	D	M	GC-MS	0.05	15	4	LL phase separation		Capillary	5% Ph. Me S					
084	D	M	GC-MSMS	0.01	10	1			Yes	Varian CP 3800					
085	D	M	GC-MS	0.02	80%	15			Yes	Low bleed fused silica					
									4	GC-ECD					
										GC-MS					
										GC-MSMS					
										GC-MS/5%					

CHLOROTHALONIL

Determination									
Stationary phase									
Column type									
Partion Step (if any)	Clean up step	Internal Standard (if any)	Nº of points	Single level calibration	Multi-level calibration	Column type	Stationary phase	Determination	Lab Code
Extraction Solvent	Partion Step (if any)	Internal Standard (if any)	Sample Weight (g)	Recovery	RL (mg/kg)	Column type	Stationary phase	Determination	Quantification Method
Sample Weight (g)	Recovery	RL (mg/kg)	Partion Step (if any)	Extraction Solvent	Sample Weight (g)	Column type	Stationary phase	Determination	Quantification using Standards in solvents or matrix
Partion Step (if any)	Extraction Solvent	Sample Weight (g)	Recovery	Partion Step (if any)	Extraction Solvent	Column type	Stationary phase	Determination	Confirmation method
086 D M GC-MS/MS 0.01	72	15	4			Yes	4	30x25x25	DB-5,DB-17
087 D M GC-MS 0.01	10	4				Yes	3	HP-5	GC-FPD/EC/CD/MS
088 D S MS 0.03	15	4				Yes	5	NarrowBore	GC-MS
089 D M GC-MS 0.05	15					Yes	4	Capillary	BP5
090 D M GC-MS 0.01	75	15	1	TPP(QC) TIP. Anthracene	Yes	1	Capillary	DB-5ms/DB-1701	GC-ECD/PFPD
091 D M GC-MS/MS 0.05	15	1			Yes	3	HP-5MS		GC-ITD
092 D M GC-MS/MS 0.01	5.2	4			Yes		Factor Four		GC-MS/MS-ECD
093 ND S GC-MS 0.01	15	4				Yes	4	30m/25x25	CPSIL8CB
094 D M MS/MS	15	4				Yes	4	30mx0.25x0.2	CPSIL8
095 D M GC-ECD 0.02	85	75	1	GPC	Yes	Yes	3	5CB,CPSIL19CB,CPSI L8CB	GC-ECD,GC-NPD,GC-PFPD
096 D S GC-ITD 0.01	109	15	4		(Injection only)	Yes	5	Capillary 25mx0.22mm ID	BPX-5.0,25mm(SGE)
									GC-ECD

CHLOROTHALONIL

APPENDIX 7. Methods used by participants for determining pesticides

	Lab Code		Quantification Method	Confirmation method	RL (mg/kg)	Recovery	Sample Weight (g)	Extraction Solvent	Partition Step (if any)	Clean up step	Internal Standard (if any)	Single level calibration	Multi-level calibration	Nº of points	Column type	Stationary phase	Determination	
	Method	Sample Type																
097	D	M	GC-MS	0.01	69	30	1			GPC	Triphenyl Phosphate	Yes	5	Capillary	DB-5	GC-MS		
098	D	M	GC-MS	0.01		10	1	SPE	Yes					CP-SIL13CB		GC-MS		
099	D	M	GC-MS	0.01	81	30	1	O				Yes	4	ZB5	30%Phenyl	GC-MS		
100	D	M	GC-MS	0.01	106	10	1	GPC				Yes	3		DB5	GC-MS		
101	D	M	GC-MS	0.01	89	50	5	SPE	Yes					HP 5/HP 50	See column type	GC-ECD/GC-NPD		
102	D	M	GC-ITD	0.01	94	50	1	GPC	Yes					At-1701	Attn-1701	GC-FPD		
103	D	S	GC-MS	0.02	107	20	3	LL				Yes	2	HP-5		GC-ECD/NPD		
104	D	M	GC-MS	0.02	137	75	1	1/20	GPC	Aldrin		Yes	5	Narrowbore	AT-5/M/S	GC-MS		
105	NA																	
106	D	M	GC-MS	0.01	7.5	1	Ethyl-Parathion	Yes						DB-1	0.25mm	GC-ECD/M/S		
107	D	M	GC-MS	0.01	80.30	20	4					Yes	4	Capillary 30 m	HP-5	GC-ECD/NPD		
108	D	S	GC-ECD	0.01	102	5	3					Yes	3	HP-5	5% Phenyl Methyl Siloxane	GC-ECD		
109	D	S	GC-EC	0	112%	100	3					Yes	3	HP-1	Me-SiOH	GC-ECD		
110	D	M	GC-MS	0.01	100.80	20	3	SPE				Yes	5	DB-5		GC-ECD/GC-NPD		
111	D	M		0.02	94.80	10	4	(Silica)				Yes	5	HP-5MS	5%Phenyl	GC-ECD/NPD		
112	D	S	GC-NPD	0.01	90	100	3	O	Yes					HP1	MeSilicone	GC-ECD		
113	D	M	GC-ITD	0.01	91	15	4	NO	Hexachloro-benzene			Yes	5	DB-5MS		GC-ITD		

APPENDIX 7. Methods used by participants for determining pesticides

CHLOROTHALONIL

Lab Code									
Quantification Method									
Quantification using Standards in solvents or matrix									
Confirmation method									
RL (mg/kg)									
Recovery									
Sample Weight (g)									
Extraction Solvent									
Partition Step (if any)									
Clean up step									
Internal Standard (if any)									
Single level calibration									
Multi-level calibration									
Nº of points									
Column type									
Stationary phase									
Determination									
114	D	S	GC-ECD	0.02	18	1	GPC	Yes	4
115	ND		GC/MS	0.01	50	4	Dichloromethane	Yes	HP5
116	NA								GC-ECD
117	NA								GC/MS
118	D	M	GC-MS	0.02	100	10	6	SPE	Yes
119	D	M	GC-MS	0.005	10	1		NO	Yes
120	D	M		0.01	50	1	GPC		Capillary
121	D	M	GC-MS	0.005	25	1	GPC	Yes	DB-35MS
122	D	S	GC-MS	0.01	94.60	30	1	GPC	Yes
123	D	S	HPLC	0.01	30	1		Chrompack 50mx0.25mm x0.25mm	35%Phenyl
124	D	S		0.01	30	1	GPC	Yes	DB-5MS
125	D	S	GC-MS	0.01	91	25	1	GPC	GC-MS
									GC-MS(single quad)ECD, NPD
									GC-ECD
									GC-ECD
									GC-ECD
									GC-ECD
									RTX 17025

APPENDIX 7. Methods used by participants for determining pesticides

CHLOROTHALONIL

		Lab Code									
		Quantification Method									
		Quantification using Standards in solvents or matrix									
		Confirmation method									
		RL (mg/kg)									
		Recovery									
		Sample Weight (g)									
		Extraction Solvent									
		Partition Step (if any)									
		Clean up step									
		Internal Standard (if any)									
		Single level calibration									
		Multi-level calibration									
		Nº of points									
		Column type									
		Stationary phase									
		Determination									
		NO RESULTS									
126											
127	D	S	0.005	30	1	GPC	Yes		30mx0.25mm x0.25um	RTX-5Sil MS/Integria Guard	GC-ECD
128	D	S	GC-ECD	0.05	110	30	1		30mx0.25mm x0.25um	RTX-CLP	GC-NPD
129	D	S		0.01		GPC		Yes	3	Restek	GC-ECD
130	D	M	GC-MS	0.01	30	1	Yes	3	Restek	RTX-5MS	GC-ECD

APPENDIX 7. Methods used by participants for determining pesticides

DIAZINON

Lab Code	Quantification Method						Determination
	Confirmation method						
001	D	S	GC-ECD	0.01	86	15	RL (mg/kg)
002/0 03	D	M	GC-MS	0.02	91	25	Recovery
004	D	M	GC-PFPD	0.02	119	5	Sample Weight (g)
005	D	M	GC-MS	0.01	90	50	Extraction Solvent
006	D	M	GC-MS		10	1	Partition Step (if any)
007	D	M	GC-ITD	0.02	102	25	Clean up step
008	D	M	GC-ITD	0.02	102	25	Internal Standard (if any)
009	D	M	GC-MS	0.01	95,0	50	Single level calibration
010	D	M	GC-NPD	0.03	100	3	Multi-level calibration
011	D	S	GC-MS	0.02	120	25	Nº of points
							Column type
							Stationary phase
							Determination

APPENDIX 7. Methods used by participants for determining pesticides

DIAZINON

Lab Code		Quantification Method		Confirmation method		RL (mg/kg)		Recovery		Sample Weight (g)		Extraction Solvent		Partition Step (if any)		Clean up step		Internal Standard (if any)		Single level calibration		Multi-level calibration		Nº of points		Column type		Stationary phase		Determination	
012	D	M	GC-MS	0.01	70-110	50	3			Yes	8	HP5MS																		GC-MS	
013	D	S	GC-PFPD	0.01	88	25	Ethyl Acetate-Dichloromethane																						GC-FPD/NPD GC-MS		
014	D	S	GC-MS	0.02		40	3		L								Yes	3	GC-Capillary	DB5									GC-ECD/NPD-MS		
015	D	S	GC-MS	0.01		50	1,1DCM (80/20)			Yes	Yes																	GC-FPD, GC-TSD			
016	D	S		0.02		50	3	Yes	SPME			Yes																GC-ECD/PFPD/NPD/MS			
017	D	S	GC-MS	0.01		40	2		GPC			Yes																GC-ECD/PND			
018	D	S	GC-MS	0.013		100	2		GPC	Yes	Yes	5	AT1701/AT1/DB05-MS														DB5/DB1701				
019	D	M	GC-TOF	0.01	100	10	5		PCB 138		Yes	04-may															GC-ECD/NPD/FPD				
020	D	M	GC-MS	0.010		25	2		GPC		Yes	3	DB5														GC-MS				
021	D	S	GC-MS	0.02	97.6	50	2		GPC		Yes															BD1/5/35/1301. HP50/1701/5MS	GC-NPD				
022	D	S		0.01	10 ⁻¹	50	6; Acetone	Cyclohexane/Ethyl acetate	GPC	Trans-HCEO	Yes															Capillary	DB5/210	GC-ECD			
023	D	M	GC-MS	0.02		50	2		GPC		Yes	5	30mmx0.25mm														HP5MS	GC-MS			

APPENDIX 7. Methods used by participants for determining pesticides

DIAZINON

Determination									
Stationary phase									
Column type									
Partion Step (if any)	Clean up step	Internal Standard (if any)	Single level calibration	Multi-level calibration	Nº of points	Column type	Stationary phase	Determination	
Extraction Solvent									
Sample Weight (g)									
Recovery									
RL (mg/kg)									
Confirmation method									
Quantification using Standards in solvents or matrix									
Quantification Method									
Lab Code									
024	D	M	GC-MS/ECD	0.04	10	Acetonitril	PCB 209	Yes	9
025	D	M	GC/MS	0.02	50	3	GPC	Yes	3
026	D	S	GC-MS	0.01	10	5	SPE	Yes	3
027	D	M	GC-MS	0.02	100	2	GPC	EPSILON-HCH	Varian VF-5MS
028	D	M	GC-MSD	0.01	102	10	others	30mx0.25x0.25	DB-5. Factor four
029	D	M	GC-MS	0.005	100	2	Triphenyl-phosphat	GC	GC-ECD
030	D	M	GC-NPD	0.02	50	6	GPC SPE	HP5-MS DB17	HPMS
031	D	S	GC-MS	0.01	50	2	Bromophos-ethyl	DB5	GC-MS (single-quad)
032	D	M	GC-MS	0.01	105.9	30	Ethyl acetate/Ciclohexane (2:1)	ZB5	GC-ECD GC-NPD GC-MS
033	D	M	GC-ECD	0.01	100	2	GPC	DB5	GC-ECD GC-NPD GC-MS
							25m x 0.32mmx0.52μm	Ultra2	GC-ECD/NPD

APPENDIX 7. Methods used by participants for determining pesticides

DIAZINON

Lab Code		Quantification Method		Confirmation method		RL (mg/kg)		Recovery		Sample Weight (g)		Extraction Solvent		Partition Step (if any)		Clean up step		Internal Standard (if any)		Single level calibration		Multi-level calibration		Nº of points		Column type		Stationary phase		Determination	
034	D	M	GC-MS	0.01	100	2		GPC	Parathion-methyl	Yes				30mx0.3mmx0.5μm		DB5	DB17			GC-PND											
035	D	M	GC-MS	0.01	50	3		GPC			Yes	4	HP-5MS							GC-MS/ECD/FPD											
036	D	S	GC-MS	0.02	101	3		GPC		Yes									GC-ECD/NPD/MS												
037	D	M	GC-MS	0.02	100	2		GPC	TPP		Yes	4	HP5/HP1		HP1(Me Siloxan)				GC-MS/FPD												
038	D	M	GC-MS	0.02	88	20	2		Triphenyl-phosphat		Yes	3	Capillary-GC		Optimas, DB17-MS				GC-ECD, GC-NPD, GC-MS												
039	D	S	GC-MS	0.02	45	3	LL	GPC	Parathion		Yes	3	Restex 30x0.25x0.25		Rtx-CLPesticides I+II				GC-NPD												
040	D	S	GC-ECD	0.02	340	2		O+GPC	Parathion	Yes			Capillary						GC-ECD												
041	D	S	GC-MS	0.02	103	75	Acetone Followed by Cyclohexan and Ethyl Acetate	GPC		Yes	3	HP5							GC-ECD, GC-NPD												
042	D	M	GC-MS	0.01	100	2		GPC	EPN	Yes	3	Capillary		DB5, DB1701, DB35				GC-ECD, GC-NPD, GC-FPD, GC-MS (single-quad)													
043	D	S	GC-MS	0.02	50	3		LL		Yes			Capillary		DB5				GC-ECD												

APPENDIX 7. Methods used by participants for determining pesticides

DIAZINON

Determination									
Stationary phase									
Column type									
Partion Step (if any)	Clean up step	Internal Standard (if any)	Single level calibration	Multi-level calibration	Nº of points	Column type	Stationary phase	Determination	
Extraction Solvent	Partion Step (if any)	Internal Standard (if any)	Single level calibration	Multi-level calibration	Nº of points	Column type	Stationary phase	Determination	
Sample Weight (g)	Extraction Solvent	Internal Standard (if any)	Single level calibration	Multi-level calibration	Nº of points	Column type	Stationary phase	Determination	
Recovery	Sample Weight (g)	Internal Standard (if any)	Single level calibration	Multi-level calibration	Nº of points	Column type	Stationary phase	Determination	
RL (mg/kg)	Extraction Solvent	Internal Standard (if any)	Single level calibration	Multi-level calibration	Nº of points	Column type	Stationary phase	Determination	
Confirmation method	Partion Step (if any)	Internal Standard (if any)	Single level calibration	Multi-level calibration	Nº of points	Column type	Stationary phase	Determination	
Quantification using Standards in solvents or matrix	Clean up step	Internal Standard (if any)	Single level calibration	Multi-level calibration	Nº of points	Column type	Stationary phase	Determination	
Quantification Method	Partion Step (if any)	Internal Standard (if any)	Single level calibration	Multi-level calibration	Nº of points	Column type	Stationary phase	Determination	
Lab Code	Clean up step	Internal Standard (if any)	Single level calibration	Multi-level calibration	Nº of points	Column type	Stationary phase	Determination	
044 D M GC-MS 0.01	100 2	GPC	Yes	5	OV17, DB6	GC-NPD			
045 D M GC-ECD, GC-NPD, GC-MS 0.01	100 2	GPC	Yes	5	DB5, DB1701	GC-ECD, GC-NPD, GC-MS			
046 D M GC-ECD 0.02	101 50 3	GPC	Yes	3	Capillary	DB17, HP5	GC		
047 D M GC-FPD 0.01	100 2	GPC	Yes	3 . 5	Capillary	DB5, DB1701	GC-ECD, GC-NPD, GC-FPD		
048 D S GC-FPD,MS 0.01	50 1	GPC	Yes	3	Capillary	DB1701	GC-FPD, GC-MS		
049 D M GC-MS 0.02	83.3 15 4	GPC	Yes	2	DB5MS	GC-ECD, GC-IT			
050 D M GC-NPD 0.02	120 15 4	GPC	Yes	6	Capillary	DB1701	GC-NPD		
051 D M GC-TSD-VA17 0.01	103 25 6	GPC	Yes	5	HP608	GC-ECD			
052 D M GC-ECD 0.02	87.1 50 1	GPC	Yes	5	HP5MS	GC-NPD			
053 NA									
054 D M GC-NPD 0.02	90.3 25 1	GPC	Yes	3	DB1701	14%Cyanopropyl Phenyl	GC-NPD		
055 D M GC-ECD 0.02	101 15 4	GPC	Yes	5	HP1701	Me Siloxane	GC-ECD		
056 D M GC-MS 0.05	99.05 15 4	GPC	Yes	3	DB5/1701		GC-FPD		
057 D S GC-MS 0.02	20 6	GPC-SPE	Yes	3	DB5	Me Polisiloxan	GC-MS		

APPENDIX 7. Methods used by participants for determining pesticides

DIAZINON

Determination									
Stationary phase									
Column type									
Partion Step (if any)	Clean up step	Internal Standard (if any)	Nº of points	Column type	Stationary phase	Determination			
Extraction Solvent									
Sample Weight (g)									
Recovery									
RL (mg/kg)									
Confirmation method									
Quantification using Standards in solvents or matrix									
Quantification Method									
Lab Code									
058	D	M	GC-MS	0.05	15	1	GPC	Yes	Yes
059	D	S	GC-MS	0.02	5	1	SPE	Yes	2
060	D	S	GC-NPD	0.03	87.2	10	6(Dichloromethane)	Yes	03/04/2 07
061	D	M	GC-MS	0.02	105	15	6(Dichloromethane)	Yes	5
062	D	S	GC-NPD	0.005	75	50	Acetone+ Methanol (1:1)	Empore Disk C8	Fendorfors
063	D	S	GC-ECD	0.01	95	25	3	EXTRELUT 20	Yes
064	D	S	GC-MS		80.8	10	6	O	Yes
065	D	S	GC-MS	0.005	89.7	10	Dichloromethane	Yes	HP5
066	D	S	GC-MS	0.05	95	25	5	Yes	POLYSILOXANE
067	D	M	GC-MS		10	1	EXTRELUT	GPC	GC-NPD
068	D	M	GC-MS	0.02	89.2	5	6	1	TFF
							Yes	5	C/P SIL 8CB
									FUSED SILICA
									GC-MS/HPLC-FL

APPENDIX 7. Methods used by participants for determining pesticides

DIAZINON

Determination										
Stationary phase										
Column type										
Nº of points										
Single level calibration										
Multi-level calibration										
Clean up step										
Internal Standard (if any)										
Extraction Solvent										
Sample Weight (g)										
Recovery										
RL (mg/kg)										
Quantification Method										
Quantification using Standards in solvents or matrix										
Confirmation method										
NO RESULTS										
069	D	S	GC-MS	83	5	1	ASE(Dionex)	GPC	Ronnel+Azobe noene+ Triphenyl phosphate	Yes
070	D	S	GC-NPD	0.01	15	6		GPC	M-series	Yes
071	D	M	GC-MS	0.01	15	6		GPC		
072	NA									
073	D	S	GC-ECD	0.01	10	Dichloromethane				Yes
074										3
075	D	S	GC-MS	0.02	10	3		SP-E	Fenchlorphos	Yes
076	D	M	GC-MS	0.01	10	5	Yes	O		
077	D	S	GC-MS	0.02	92	25	2	GPC		Yes
078	D	M	GC-MS	0.02	99	50	1	GPC		Yes
079	D	M	GC-MS	0.02	15	4			Narrowbore	4
080								SPB-1701		1
NO RESULTS										
GC-ECD; GC-NPD. GC-MS (SINGLE-QUAD)										
GC-MS										
GC-FPD										
GC-PFPD										
GC-NPD										

APPENDIX 7. Methods used by participants for determining pesticides

DIAZINON									
Lab Code									
Quantification Method									
Quantification using Standards in solvents or matrix									
Confirmation method									
RL (mg/kg)									
Recovery									
Sample Weight (g)									
Extraction Solvent									
Partition Step (if any)									
Clean up step									
Internal Standard (if any)									
Single level calibration									
Multi-level calibration									
Nº of points									
Column type									
Stationary phase									
Determination									
081	D	M	GC-MS	0.01	20	1		Yes	3-6
082	D	M	GC-MS	0.02	15	4		Yes	Capillary
083	D	M	GC-MS	0.01	15	4	LL phase separation	Yes	HP-5MS
084	D	M	GC-MSMS	0.01	10	1		Yes	Varian CP 3800
085	D	M	GC-MS	0.02	95	15		Yes	Low bleed fused silica
086	D	M	GC-MSMS	0.01	79	15		Yes	GC-MS/MS
087	D	M	GC-MS	0.01	10	4		Yes	CP-Sil 8
088	D	MS			15	4		Yes	PhMSil 5%
089	D	M	GC-MS/MS	0.05	15			Yes	DB-5, DB-17
090	D	M	GC-MSMS	0.02	89.30	15		Yes	GC/FPD/ECM/MS
091	D	M	GC-MSMS	0.05	15	1	TIP(QC)	Yes	GC/NPD/FPD/MS
092	D	M	GC-MSMS	0.02	5.2	4	TIP ^P , Anthracene	Yes	BP5
								Yes	GC-MS/MS
								Yes	GC-MS/MS-ECD
								Yes	GC-ITID
								Yes	Factor Four

APPENDIX 7. Methods used by participants for determining pesticides

DIAZINON

Determination									
Stationary phase									
Column type									
Partion Step (if any)	Clean up step	Internal Standard (if any)	Nº of points	Column type	Stationary phase	Determination			
Extraction Solvent	Partion Step (if any)	Internal Standard (if any)	Single level calibration	Column type	Stationary phase	Determination			
Sample Weight (g)	Extraction Solvent	Internal Standard (if any)	Multi-level calibration	Column type	Stationary phase	Determination			
Recovery	Sample Weight (g)	Internal Standard (if any)	Nº of points	Column type	Stationary phase	Determination			
RL (mg/kg)	Extraction Solvent	Internal Standard (if any)	Single level calibration	Column type	Stationary phase	Determination			
Confirmation method	Partion Step (if any)	Internal Standard (if any)	Multi-level calibration	Column type	Stationary phase	Determination			
Quantification using Standards in solvents or matrix	Partion Step (if any)	Internal Standard (if any)	Nº of points	Column type	Stationary phase	Determination			
Quantification Method	Partion Step (if any)	Internal Standard (if any)	Single level calibration	Column type	Stationary phase	Determination			
Lab Code	Partion Step (if any)	Internal Standard (if any)	Multi-level calibration	Column type	Stationary phase	Determination			
093 D S GC-MS 0.02	15	4	4	30m/25x25	CPSIL8CB	GC-MS			
094 D M MS/MS	15	4	4	30mx0.25x0.2	CPSIL8	GC-ECD,GC-NPD,GC-PFPD			
095 D M GC-FPD	0.02	88	75	1	CP-SIL 5CB,CPSIL,19CB,CPSIL 8CB	GC-ECD,GC-FPD,GC-ITD			
096 D M GC-ITD	0.025	97	15	4	Capillary,25mx0.22mmI D	GC-ITD			
097 D M GC-MS	0.02	92	25	1	O Triphenyl Ethylene	GC-MS			
098 D M GC-MS	0.02	10	1		SPE Yes	GC-MS			
099 D M GC-MS	0.01	91	30	1	O	GC-MS			
100 D M GC-MS	0.02	84	10	1	GPC Yes	GC-MS			
101 D M GC-MS	0.02	87	50	5	SPE Yes	GC-MS			
102 D M GC-ITD	0.01	93	50	1	GPC Yes	GC-MS			
103 D S GC-MS	0.02	90	20	3	LL HP-5	GC-ECD/NPD			
104 D M GC-MS	0.02	111	75	1	1/20 Aldrin	Narrowbore AT-5MS	GC-MS		
105 D M GC-FTD	0.02	25,0 33	6	2	LL Dithalimphos	NB-54 GC-FTD			
106 D M GC-MS	0.02	7,5	1		Ethyl-parathion Yes	DB-1 GC-ECD/MS			

APPENDIX 7. Methods used by participants for determining pesticides

DIAZINON											
Lab Code	Quantification Method										
	Quantification using Standards in solvents or matrix										
Confirmation method	RL (mg/kg)							Recovery			
	Sample Weight (g)							Extraction Solvent			
Partion Step (if any)	Clean up step							Internal Standard (if any)			
	Single level calibration							Multi-level calibration			
Nº of points	Column type							Stationary phase			
	Determination										
107 D M GC-MS 0.02 89.10 20 4 Yes 4 Capillary 30 m HP-5 GC-ECD/NPD											
108 D S GC-ECD 0.02 100 5 3 Yes 3 HP-5 5% Phenyl Methyl Siloxane GC-ECD											
109 D S GC-EC 0 89 100 3 Yes 3 HP-1 Me-SiOH GC-ECD											
110 D M GC-MS 0.01 87 20 3 SPE Yes 5 DB-5 GC-ECD,GC-NPD											
111 D S 0.02 92.80 10 4 SPE (Silica) Yes 5 HP-5MS 5%Phenyl GC-ECD/NPD											
112 D S GC-ECD 0.02 95 100 3 O Yes HP17 50%PhMe Silicone GC-NPD											
113 D M GC-ITD 0.01 91 15 4 Hexachlorobenzene Yes 5 DB-5MS GC-ITD											
114 D M GC-FPD 0.02 18 1 GPC Yes Capillary HP5 GC-FPD											
115 D S GC/MS 0.02 50 4 Dichloromethane LL Yes HP(30x0.25x0.25) 5MS GC/MS											
116 D S GC/MS 0.01 124 50 2 SPE Yes HP5-MS GC/MS											
117 D M GC-MS 0.02 105.20 20 4 Yes GPC Yes 5 HP-5MS GC/MS											
118 D M GC-MS 0.02 96 10 6 SPE Yes DB-35MS 35%Phenyl GC/MS											
119 D M GC-MS 0.02 10 1 NO Yes Capillary DB-5MS GC/MS											

APPENDIX 7. Methods used by participants for determining pesticides

DIAZINON

Determination										
Stationary phase										
Column type										
Partion Step (if any)	Clean up step	Internal Standard (if any)	Single level calibration	Multi-level calibration	Nº of points	Column type	Stationary phase	Determination		
Extraction Solvent										
Sample Weight (g)										
Recovery										
RL (mg/kg)										
Confirmation method										
Quantification using Standards in solvents or matrix	Quantification Method	Lab Code								
120 D M GC-FPD 0.01	50	1	GPC		Yes	2	30mmx0.26mm	DB-5MS	GC-ITD	
121 D M GC-MS 0.03	25	1	GPC		Yes	5	Capillary	DB-5MS	GC-MS(single quad)ECD, NPD	
122 D S GC-MS 0.02	87.70	30	1	GPC	Yes	3	Chrompack 30mmx0.25mm x0.25mm	CPSIL-8CB	GC-NPD	
123 D S GC-MS 0.02	30	1	GPC	Yes			30mmx0.53mm	CP-Sil 8 CB	GC-NPD	
124 D S 0.02	30	1			Yes	3	SGE	BPX-5	GC-NPD	
125 D S GC-MS 0.02	108	25	1	GPC	Yes	5	Capillary	CPSyl 8CB	GC-NPD	
126	NO RESULTS									
127 D S GC-MS 0.001	30	1	GPC		Yes	3	50mmx0.32mm x0.25um	CP-Sil 8 CB	GC-ECD	
128 D S HPLC-DAD 0.02	93	30	1	GPC	Yes	3	30mmx0.25mm x0.25um	RTX-CLP	GC-NPD	
129 D S 0.02	30	1	GPC	Yes	3	Varian	Factor Four V/F	GC-NPD		
130 D M GC-MS 0.02	30	1		Yes	3	Varian	CPSil 8CB	GC-NPD		

APPENDIX 7. Methods used by participants for determining pesticides

DIMETHOATE

Lab Code	Quantification Method						Determination
	Quantification using Standards in solvents or matrix						
Confirmation method	RL (mg/kg)						
	Recovery						
Sample Weight (g)	Extraction Solvent						
	Partition Step (if any)						
Clean up step	Internal Standard (if any)						
	Single level calibration						
Nº of points	Multi-level calibration						
	Column type						
Stationary phase	Determination						
	GC-ECD, GC-NPD, GC-ITD						
001 D M LC-MS-MS 0.01 106 15 4	DB5 DB1701						ECD NPD MS
002 D M GC-MS 0.02 85 25 4	Tributyl-phosphat						GC-FPD
003 D S GC-PFPD 0.02 102 25 Acetone. Petrololumether.	Yes 3 30x0.32x0.25						DB1301
004 D S GC-PFPD 0.02 102 25 Dichloromethane	Yes 6 100x2.1						LC-MS/MS
005 D M GC-MS 0.009 85 20 1	C18 SPE						RTX-WAA
006 NA	Yes 5 90x0.83x0.5						GC-NPD
007 D M GC-NPD 0.01 106 25 2	J&W 0.32/0.25mmID						GC-ECD, GC-NPD, GC-ITD
008 D M	DB5 DB1701						GC-ECD/NPD/ITD
009 D M GC-MS 0.01 108.0 50 4 Yes	1701/05/01/35						GC-ECD/NPD/ITD
010 D M GC-EC 0.05 100 3 LL	GC-ECD/NPD/ITD						GC-ECD/NPD/ITD
011 D S GC-MS 0.02 113 25 1	Yes 3 Capillary						GC-ECD GC-NPD
012 D M GC-MS 0.02 60-70 50 3	Yes 5 30mx0.53mmx1.5μm						GC-NPD, GC-FPD, GC-MS
	DB5, DB608						
	HP5MS						GC-MS

APPENDIX 7. Methods used by participants for determining pesticides

DIMETHOATE

Determination									
Stationary phase									
Column type									
Lab Code	Quantification Method	Quantification using Standards in solvents or matrix	Confirmation method	RL (mg/kg)	Recovery	Sample Weight (g)	Extraction Solvent	Partition Step (if any)	Clean up step
013 D S GC-PFPD 0.01 80 25 Ethyl Acetate-Dichloromethane									GC-FPD/NPD/GC-MS
014 D S GC-MS 0.02 40 3 LL					Yes	3	GC-Capillary		DB5
015 D S GC-MS 0.02 50 1/DCm (80/20)					Yes	Yes	Capillary		GC-ECD-NPD-MS
016 D S 0.02 50 3 Yes	SPE			Yes	4	AT1701/AT1/DB05-MS			GC-FPD, GC-TSD
017 D S GC-MS 0.01 40 2 GPC				Yes	5	DB5/DB1701			GC-ECD/PFPD/NPD/MS
018 D M GC-ECD 0.01 10 5 SPE	Yes			Yes	5	AtlantisDc18			GC-ECD/PND
019 D M 5 Transitions 0.005 90 10 5 Triphenyl-phosphat				Yes	4	ZORBAX C18			LC-MS/MS
020 D M GC-MS 0.010 25 2 GPC				Yes	3	FSOT			HP-5-MS
021 D S GC-MS 0.02 123.8 50 2 GPC				Yes			BD1/5/35/1301. HPS0/1701/5MS		GC-NPD
022 D M 0.01 104 10 6;(Methanol) CEM ELUT				Yes	7	2.1x125mm; 3µm	RP-C18		LC-MS/MS
023 D M LC-MS/MS 0.01 10 Methanol				Yes	5	2.1x100mm	Atlantis C18		LC-MS/MS
024 ND	0.1								

APPENDIX 7. Methods used by participants for determining pesticides

DIMETHOATE

Determination									
Stationary phase									
Column type									
Partion Step (if any)					Clean up step				
Extraction Solvent					Internal Standard (if any)				
Sample Weight (g)					Single level calibration				
Recovery					Multi-level calibration				
RL (mg/kg)					Nº of points				
Confirmation method					Column type				
Quantification using Standards in solvents or matrix					Stationary phase				
Lab Code					Determination				
025	D	M	LC-MS/MS	0.01	20	6		Yes	2x4
026	D	S	GC-MS	0.01	100	2		Yes	3
027	D	M	GC-MS	0.05	100	2		Capillary	Varien VF-5MS
028	D	M	GC-MSD	0.01	90.3	10	5	EPSILON-HCH	30mx0.25x0.25
029	D	M	GC-MS	0.005	100	2	1	Others	DB-5. Factor four
030	D	M	GC-NPD	0.02	50	6		Triphenyl-phosphat	DB-5. Factor four
031	D	S	GC-MS	0.01	50	2		Yes	7
032	D	M	LC-MS/MS	0.01	81.3	10	6(Methanol)	KS	HPSMS
033	D	M	GC-ECD	0.02	100	2		Yes	3.5
034	D	M	GC-MS	0.01	100	2		GPC	GC-ECD
035	D	M	GC-MS	0.02	50	3		Bromophos-ethyl	GC-NPD
									GC-MS
									GC-MS/ECDFPD

APPENDIX 7. Methods used by participants for determining pesticides

DIMETHOATE

Determination									
Stationary phase									
Column type									
Lab Code	Quantification Method	Quantification using Standards in solvents or matrix	Confirmation method	RL (mg/kg)	Recovery	Sample Weight (g)	Extraction Solvent	Partition Step (if any)	Clean up step
036 D S GC-MS 0.02	101 3 GPC	Yes		HP5/HP1	HP1(Me Siloxan)				GC-ECD/NPD/MS
037 D M GC-MS 0.02	100 2 GPC	Yes	4	HP5/HP1	HP1(Me Siloxan)				GC-ECD/NPD/MS
038 D M LC-MS/MS 0.01	92 2 Triphenyl-phosphat	Yes	3	Capillary-GC	Optima5. Optima 17. DB17. MS C18. Aqua5U				GC-ECD/NPD/MS
039 D S 0.01	10 5 PSA	Yes	6	Aqua (phenomex)	C18				GC-MS/FPD
040 D S GC-ECD 0.02	340 2 O+GPC Parathion	Yes		Capillary					GC-MS/FPD
041 D M LC-MS. GC-MS 0.02	80 10 Methanol. Acetone Followed by Cyclohexan and Ethyl Acetate	Yes	7.3	Altanis. HP5	dc18				GC-ECD
042 D M GC-MS 0.01	100 2 GPC EPN	Yes	3	Capillary	DB5. DB1701. DB35				GC-MS/MS
043 D M LC-MS/MS 0.006	10 6 (Methanol) SPE	Yes	2	C18					GC-ECD. GC-NPD. GC-FPD. GC-MS (single-quad)
044 D M 0.02	100 2	Yes	4	RP8					LC-MS/MS
045 D M GC-ECD. GC-NPD. GC-MS 0.02	100 2 GPC	Yes	5	DB5. DB1701					GC-ECD. GC-NPD.GC-MS

APPENDIX 7. Methods used by participants for determining pesticides

DIMETHOATE									
	Lab Code	Quantification Method							
		Quantification using Standards in solvents or matrix							
		Confirmation method							
		RL (mg/kg)							
		Recovery							
		Sample Weight (g)							
		Extraction Solvent							
		Partition Step (if any)							
		Clean up step							
		Internal Standard (if any)							
		Single level calibration							
		Multi-level calibration							
		Nº of points							
		Column type							
		Stationary phase							
		Determination							
046	D	M	GC-ECD	0.01	102	50	3	GPC	Yes
047	D	M	GC-FPD	0.005	100	2		GPC	Yes
048	D	S	GC-FPD/MS	0.02	50	1			3.5
049	D	M	GC-MS	0.02	92	15	4		Capillary
050	D	M	GC-NPD	0.02	140	15	4		DB5, DB1701
051	D	M	GC-TSD	0.02	103	25	1		DB1701
052	D	M	GC-TSD-VA17	0.02	104	50	1		DB5/MS
053	NA								GC-ECD/GC-IT
054	D	M	GC-NPD	0.02	94.5	25	1		GC-NPD
055	NA								GC-NPD
056	D	M	GC-NPD	0.04	92.32	15	4	GPC	Yes
057	D	S	GC-MS	0.05	20	6		GPC-SPE	Yes
058	D	M	GC-MS	0.05	15	1		GPC	Yes
059	ND			0.02					
060	NA								

APPENDIX 7. Methods used by participants for determining pesticides

DIMETHOATE

Determination									
Stationary phase									
Column type									
Lab Code	Quantification Method	Quantification using Standards in solvents or matrix	Confirmation method	RL (mg/kg)	Recovery	Sample Weight (g)	Extraction Solvent	Partion Step (if any)	Clean up step
061 D M GC-MS 0.02	6(Dichloro-methane)	Acetone+ Methanol (1:1)	Empore Disk C8	Fendofos	Yes	5	DB5	5% Phenyl 95% Dimethyl-Polysiloxane	GC-ECD/NPD/MS
062 ND S GC-NPD 0.01	50	EXTRELUt 20	O	Yes	Yes	3	SPPB5	GC-FCD/NPD	GC-FPD
063 D S GC-FPD 0.05	87	25	Capillary	HP-5	Capillary	6	HP5	POLYSILOXANE	GC-ECD
064 D S GC-MS	109.9	10	HP-5	POLYSILOXANE	Yes	0.05	GC-MS	GC-NPD	GC-ECD/NPD
065 D S GC-MS	0.01	79.1	Dichloromethane	GC-MS	Yes	0.05	GC-MS	GC-MS	GC-MS
066 D S GC-MS	0.05	90	25	GC-MS	Yes	5	GC-MS	GC-MS	GC-MS
067 D M GC-MS		10	1	GC-MS	Yes	1	GC-MS	GC-MS	GC-MS
068 D M GC-MS	0.02	98.5	5	GC-MS	Yes	1	GC-MS	GC-MS/HPLC-FL	GC-MS/HPLC-FL
069 D S GC-MS	79	5	1	GC-MS	Ronnel+ Azobencene+ Triphenyl phosphate	MDN-5S	5%PHENYL SILICONE	GC-ECD/NPD/FID	GC-ECD/NPD/FID
070 D S GC-NPD	0.01	15	6	GC-MS	GPC	M-series	25m ID 0.32 mm 0.25um	SE 54; OV1701	GC-ECD/NPD
071 D M GC-MS	0.01	15	6	GC-MS	GPC			SE54. OV1701	GC-NPD/FID

APPENDIX 7. Methods used by participants for determining pesticides

DIMETHOATE

Determination								
Stationary phase								
Column type								
Lab Code	Quantification Method	Quantification using Standards in solvents or matrix	Confirmation method	RL (mg/kg)	Recovery	Sample Weight (g)	Extraction Solvent	Partition Step (if any)
074	NO RESULTS							
075	D	S	GC-MS	0.05	10	3	SPE Fenchloriphos	Yes Yes Yes
076	D	M	GC-MS	0.02	10	5	Yes GPC GPC	O 4 4
077	D	S	GC-MS	0.05	91	25	2 Fused Silica HP-50+	5%Phenyl- 50%Phenyl- MethylSilicone
078	D	M	GC-MS	0.02	82	50	1 GPC	Narrowbore DB 5
079	D	M	GC-NPD	0.02	15	4	Yes Yes	SPB 1701 GC-NPD
080	NO RESULTS							
081	D	M	GC-MS	0.01	20	1	Yes Yes	3-6 HP-5MS
082	D	M	GC-MS	0.02	15	4	Yes LL phase	Capillary Varian CP 3800
083	D	M	GC-MS	0.03	15	4	Yes Yes	Low bleed fused silica GCMs
084	D	M	GC-MS/MS	0.01	10	1	Yes Yes	CP-Sil 8 GC-MS/MS

APPENDIX 7. Methods used by participants for determining pesticides

DIMETHOATE

Determination								
Stationary phase								
Column type								
Lab Code	Quantification Method	Quantification using Standards in solvents or matrix	Confirmation method	RL (mg/kg)	Recovery	Sample Weight (g)	Extraction Solvent	Partition Step (if any)
085 D M GC-MS 0.02	84	15	4	Yes	4	Capillary	PhMSil 5%	
086 D M GC-MS/MS 0.01	75	15	4	Yes	4	30x25x25	DB-5,DB-17	GC/FPD/ECD/MS
087 D M GC-MS 0.02	10	4		Yes	3	HP-5		GC-ECD/NPD/FPD/MS
088 D MS	15	4		Yes	5	NarrowBore	HP5-MS	GC-MS
089 D M GC-MS/MS 0.05	15			Yes	4	Capillary	BP5	GC-MS/MS
090 ND M GC-MS 0.05	83.2	15	1	TIP(QC) TIP ^P Anthracene	Yes	1	DB-5-ms/DB-1701	GC-ECD/PFPD
091 D M GC-MS/MS 0.05	15	1		Yes	3	HP-5MS		GC-ITD
092 D M GC-MS/MS 0.02	5.2	4		Yes	6	Factor Four		GC-MS/MS-ECD
093 D S GC-MS 0.02	15	4		Yes	4	30m/25x25	CPSIL8CB	GC-MS
094 D M MS/MS	15	4		Yes	4	30mx0.25x0.2	CPSIL8	GC-ECD GC-NPD.GC-PFPD
095 D M GC-FPD 0.02	105	75	1	GPC Yes	3	CP-SIL 19CB.CPSIL 8CB		GC-ECD GC-FPD.GC-ITD

APPENDIX 7. Methods used by participants for determining pesticides

DIMETHOATE

Determination									
Stationary phase									
Column type									
Lab Code	Quantification Method	Quantification using Standards in solvents or matrix	Confirmation method	RL (mg/kg)	Recovery	Sample Weight (g)	Extraction Solvent	Partition Step (if any)	Clean up step
096 D S GC-TID 0.01	102 7.5 4(+Na ₂ SO ₄)			Yes 4 150x3.2mmID	Altima C18.5mm	LC-MS/MS			
097 D M GC-FPD/GC-MS 0.02	89 25 1	O GPC	Triphenyl Ethylene Triphenyl Phosphate	Yes 6 Capillary	DB-5 DB-5	GC-MS GC-FPD			
098 D M GC-MS 0.02	10 1	SPE Yes	O	Yes 5 Capillary		GC-MS			
099 D M GC-MS 0.01	118 30 1	O GPC	ZB5	Yes 4 30%Phenyl	DB31301	GC-FPD			
100 D M GC-MS 0.02	93 10 1	SPE	HP 5/HP 50	Yes 3 See column type	GC-ECD/GC-NPD				
101 D M GC-MS 0.02	92 50 5	GPC	At-1701	Yes	GC-FPD				
102 D M GC-TID 0.01	87 50 1	LL	Atm-1701	Yes	GC-FPD				
103 D S GC-MS 0.02	85 20 3	HP-5	GC-ECD/NPD	Yes 2					
104 D M GC-MS 0.02	96 75 1	HP-5MS	GC-MS	Yes 5 Narrowbore					
105 ND	0.02								
106 D M GC-MS 0.02	7.5 1	Ethylparathion	DB-1	Yes 0.25mm	GC-ECDD/MS				
107 D M GC-MS 0.02	112.5 20 4			Yes 4 Capillary 30 m	HP-5	GC-ECDD/NPD			
108 D S GC-ECD 0.02	102 5 3			Yes 3 HP-5	5% Phenyl Methyl Siloxane	GC-ECD			
109 D S GC-EC 0 116 100 3				Yes 3 HP-1	MeSiOH	GC-ECD			
110 D M GC-MS 0.01	93.10 20 3	SPE	DB-5	Yes 5		GC-ECD/GC-NPD			
111 D M 0.01 82.60 10 4	(Silica)	SPE	HP-5MS	Yes 5 5%Phenyl		GC-ECD/NPD			

APPENDIX 7. Methods used by participants for determining pesticides

Determination									
Stationary phase									
Column type									
Partion Step (if any)	Clean up step	Internal Standard (if any)	Single level calibration	Multi-level calibration	Nº of points				
Extraction Solvent	Sample Weight (g)	Recovery							
Confirmation method	RL (mg/kg)	Lab Code	Quantification Method	Quantification using Standards in solvents or matrix	Lab Code	Quantification Method			
							DIMETHOATE		
112 D S GC-ECD 0.02	92	100	3	O	Yes	HP17	50%PhMe Silicone		
113 D M GC-ITD 0.025	90	15	4	NO Hexachlorobenzene	Yes	5	GC-NPD		
114 D M GC-FPD 0.04	18	1		GPC	Yes	Capillary	GC-ITD		
115 ND GC/MS 0.02	50	4	Dichloromethane	LL	Yes	HP(30x0.25x0.25)	HP5		
116 D S GC/MS 0.02	91	50	2	SPE	Yes	5MS	GC-FPD		
117 D M GC-MS 0.02	108	20	4	Yes	GPC	HP5-MS	GC/MS		
118 D M GC-MS 0.02	95	10	6	SPE	Yes	5	GC-MS		
119 D M GC-MS 0.02	10	1		NO	Yes	DB-35MS	GC-MS		
120 ND M GC-FPD 0.01	50	1		GPC	Yes	35%Phenyl	GC-MS		
121 D M GC-MS 0.03	25	1		GPC	Yes	DB-5MS	GC-ITD		
122 D S GC-MS 0.02	102.	30	1	NO	5	Capillary	GC-MS(Single quad)ECD NPD		
123 D S GC-MS 0.02	30	1		GPC	Yes	3	Chromopack 30mx0.32mm x0.25mm	CP-SII 8 CB	
								GC-NPD	

APPENDIX 7. Methods used by participants for determining pesticides

DIMETHOATE

Determination									
Stationary phase									
Column type									
Lab Code	Quantification Method	Quantification using Standards in solvents or matrix	Confirmation method	RL (mg/kg)	Recovery	Sample Weight (g)	Extraction Solvent	Partition Step (if any)	Clean up step
126									
127	D	S	GC-MS	0.005	30	1	GPC	Yes	3
128	D	S	GC-ECD	0.05	104	30	1	GPC	Yes
129	D	S		0.02	30	1	GPC	Yes	3
130	D	M	GC-MS	0.02	30	1		Yes	3

NO RESULTS

APPENDIX 7. Methods used by participants for determining pesticides

ENDOSULFAN

	Lab Code		Quantification Method		Confirmation method		RL (mg/kg)		Recovery		Sample Weight (g)		Extraction Solvent		Partition Step (if any)		Clean up step		Internal Standard (if any)		Single Level Calibration		Multi-Level Calibration		Nº of points		Column type		Stationary phase		Determination	
001	D	S	GC-MS	0.01	96	15	4													Yes	3.4	DB5 DB 1701						ECD NPD MS				
002/ 003	D	M	GC-MS	0.01	107	15	4													PCB97	Yes	3	60x0.25x0.1 60x0.25x0.15	DB5/1701					GC-ECD			
004	D		GC-MS	0.05	102	20	6												Nitrofen+ Mirex	Yes	5	CAP		Restele CLP			GC-ECD					
005	D	M	GC-MS	0.005	95	20	1												SPE	Yes	5	90x0.83x0.5	RTX-WMA				GC-NPD					
006	D	M	GC-MS		100	10	1				O									Yes	5	DB 5MS					MS			GC-MS		
007	D	M	GC-ECD	0.01	82	25	2				GPC								Dithalimphos	Yes	5	J&W 0.32/0.25mmID	DB5 DB1701				GC-ECD, GC-NPD, GC-ITD					
008	D	M		0.01	71	25	2				GPC									Yes	5	DB	1701/05/01/35				GC-ECD/NPD/ITD					
009	D	M	GC-MS	0.01	107.0	50	4	Yes	LL									dithalimphos	Yes							H-P-5 DB-1701P	NPD ECO					
010	D	M	GC-EC	0.03	100	3	LL	no	aldrin										Capillary	Yes	3	Capillary	DB 5 DB1701				GC-ECD GC-NPD					
011	D	S	GC-MS	0.01	75	25	1				SPE									Yes	4	30mx0.53mmx1.5μm	DB5, DB608				GC-ECD, GC-MS					

APPENDIX 7. Methods used by participants for determining pesticides

ENDOSULFAN

Lab Code		Quantification Method		Confirmation method		RL (mg/kg)		Recovery		Sample Weight (g)		Extraction Solvent		Partition Step (if any)		Clean up step		Internal Standard (if any)		Single Level Calibration		Multi-Level Calibration		Nº of points		Column type		Stationary phase		Determination	
012	D	M	GC-MS	0.01	70-110	50	3					Yes	8	HP5MS					GC-MS		GC-ECD/ELCD										
013	D	S	GC-ELCD	0.01	80	25	Ethyl Acetate-Dichloromethane		SPRE									Yes	3	GC-Capillary	DB5	GC-ECD-NPD-MS									
014	D	S	GC-MS	0.05		40	3		LL									Yes	3	Capillary 0.32	DB5, DB35	GC-ECD									
015	D	S	GC-MS	0.01		50	1/DCM (80/20)		SPRE Florisil									Yes	3	AT1701/AT1/DB05-MS	GC-ECD/PFPD/NPD/MS										
016	D	S		0.02		50	3	yes	SPRE			Yes	4																		
017	D	S	GC-MS	0.01		40	2		GPC			Yes	5	DB5/DB1701																	
018	D	S	GC-MS	0.003		100	2		GPC	YES		Yes	5	DB-5/1/1701																	
019	D	M	GC-TOF	0.01	96	10	5		PCB 138			Yes	1	DB5																	
020	D	M	GC-MS	0.010		25	2		GPC		In Matrix	3	FSOT	HP-5-MS																	
021	D	S	GC-MS	0.005	91.7	50	2		GPC			Yes		BD 1/5/35/1301. HPSO/1701/5MS																	
022	D	S		0.01	100	50	6; ACETONE	Cyclohexane-Ethy acetaate	GPC	Trans-HCEO	Yes			Capillary	DB5/210																
023	D	M	GC-MS	0.05		50	2		GPC			Yes	5	30mmx0.25mm	HP5MS																

APPENDIX 7. Methods used by participants for determining pesticides

ENDOSULFAN

Determination									
Stationary phase									
Column type									
Partion Step (if any)	Clean up step	Internal Standard (if any)	Single Level Calibration	Multi-Level Calibration	Nº of points	Column type	Stationary phase	Determination	ENDOSULFAN
Extraction Solvent	Partion Step (if any)	Internal Standard (if any)	Single Level Calibration	Multi-Level Calibration	Nº of points	Column type	Stationary phase	Determination	ENDOSULFAN
Sample Weight (g)	Recovery	RL (mg/kg)	Partion Step (if any)	Clean up step	Internal Standard (if any)	Single Level Calibration	Multi-Level Calibration	Nº of points	Column type
Confirmation method	Quantification using Standards in solvents or matrix	Quantification Method	Extraction Solvent	Partion Step (if any)	Internal Standard (if any)	Single Level Calibration	Multi-Level Calibration	Nº of points	Column type
Lab Code	Quantification Method	Quantification using Standards in solvents or matrix	Confirmation method	Partion Step (if any)	Internal Standard (if any)	Single Level Calibration	Multi-Level Calibration	Nº of points	Column type
024 D M GC-MSECD 0.01	Acetonitril	PCB 209	Yes	9	ZP 5	Dimethyl-Polysiloxan	GC-MSECD		
025 D M GC/MS 0.01			Yes	3	Capillary-GC	DB1701 (HPG-MS)	GC-ECD/PND/FPD		
026 D S GC-ECD 0.01			Yes	3	Capillary	Varien VF-5MS	GC-ECD GC-NPD GC-MS		
027 D M GC-MS 0.05			Yes	7	KS	HPMS	GC-ECD		
028 D M GC-ECD 0.01	98.7	100	Yes	3-5	GC	DB5 HP5-MS DB17	GC-ECD GC-NPD GC-MS (single-quad)		
029 D M GC-MS 0.005			Yes	3-5	GC	DB5 HP5-MS DB17	GC-ECD GC-NPD GC-MS (single-quad)		
030 D M GC-ECD 0.005			Yes	2	Capillary	DB1	GC-ECD		
031 D S GC-MS 0.01			Yes	2	Capillary	HP 5	GC-ECD GC-NPD GC-MS		
032 D M GC-MS 0.01	Ethyl acetate/ Ciclohexane (2:1)		Yes	4	DB5		GC-ECD, GC-NPD, GC-MS		
033 D M GC-ECD 0.01			Yes	3	25m x 0.32mmx0.52µm	Ultra2	GC-ECD/NPD		

APPENDIX 7. Methods used by participants for determining pesticides

ENDOSULFAN

Determination											
Stationary phase											
Column type											
Part Step (if any)	Clean up step	Internal Standard (if any)	Single Level Calibration	Multi-Level Calibration	Nº of points	Column type	Stationary phase	Determination			
Extraction Solvent	Part Step (if any)	Internal Standard (if any)	Single Level Calibration	Multi-Level Calibration	Nº of points	Column type	Stationary phase	Determination			
Sample Weight (g)	Extraction Solvent	Internal Standard (if any)	Single Level Calibration	Multi-Level Calibration	Nº of points	Column type	Stationary phase	Determination			
Recovery	Sample Weight (g)	Internal Standard (if any)	Single Level Calibration	Multi-Level Calibration	Nº of points	Column type	Stationary phase	Determination			
RL (mg/kg)	Recovery	Internal Standard (if any)	Single Level Calibration	Multi-Level Calibration	Nº of points	Column type	Stationary phase	Determination			
Quantification Method	RL (mg/kg)	Internal Standard (if any)	Single Level Calibration	Multi-Level Calibration	Nº of points	Column type	Stationary phase	Determination			
Quantification using Standards in solvents or matrix	Quantification Method	Internal Standard (if any)	Single Level Calibration	Multi-Level Calibration	Nº of points	Column type	Stationary phase	Determination			
Lab Code	Quantification Method	Internal Standard (if any)	Single Level Calibration	Multi-Level Calibration	Nº of points	Column type	Stationary phase	Determination			
034 D M GC-MS 0.01	100 2	PCB 153	Yes 2	30mx0.3mmx0.25µm	DB5 DB1701	GC-ECD					
035 D M GC-MS 0.02	50 3	GPC	Yes 4	HP-5MS		GC-MS/ECD/FPD					
036 D S GC-MS 0.05	101 3	GPC	Yes			GC-ECD/NPD/MS					
037 D M GC-MS 0.02	100 2	GPC	Yes 4	HP5		GC-MS					
038 D M GC-MS 0.02	101 20	Triphenyl-Phosphat	Yes 3	Capillary-GC	Optima5 Optima 17. DB17-MS	GC-ECD, GC-NPD, GC-M/S					
039 D S GC-MS 0.02	45 3	Aldrin	Yes 3	Restek 30x0.25x0.25	RTX-CL Pesticide I+II	GC-ECD					
040 D S GC-ECD 0.05	340 2	O+GPC	Parathion Yes	Capillary		GC-ECD					
041 D S GC-MS 0.05	78 75	GPC	Yes 3	HP5		GC-ECD					
042 D M GC-MS 0.01	100 2	GPC	Yes 3	Capillary	DB5, DB1701, DB35	GC-ECD, GC-NPD, GC-FPD, GC-MS (single-quad)					
043 D S GC-MS 0.006	50 3	GPC	Yes	Capillary	DB5	GC-ECD					
044 D M GC-MS 0.01	100 2	GPC	Yes 5	ZB35, DB5		GC-ECD					

APPENDIX 7. Methods used by participants for determining pesticides

Lab Code										
Quantification Method										
Quantification using Standards in solvents or matrix										
Confirmation method										
RL (mg/kg)										
Recovery										
Sample Weight (g)										
Extraction Solvent										
Partition Step (if any)										
Clean up step										
Internal Standard (if any)										
Single Level Calibration										
Multi-Level Calibration										
Nº of points										
Column type										
Stationary phase										
Determination										
045	D	M	GC-ECD, GC-MS	0.01	100	2	GPC	Yes	5	DB5 DB1701 GC-ECD, GC- NPD,GC-MS
046	D	M	GC-ECD	0.005	107	50	3	GPC	Yes	Capillary GC
047	D	M	GC-ECD	0.005	100	2	GPC	Yes	3.5 Capillary DB5, DB1701 GC-ECD, GC- NPD,GC-FID	
048	D	S	GC-ECD, MS	0.01	7	4	Yes	Yes	3 Capillary DB5 GC-ECD, GC-MS	
049	D	M	GC-MS	0.05	101	15	4		2 DB5MS GC-ECD/GC-IT	
050	NA									
051	D	M	GC-ECD	0.01	81	25	6	Yes	5 DB-5MS 95%Dymethyl- Polysiloxane GC-ECD	
052	NA									
053	NA									
054	D	M	GC-ECD	0.02	96	25	6	Yes	0 HP5MS 5%Phenyl GC-ECD	
055	D	M	GC-ECD	>0.02	99	15	4	Yes	5 HP1701 Me Siloxane GC-ECD	
056	D	M	GC-ECD	0.02	94.08	15	4	GPC	Yes 5 HP5MS GC-MS	
057	D	S	GC-MS	0.02	20	6	SP-E Yes	Yes	3 DB5 Me Polisiloxan GC-MS	
058	D	M	GC-MS	0.05	15	1	GPC Yes	Yes	3 HP5MS 5%PHSi GC-MS	

ENDOSULFAN

APPENDIX 7. Methods used by participants for determining pesticides

ENDOSULFAN

Determination									
Stationary phase									
Column type									
Partion Step (if any)	Clean up step	Internal Standard (if any)	Single Level Calibration	Multi-Level Calibration	Nº of points	Column type	Stationary phase	Determination	
Extraction Solvent									
Sample Weight (g)									
Recovery									
RL (mg/kg)									
Confirmation method	Quantification Method	Quantification using Standards in solvents or matrix							Lab Code
059 D S GC-MS 0.05	5	1	SPE		Yes	2	Capillary	ZB5	GC-ECD
060 NA									
061 D M GC-MS 0.02	85.5	15	6(Dichloromethane)			Yes	5	DB5	5%Phenyl-95%Dimethyl-Polysiloxane GC-ECD/NPD/MS
062 D S GC-ECD 0.005	89	50	Acetone+ Methanol (1:1)		Empore Disk C8 Fenolofos	Yes	3		GC-ECD/NPD
063 D S GC-ECD 0.02	90	25	3	EXTRRELUT 20		Yes		DB-608. SPB1	GC-ECD
064 D S GC-MS	116.1	10	6	O	YES	Yes	Capillary	HP-5	GC-ECD
065 D S GC-MS 0.005	98.8	10	Dichloromethane			Yes		HP5 POLYSILOXANE	GC-ECD
066 D S GC-MS 0.05	110	25	5	Yes	SPE	YES	Capillary	5%PbSil	GC-ECD/NPD
067 D M GC-MS	10	1		EXTRRELUT	GPC	YES	4		GC-MS
068 D M GC-MS 0.05	90.1	5	6	1	TFF	Yes	5	CIP SIL 8CB	FUSED SILICA
069 D S GC-MS	85	5	1	ASE(Dionex)	GPC	Romiel+ Azobencene+ Triphenyl Phosphate	MDN-5S	5%PHENYL SILICONE	GC-ECD/NPD/FPD

APPENDIX 7. Methods used by participants for determining pesticides

ENDOSULFAN

Determination									
Stationary phase									
Column type									
Partion Step (if any)	Clean up step	Internal Standard (if any)	Single Level Calibration	Multi-Level Calibration	Nº of points	Column type	Stationary phase	Determination	Lab Code
Extraction Solvent	Sample Weight (g)	Recovery	Sample Weight (g)	Extraction Solvent	Partion Step (if any)	Clean up step	Internal Standard (if any)	Single Level Calibration	Quantification Method
Partion Step (if any)	Clean up step	Internal Standard (if any)	Single Level Calibration	Multi-Level Calibration	Nº of points	Column type	Stationary phase	Determination	Lab Code
070 D S GC-ECD 0.01	GPC	M-series	Yes		25m ID 0.32 mm 0.25um	SE 54; OV1701		GC-ECD/NPD	Quantification Method
071 D M GC-MS 0.01	GPC			Yes	2	SE54. OV1701		GC-ECD	Quantification using Standards in solvents or matrix
072 D S 0.01	GPC	Diclofenon	Yes		HP1. HP5			GC-ECD/NPD	Confirmation method
073 D S GC-ECD 0.01			Yes	3	Capillary	SE30/SE54		GC-ECD/NPD	RL (mg/kg)
074	NO RESULTS								
075 D S GC-MS 0.05	SPE	Fenchloraphos	Yes		DB5	GC-ECD, GC-NPD, GC-MS (Single-Qquad)			
076 D M GC-MS 0.05	10	5	YES	O		GC-M/S			
077 D S GC-MS 0.05	25	2		GPC		5%Phenyl-Methyl Siloxane			
078 D M GC-MS 0.05	68	50	1	GPC	Yes	4	HP-5MS	GC-MS	
079 D M GC-MS 0.05	15	4			Yes	4	Narrowbore	DB 5	
080	NO RESULTS								
081 D M GC-MS 0.02			Yes	3-6	HP-5MS	5% Ph. Me S		GC-ECD/FPD/TD	
082 D M GC-MS 0.02			Yes		Capillary	HP-5MS		GC-ECD	

APPENDIX 7. Methods used by participants for determining pesticides

ENDOSULFAN

Lab Code		Quantification Method		Confirmation method		RL (mg/kg)		Recovery		Sample Weight (g)		Extraction Solvent		Partition Step (if any)		Clean up step		Internal Standard (if any)		Single Level Calibration		Multi-Level Calibration		Nº of points		Column type		Stationary phase		Determination	
083	D	M	GC-MS	0.02		15	4		LL Phase Separation			Yes	4	Varian CP 3800		Low Bleed Fused Silica		GC-MS													
084	D	M	GC-MS/MS	0.01		10	1					Yes	4	C/P-Sil 8				GC-MS/MS													
085	D	M	GC-MS	0.02	79	15	4					Yes	4	Capillary		PhMSil 5%															
086	D	M	GC-MS/MS	0.05	84	15	4					Yes	4	30x25x25		DB-5, DB-17		GC/FPD/ECD/MS													
087	D	M	GC-MS	0.03		10	4					Yes	3	HP-5				GC-ECD/NPD/FPD/MS													
088	D		MS			15	4					Yes	5	NarrowBore		HP-5-MS															
089	D	M	GC-MS/MS	0.05		15						Yes	4	Capillary		BP5		GC-MS/MS													
090	D	M	GC-MS/MS	0.05	90	15	1					Yes	3	Capillary		DB-5ms		GC-MS/MS													
091	D	M	GC-MS/MS	0.05		15	1					TIP. Anthracene	Yes	3	HP-5MS				GC-ITD												
092	D	M	GC-MS/MS	0.05		5.2	4					Yes	6	Factor Four				GC-MS/MS-ECD													
093	D	S	GC-MS	0.05		15	4					Yes	4	30m/25x25		CPSIL8CB		GC-MS													
094	D	M	MS/MS			15	4					Yes	4	30mx0.25x0.2		CPSIL8		GC-ECD,GC-NPD,GC-PFPD													

APPENDIX 7. Methods used by participants for determining pesticides

Lab Code	Quantification Method		Confirmation method		RL (mg/kg)	Recovery	Sample Weight (g)	Extraction Solvent	Partition Step (if any)	Clean up step	Internal Standard (if any)	Single Level Calibration	Multi-Level Calibration	Nº of points	Column type	Stationary phase	Determination		
	Quantification using Standards in solvents or matrix																		
095	D	M	GC-ECD	0.01	74	75	1			GPC	Yes	Yes	3	CP-SIL 5CB.CPSI-19CB.CPSIL 8CB		GC-ECD	GC-FPD	GC-ITD	
096	D	M	GC-ITD	0.01	97	15	4			(Injection Only)	Yes	Yes	5	Capillary.25mx0. 22mmID	BPX-5.0.25mm(SGE)		GC-ECD		
097	D	M	GC-MS	0.05	106	30	1			GPC	Triphenyl Phosphate	Yes	5	Capillary	DB-5		GC-MS		
098	D	M	GC-MS	0.05		10	1			SPE	Yes	Yes	4	CP-SIL13CB			GC-MS		
099	D	M	GC-MS	0.02	79	30	1			O		Yes	4	ZB5	30%Phenyl		GC-MS		
100	D	M	GC-MS	0.05	84	10	1			GPC		Yes	3		DB5		GC-MS		
101	D	M	GC-MS	0.01	93	50	5			SPE	Yes			HP 5/HP 50	See Column Type	GC-ECD.GC-NPD			
102	D	M	GC-ITD	0.05	90	50	1			GPC	Yes			At 1701	Attn 1701	GC-FPD			
103	D	S	GC-MS	0.01	87	20	3			LL		Yes	2	HP-5		GC-ECD/NPD			
104	D	M	GC-MS	0.02	108	75	1	1/20		GPC	ALDRIN	Yes	5	Narrowbore	AT-5MS		GC-MS		
105	D	M	GC-ECD	0.05	20.0	67	6	2		Dithalimphos Ethyl-Parathion	1	1	5	NB-1701			GC-ECD		
106	ND	M	GC-MS	0.05	7.5	1								DB-1	0.25mm	GC-EC/MS			
107	D	M	GC-MS	0.05	87.00	20	4				Yes	4	Capillary 30 m		HP-5	GC-ECD/NPD			
108	D	S	GC-ECD	0.05	98	5	3				Yes	3	HP-5	5% Phenyl Methyl Siloxane	GC-ECD				
109	D	S	GC-EC	0	93	100	3				Yes	3	HP-1	Me-SiOH		GC-ECD			

ENDOSULFAN

APPENDIX 7. Methods used by participants for determining pesticides

Lab Code	Quantification Method		Confirmation method		RL (mg/kg)	Recovery	Sample Weight (g)	Extraction Solvent	Partition Step (if any)	Clean up step	Internal Standard (if any)	Single Level Calibration	Multi-Level Calibration	Nº of points	Column type	Stationary phase	Determination	
	Quantification using Standards in solvents or matrix																	
110	D	M	GC-MS	0.01	95.30	20	3	SPE		Yes	5	DB-5				GC-ECD,GC-NPD		
111	D	S		0.03	95.90	10	4	SPE (Silica)		Yes	5	HP-5MS	5%Phenyl			GC-ECD/NPD		
112	D	S		0.05	100	100	3	O		Yes			HP1	MeSilicone		GC-ECD		
113	D	M	GC-ITD	0.026	81	15	4	NO	Hexachloro-Benzene	Yes	5	DB-5MS				GC-ITD		
114	D	S	GC-ECD	0.01		18	1	GPC		Yes	4	Capillary	HP5			GC-ECD		
115	ND	S	GC/MS	0.05		50	4	Dichloro-methane	LL	Yes		HP(30x0.25x0.25)	5MS			GC/MS		
116	D	S	GC/MS	0.05	103	50	2	SPE		Yes		HP5-MS				GC/MS		
117	NA																	
118	D	M	GC-MS	0.02	95	10	6	SPE	Yes	Yes	3	DB-35MS	35%Phenyl			GC-MS		
119	D	M	GC-MS	0.02		10	1	NO		Yes		Capillary	DB-5MS			GC-MS		
120	D	M	GC-FPD	0.01		50	1	GPC		Yes	2	30mx0.26mm	DB-5MS			GC-ITD		
121	D	M	GC-MS	0.008		25	1	GPC		Yes	5	Capillary	DB-5MS	(Single Quad)ECD, NPD		GC-MS		

ENDOSULFAN

APPENDIX 7. Methods used by participants for determining pesticides

ENDOSULFAN

Lab Code	Quantification Method							
	Quantification using Standards in solvents or matrix							
Confirmation method								
RL (mg/kg)	Recovery							
	Sample Weight (g)							
Extraction Solvent								
Partition Step (if any)	Clean up step							
	Internal Standard (if any)							
Single Level Calibration	Nº of points							
	Column type							
Stationary phase	Determination							
	NO RESULTS							
122 D S GC-MS 0.01 109.3 30 1 GPC Yes 3 Chrompack 50mx0.25mm x0.25mm CPSL-8CB GC-ECD								
123 D S GC-MS 0.02 30 1 GPC Yes 3 30mx0.32mm CP-SII 8 CB GC-ECD								
124 D S 0.05 30 1 GPC Yes 3 Restek STX-CLP GC-ECD								
125 D S GC-MS 0.05 25 1 GPC Yes 5 Capillary RTX 17025 GC-ECD								
126								
127 D S GC-MS 0.001 30 1 GPC Yes 3 50mx0.32mm x0.25um CP-SII 8 CB GC-ECD								
128 D S GC-ECD 0.05 114 30 1 GPC Yes 3 30mx0.25mm x0.25um RTX-CLP GC-ECD								
129 D S 0.05 30 1 GPC Yes 3 Restek Stx-Cl-Pest. GC-ECD								
130 D M GC-MS 0.05 30 1 GPC Yes 3 Restek RTX-5MS GC-ECD								

APPENDIX 7. Methods used by participants for determining pesticides

Lab Code	Quantification Method						Determination
	Confirmation method						
001 D M LC-MS-MS 0.05	85	15	4				GC-ECD NPD MS
002/ 003 D M GC-MS 0.1	130	15	4	PCB97	Yes	3	DB5 60x0.25x0.1 60x0.25x0.15
004 D S LC-MS 0.02	96	25	Acetone. Petroleumether. Dichloromethane		Yes	6	DB31701 C18 LC-MS/MS
005 ND	0.06	50	6	O+LL	Yes	4	100x2.1 30x0.5x3x0.5 RT-X-WAR GC-NPD
006 D M	0.02	90	10	5		5	R-P C18 LC-MS/MS
007 D M LC-MS/MS 0.01	83	10	Methanol Ammonia Acetate (95%;5% 20mM Acetic Acid)	¹³ C ₆ -Carbaryl	Yes	3	4u.10cm.3mm ID C18 LC-MS/MS
008 D S	0.05	124	25	6		4	DB 1701/01 GC-ECD
009 D S LC-MS/MS 0.01	75.0	50	4	Yes LL	Yes	3	3mmx25mm C18 LC-MS/MS
010 ND	0.5	100	3	LL	No Aldrin	3	DB 5 DB31701 GC-ECD GC-NPD Capillary
011 NA							

IMAZALIL

APPENDIX 7. Methods used by participants for determining pesticides

IMAZALIL

Lab Code		Quantification Method		Confirmation method		RL (mg/kg)		Recovery		Sample Weight (g)		Extraction Solvent		Partition Step (if any)		Clean up step		Internal Standard (if any)		Single Level Calibration		Multi-Level Calibration		Nº of points		Column type		Stationary phase		Determination		
012	D	M	GC-MS	0.01	70-110	50	3										Yes	8	HP5MS										GC-MS			
013	D	S	GC-NPD	0.02	81	25	Ethyl Acetate-Dichloromethane																					GC-FPD/NPD/GC-MS				
014	D	S	GC-MS	0.05		40	3		L								Yes	3	GC-Capillary		DB5								GC-ECD/NPD-MS			
015	D	S	GC-MS	0.15		50	1/DCM (80/20)									Yes	2	Capillary 0.32		DB5, DB200									GC-NPD			
016	D	S		0.05		50	3	Yes	SPE							Yes	4	AT1701/AT1/DB05-MS		GC-ECD/PFPD/NPD/MS												
017	NA																															
018	D	M	GC-MS	0.01		10	5		SPE	Yes							Yes	5	AtlantisDc18		LC-MS/MS											
019	D	M	GC-TOF	0.005	89	10	5										Yes	4	ZORBAX	C18	LC-MS/MS											
020	D	M	LC-MS/MS	0.010		10	6		SPE								Yes	3	Aqua 3	C18	LC-MS/MS											
021	D	S	GC-MS	0.05	79	50	2		GPC										BD1/5/35/1301. HPS0/1701/5MS		GC-NPD											
022	D	M		0.01	133	10	6,(Methanol)									Yes	7	2.1x125mm; 3µm	RP-C18	LC-MS/MS												
023	D	M	LC-MS/MS	0.01		10	Methanol									Yes	5	2.1x100mm	Atlantis C18	LC-MS/MS												

APPENDIX 7. Methods used by participants for determining pesticides

IMAZALIL

Determination									
Stationary phase									
Column type									
Partion Step (if any)	Clean up step	Internal Standard (if any)	Single Level Calibration	Multi-Level Calibration	Nº of points	Column type	Stationary phase	Determination	
Extraction Solvent									
Sample Weight (g)									
Recovery									
RL (mg/kg)									
Confirmation method	Quantification using Standards in solvents or matrix	Quantification Method	Lab Code						
024 D M LC-MS/MS 0.005	10 Acetonitril	Yes 5 Aqua	024	D M LC-MS/MS 0.005	10 Acetonitril	Yes 5 Aqua	C18	LC-MS/MS	
025 D M LC-MS/MS 0.01	20 6	Yes 2x4 HPLC(RP)	025	D M LC-MS/MS 0.01	20 6	Yes 2x4 HPLC(RP)	RPC18)	LC-MS/MS	
026 D S GC-MS 0.05	10 5	Yes 3 Capillary	026	D S GC-MS 0.05	10 5	Yes 3 Capillary	Varien VF-5MS	GC-ECD GC-NPD GC-MS	
027 D M GC-MS 0.02	100 2	Yes 30mx0.25x0.25	027	D M GC-MS 0.02	100 2	Yes 30mx0.25x0.25	Factor Four	GC-NPD	
028 D M GC-MSD 0.02	102 10	Yes 7 KS	028	D M GC-MSD 0.02	102 10	Yes 7 KS	HPMS	GC-MS	
029 D M GC-MS 0.04	100 2	Yes 3-5 GC	029	D M GC-MS 0.04	100 2	Yes 3-5 GC	DB5 HP5-MS DB17	GC-ECD GC-NPD GC-MS (single-quadrupole)	
030 D M LC-MS 0.02	10 Methanol	Yes 3 Synergi-polar RP	030	D M LC-MS 0.02	10 Methanol	Yes 3 Synergi-polar RP	LC-MIS		
031 D S GC-MS 0.02	10 5	Yes 2 Capillary-LC	031	D S GC-MS 0.02	10 5	Yes 2 Capillary-LC	HP5 RP18	GC-ECD GC-NPD GC-MS	
032 D M LC-MS/MS 0.01	83 10 6(Methanol)	Yes 5 Aqua5μ	032	D M LC-MS/MS 0.01	83 10 6(Methanol)	Yes 5 Aqua5μ	C18	LC-MS/MS	
033 D M 0.02	100 2	Yes 3 25m x 0.32mmx0.52μm	033	D M 0.02	100 2	Yes 3 25m x 0.32mmx0.52μm	Ultra2	GC-ECD/NPD	

APPENDIX 7. Methods used by participants for determining pesticides

IMAZALIL

Lab Code		Quantification Method		Confirmation method		RL (mg/kg)		Recovery		Sample Weight (g)		Extraction Solvent		Partition Step (if any)		Clean up step		Internal Standard (if any)		Single Level Calibration		Multi-Level Calibration		Nº of points		Column type		Stationary phase		Determination	
034	D	M	GC-MS	0.03	100	2		GPC	Parathion-Methyl	Yes				30mx0.3mmx0.5μm		DB5	DB17			GC-PND											
035	D	M	LC-MS/MS	0.01	10	5		O			Yes	5	Aqua5μl. 50x2mm		RP18					LC-MS/MS											
036	D	S	GC-MS	0.02	101	3		GPC		Yes									GC-ECD/NPD/MS												
037	D	M	GC-MS	0.02	100	2		GPC	TPP		Yes	4	HP5						GC-MS												
038	D	M	LC-MS/MS	0.01	73	20	2		Triphenyl-Phosphat		Yes	3	Capillary-GC	Optimas, DB17-MS		GC-ECD, GC-NPD, GC-MS															
039	D	S		0.01	10	5	LL	PSA	Methoxi-Fenoziide		Yes	6	Aqua(phenomex)	C18		LC-MS/MS															
040	ND	S	GC-ECD	0.01	340	2		O+GPC	Parathion	Yes			Capillary			GC-ECD															
041	D	M	LC-MS	0.02	103	10	Methanol			Yes	7	Atlantis	dc18		LC-MS/MS																
042	D	M	GC-MS	0.02	100	2		GPC	EPN	Yes	3	Capillary	DB5, DB1701, DB35	GC-ECD, GC-NPD, GC-FPD, GC-MS (single-quad)																	
043	D	M	LC-MS/MS	0.006	10	6 (Methanol)		SPE		Yes	2		C18		LC-MS/MS																
044	D	M		0.01	100	2				Yes	4		RP8		LC-MS/MS																

APPENDIX 7. Methods used by participants for determining pesticides

Lab Code									
Quantification Method									
Quantification using Standards in solvents or matrix									
Confirmation method									
RL (mg/kg)									
Recovery									
Sample Weight (g)									
Extraction Solvent									
Partition Step (if any)									
Clean up step									
Internal Standard (if any)									
Single Level Calibration									
Multi-Level Calibration									
Nº of points									
Column type									
Stationary phase									
Determination									
045	D	M	LC-MS/MS	0.02	10	5	SPE	Yes	3
046	D	M	GC-PND	0.02	82	50	GPC	Yes	Luna
047	D	M	GC-NPD	0.01	100	2	GPC	Yes	C18
048	NA						Capillary	DB17. HP5	LC-MS/MS
049	ND	M	GC-MS	0.5	15	4	Capillary	3. 5	GC
050	NA						DB5	DB1701	NPD,GC-FID
051	NA						DB5MS		GC-ECD, GC-FID
052	NA								NPD,GC-FID
053	NA								
054	NA								
055	NA								
056	NA								
057	D	S	HPLC-UV	0.05	20	1	LL	Yes	3
058	ND	M	GC-MS	0.03	15	1	GPC	Yes	HP5MS
									5%PHSi
									GC-MS

IMAZALIL

APPENDIX 7. Methods used by participants for determining pesticides

IMAZALIL

Determination									
Stationary phase									
Column type									
Partion Step (if any)	Clean up step	Internal Standard (if any)	Single Level Calibration	Multi-Level Calibration	Nº of points	Column type	Stationary phase	Determination	
Extraction Solvent									
Sample Weight (g)	Recovery	RL (mg/kg)							
Confirmation method									
Quantification using Standards in solvents or matrix	Quantification Method	Lab Code							
059 N/A		059							
060 ND	0.1	060	ND	0.1	10	6(Dichloromethane)			
061 D M	GC-MS	061	M	GC-MS	0.02	99.1	15	6(Dichloromethane)	
062 ND S	HPLC-DAD	062	S	HPLC-DAD	0.25	10	Dichloromethane	LC-NH ₂ Columns	
063 D S	GC-ECD	063	S	GC-ECD	0.05	84	25	3	EXTRRELUT 20
064 D S	GC-MS	064	D	GC-MS		72.1	10	6	O
065 D S	GC-MS	065	D	GC-MS	0.005	81.2	10	Dichloromethane	
066 D S	GC-MS	066	D	GC-MS	0.05	92	25	5	Yes
067 D M	GC-MS	067	D	GC-MS		10	1		EXTRRELUT
068 D M	GC-MS	068	D	GC-MS	0.05	77.5	5	6	TFF
069 D S	GC-MS	069	S	GC-MS		79	5	1	ASE(Dionex) GPC
Ronnel+ Azobencene+ Triphenyl phosphate									
5%PHENYL SILICONE									
MDN-SS									
GC-ECD/NPD/FPD									

APPENDIX 7. Methods used by participants for determining pesticides

IMAZALIL

Determination										
Stationary phase										
Column type										
Single Level Calibration										
Lab Code	Quantification Method	Quantification using Standards in solvents or matrix	Confirmation method	RL (mg/kg)	Recovery	Sample Weight (g)	Extraction Solvent	Partition Step (if any)	Clean up step	Internal Standard (if any)
070 D S GC-ECD 0.02	GPC	M-series	Yes	25m ID 0.32 mm 0.25um	SE 54; OV1701					
071 D M GC-MS 0.02	GPC		Yes	2	SE54; OV1701					GC-ECD/NPD
072 D S 0.02 90	GPC	Diclofenon	Yes		HP1; HP5					GC-ECD/NPD
073 D S GC-ECD 0.01			Yes	3	SE30/SE54					GC-ECD/NPD
074	NO RESULTS									
075 D S GC-MS 0.02	SPE	Fenchloraphos	Yes	DB5	GC-ECD, GC-NPD, GC-MS (SINGLE-QUAD)					
076 D M GC-MS 0.01	10	5	Yes O	3	HP-5MS					GC-MS
077 ND	0.02									
078 D M GC-MS 0.05 79	50	1	GPC	4	Narrowbore	HP 50				GC-NPD
079 D M GC-MS 0.02	15	4		1	SPB 1701					GC-NPD
080	NO RESULTS									
081 D M GC-MS 0.01	20	1		3-6	HP-5MS	5%Ph. Me S				GC-ECD/FPD/TD
082 D M GC-MS 0.05	15	4	Yes		Capillary	HP-5MS				GC-NPD

APPENDIX 7. Methods used by participants for determining pesticides

IMAZALIL

		Lab Code	
		Quantification Method	
		Quantification using Standards in solvents or matrix	
		Confirmation method	
		RL (mg/kg)	
		Recovery	
		Sample Weight (g)	
		Extraction Solvent	
		Partition Step (if any)	
		Clean up step	
		Internal Standard (if any)	
		Single Level Calibration	
		Multi-Level Calibration	
		Nº of points	
		Column type	
		Stationary phase	
		Determination	
083	NA		
084	D	M	GC-MS/MS 0.01
085	D	M	GC-MS 0.02
086	D	M	LC-MS/MS 0.05
087	D	M	GC-MS 0.02
088	NA		
089	NA		
090	NA		
091	NA		
092	D	M	GC-MS/MS 0.02
093	NA		
094	D	M	MS/MS
		15	4
		Yes	4
		5.2	4
		Yes	6
		Factor Four	
		GC-MS/MS-ECD	
		GC-MS/ECD	
		GC-ECD/GC-NPD,GC-PFPD	
		CPSIL8	
		30mx0.25x0.2	

APPENDIX 7. Methods used by participants for determining pesticides

IMAZALIL

Determination									
Stationary phase									
Column type									
Partion Step (if any)	Internal Standard (if any)	Single Level Calibration	Multi-Level Calibration	Nº of points	Sample Weight (g)	Extraction Solvent	Recovery	RL (mg/kg)	Lab Code
Clean up step	Internal Standard (if any)	Single Level Calibration	Multi-Level Calibration	Nº of points	Sample Weight (g)	Extraction Solvent	Recovery	RL (mg/kg)	Quantification Method
095 D M LC-MS/MS 0.01 109 75 1 Yes Yes									Quantification using Standards in solvents or matrix
096 D S LC-MS/MS 0.01 90 7.5 4(+Na ₂ SO ₄)		Yes 4 150x3.2mmID	Altima C18.5mm	LC-MS/MS					LC-MS/MS
097 D M LC-MS/MS 0.02 95 10 5		Yes 5 Narrow Bore	C18	LC-MS/MS					
098 D M LC-MS/MS 0.02 10 1	LL Yes	Yes 4 RP-HPLC	C-18	LC-MS/MS					
099 D M GC-MS 0.01 62 30 1 O		Yes 4 ZB5	30%oPhenyl	GC-MS					
100 D M LC-MS/MS 0.02 88 10 1		Yes 4 C-18		LC-MS/MS					
101 ND M GC-MS 0.02 50 5	SPE	Yes HP 5/HP 50	See Column Type	GC-ECD,GC-NPD					
102 ND 0.02 50 1	GPC	Yes At-1701	Attn-1701	GC-FPD					
103 D S GC-MS 0.02 102 20 3 LL		Yes 2 HP-5							
104 D M GC-MS 0.01 115 75 1 1/20 GPC Aldrin		Yes 5 Narrowbore	A ⁻ T-5MS	GC-MS					
105 NA									
106 D M LC-MS/MS 0.02 7.5 1		Yes C-18		GC-ECD/NPD					
107 D S LC-MS 0.02 103.5 20 4		Yes 6 Xterra 100 mm	C-18	LC-MS/MS					
108 NA									
109 NA									

APPENDIX 7. Methods used by participants for determining pesticides

IMAZALIL

Lab Code	Quantification Method						Determination					
	Quantification using Standards in solvents or matrix											
Confirmation method						RL (mg/kg)	Determination					
Recovery						Sample Weight (g)	Determination					
Extraction Solvent						Sample Weight (g)	Determination					
Partition Step (if any)						Sample Weight (g)	Determination					
Clean up step						Sample Weight (g)	Determination					
Internal Standard (if any)						Sample Weight (g)	Determination					
Single Level Calibration						Sample Weight (g)	Determination					
Multi-Level Calibration						Sample Weight (g)	Determination					
Nº of points						Sample Weight (g)	Determination					
Column type						Sample Weight (g)	Determination					
Stationary phase						Sample Weight (g)	Determination					
Determination						Sample Weight (g)	Determination					
110	D	M	GC-MS	0.02	107.2	20	3	Yes	3	DB-5	GC-ECD,GC-NPD	
111	NA											
112	NA											
113	D	M	GC-TID	0.05	105	15	4	No	Hexachloro-Benzene	Yes	5	DB-5MS
114	D	S	GC-ECD	0.05	37	6		GPC		Yes	4	GC-TID
115	ND		GC/MS	0.02	50	4		Dicloro-methane	LL	Yes		Capillary
116	D	S	GC/MS	0.02	108	50	2	SPE		Yes		HP5
117	ND	M	GC-MS	0.02	120.4	20	4	Yes	GPC	Yes	5	HP5-MS
118	D	M	GC-MS	0.02	84	10	6	SPE	Yes	Yes	3	GC-MS
119	D	M	LC-MS/MS	0.01	10	5	Yes	LL	DB-35MS	Yes	4	35%Phenyl
120	NA											LC-MS/MS
121	D	M	LC-MS	0.001	12.5	5	No			Yes	7	HPLC
												C-18
												LC-MS/MS

APPENDIX 7. Methods used by participants for determining pesticides

IMAZALIL

Determination										
Stationary phase										
Column type										
Single Level Calibration										
Partion Step (if any)	Internal Standard (if any)	Clean up step	Extraction Solvent	Sample Weight (g)	Recovery	RL (mg/kg)	Confirmation method	Quantification using Standards in solvents or matrix	Quantification Method	Lab Code
126	NO RESULTS									
127	D	S	0.02	30	1	GPC		Yes	3	50mx0.53mmx0.5um
128	D	S	GC-NPD	0.05	113	30	1	GPC	Yes	3
129	D	S		0.02	30	1		GPC	Yes	Purosphere RP18. 250/4mm 5um
130	D	M	HPLC-UV	0.02	30	1			Yes	3
										Varian
										Factor Four V15
										CPSII 8CB
										GC-NPD

APPENDIX 7. Methods used by participants for determining pesticides

IMIDACLOPRID

Determination									
Stationary phase									
Column type									
Nº of points									
Single Level Calibration									
Multi-Level Calibration									
Internal Standard (if any)									
Clean up step									
Partition Step (if any)									
Extraction Solvent									
Sample Weight (g)									
Recovery									
RL (mg/kg)									
Quantification using Standards in solvents or matrix									
Confirmation method									
Quantification Method									
Lab Code									
001	D	M	LC-MS-MS	0.01	97	10	6		
002/ 003	NA								
004	D	S	LC-MS	0.05	81	50	Methanol. Water	LL. O	
005	NA								
006	D	M	LC-MS-MS	0.05	85	10	5		
007	D	M	LC-MS/MS	0.01	85	10	Methanol Ammonia Acetate (95%:5%) 20mm Acetic Acid	¹³ C ₆ -carbaryl	
008	NA								
009	D	M	LC-MS/MS	0.01	112.0	50	4	Yes	5
010	NA								
011	NA								

IMIDACLOPRID

		Lab Code	
		Quantification Method	
		Quantification using Standards in solvents or matrix	
		Confirmation method	
		RL (mg/kg)	
		Recovery	
		Sample Weight (g)	
		Extraction Solvent	
		Partition Step (if any)	
		Clean up step	
		Internal Standard (if any)	
		Single Level Calibration	
		Multi-Level Calibration	
		Nº of points	
		Column type	
		Stationary phase	
		Determination	
012	NA		
013	D	S	LC-MS-MS
014	NA		
015	NA		
016	ND	S	0.05
017	ND	S	GC-MS
018	D	M	0.02
019	D	M	2 Transitions
020	D	M	LC-MSMS
021	D	S	LC-MSMS
022	D	M	0.01
023	D	M	LC-MSMS
			0.01
			99
			6-(Methanol)
			Methanol

IMIDACLOPRID

Determination																		
Stationary phase																		
Column type																		
Partion Step (if any)	Clean up step	Internal Standard (if any)	Single Level Calibration	Multi-Level Calibration	Nº of points	Column type												
Extraction Solvent	Sample Weight (g)	Recovery	Nº of points		Stationary phase													
Partition Step (if any)	RL (mg/kg)	Quantification Method	Nº of points		Stationary phase													
Confirmation method	Quantification using Standards in solvents or matrix	Nº of points		Stationary phase														
Lab Code	Lab Code	Nº of points		Stationary phase														
024	D	M	LC-MS/MS	0.005	10	Acetonitril	Yes	5	Aqua									
025	D	M	LC-MS/MS	0.01	20	6	Yes	2x4	HPLC(RP)									
026	NA								RP(C18)									
027	D	M	GC-MS	0.02	10	5	O	Yes	50x2mm									
028	D	S	HPLC-JV	0.05	78	10	1	Yes	7									
029	D	M	LC-MS/MS	0.01	10	6	1	Yes	HPLC									
030	D	M	LC-MS	0.04	10	Methanol	Isopropution D6	Yes	3-5									
031	D	S	HPLC-DAD	0.05	10	5	SPF	Yes	3									
032	D	M	LC-MS/MS	0.01	83.3	10	6(Methanol)	Yes	2									
033	NA								Capillary-LC									
034	D			0.05	100	2	GPC	Yes	HP5									
							LL		RP18									
									GC-EC									
									HPLC-DAD/HPLC-FL									
									L-C-MS/MS									
									C18									
									HPLC-UV									
									250x4x5									
									RP8									

IMIDACLOPRID

APPENDIX 7. Methods used by participants for determining pesticides

Determination									
Stationary phase									
Column type									
Single Level Calibration									
Partion Step (if any)	Clean up step	Internal Standard (if any)	Nº of points						
Extraction Solvent	Sample Weight (g)	Recovery							
Confirmation method	RL (mg/kg)	Quantification using Standards in solvents or matrix							
Lab Code	Quantification Method	Quantification Method							
035	D	M	LC-MS/MS	0.005	10	5	O	Yes	5
036	NA								Aqua5µ. 50x2mm
037	NA								RP18
038	D	M	LC-MS/MS	0.005	62	20	2	Triphenyl-Phosphat	Yes
039	D	S		0.01	10	5	UL	Methoxi-Fenoziide	Yes
040	NA							Aqua(phenomex)	6
041	D	M	LC-MS	0.05	123	10	Methanol		RP18
042	NA								LC-MS/MS
043	D	M	LC-MS/MS	0.006	10	6 (Methanol)	SPE		C18
044	D	M		0.01	100	2			LC-MS/MS
045	D	M	LC-MS/MS	0.05	10	5	SPE	Yes	4

APPENDIX 7. Methods used by participants for determining pesticides

		Lab Code	
046	NA	Quantification Method	
047	NA	Quantification using Standards in solvents or matrix	
048	NA	Confirmation method	
049	NA	RL (mg/kg)	
050	NA	Recovery	
051	NA	Sample Weight (g)	
052	NA	Extraction Solvent	
053	NA	Partition Step (if any)	
054	NA	Clean up step	
055	NA	Internal Standard (if any)	
056	NA	Single Level Calibration	
057	D	Multi-Level Calibration	
058	M	Nº of points	
059	ND	Column type	
060	NA	Stationary phase	
		Determination	

IMIDACLOPRID

APPENDIX 7. Methods used by participants for determining pesticides

IMIDACLOPRID

Lab Code									
Quantification Method									061
Quantification using Standards in solvents or matrix									062
Confirmation method									063
RL (mg/kg)									064
Recovery									065
Sample Weight (g)									066
Extraction Solvent									067
Partition Step (if any)									068
Clean up step									069
Internal Standard (if any)									070
Single Level Calibration									071
Multi-Level Calibration									D
Nº of points									M
Column type									0.05
Stationary phase									15
Determination									GPC
LC-NH₂ Columns									C18 Octadecil
LUNA 2									HPLC-DAD
LC 18									HPLC-DAD
Yes									

APPENDIX 7. Methods used by participants for determining pesticides

							Lab Code
072	NA						Quantification Method
073	NA						Quantification using Standards in solvents or matrix
074							Confirmation method
							RL (mg/kg)
							Recovery
							Sample Weight (g)
							Extraction Solvent
							Partition Step (if any)
							Clean up step
							Internal Standard (if any)
							Single Level Calibration
							Multi-Level Calibration
							Nº of points
							Column type
							Stationary phase
							Determination
NO RESULTS							
075	NA						
076	NA						
077	NA						
078	D	S	0.05	84	50	1	
079	NA						
080							
081	NA						
082	NA						
083	NA						
084	NA						

IMIDACLOPRID

APPENDIX 7. Methods used by participants for determining pesticides

IMIDACLOPRID

Lab Code									
Quantification Method									
Quantification using Standards in solvents or matrix									
Confirmation method									
RL (mg/kg)									
Recovery									
Sample Weight (g)									
Extraction Solvent									
Partition Step (if any)									
Clean up step									
Internal Standard (if any)									
Single Level Calibration									
Multi-Level Calibration									
Nº of points									
Column type									
Stationary phase									
Determination									
085	D	M	GC-MS	0.05	85	15	4	Yes	4
086	D	M	LC-MS/MS	0.02	96	5	5	Yes	4
087	NA							Luna(Phenom)	C-18
088	D			0.02	15	Dichloromethane	GPC		LC-MS/MS
089	NA								
090	D	M	LC-MS/MS	0.05	70	15	1	Yes	5
091	NA								
092	NA								
093	NA								
094	NA								
095	D	M	LC-MS/MS	0.01	89	75	1	Yes	

APPENDIX 7. Methods used by participants for determining pesticides

IMIDACLOPRID

Determination											
Stationary phase											
Column type											
Partion Step (if any)	Clean up step										
Extraction Solvent	Internal Standard (if any)										
Sample Weight (g)	Single Level Calibration										
Recovery	Multi-Level Calibration										
RL (mg/kg)	Nº of points										
Confirmation method	Column type										
Quantification using Standards in solvents or matrix	Stationary phase										
Quantification Method	Determination										
Lab Code	Determination										
096	D	S	LC-MS/MS	0.01	92	7.5	4(+Na ₂ SO ₄)			Yes	4
097	D	M	LC-MS/MS	0.02	114	10	5			Yes	5
098	D	M	LC-MS/MS	0.05		10	1	LL	Yes	Yes	4
099	NA									RP-HPLC	C-18
100	D	M	LC-MS/MS	0.05	95	10	1				LC-MS/MS
101	NA										LC-MS/MS
102	NA										LC-MS/MS
103	NA										Altima C18.5mm
104	NA										Altima C18.5mm
105	NA										LC-MS/MS
106	NA										LC-MS/MS
107	D	S	LC-MS	0.05	112.5	20	4	Yes	6	X-terra 100 mm	C-18
108	NA										LC-MS
109	NA										
110	NA										

APPENDIX 7. Methods used by participants for determining pesticides

IMIDACLOPRID

		Lab Code	
		Quantification Method	
		Quantification using Standards in solvents or matrix	
		Confirmation method	
		RL (mg/kg)	
		Recovery	
		Sample Weight (g)	
		Extraction Solvent	
		Partition Step (if any)	
		Clean up step	
		Internal Standard (if any)	
		Single Level Calibration	
		Multi-Level Calibration	
		Nº of points	
		Column type	
		Stationary phase	
		Determination	
111	NA		
112	NA		
113	NA		
114	NA		
115	ND	GC/MS	0.05
116	NA		
117	NA		
118	NA		
119	D	M	LC-MS/MS
120	NA		
121	D	M	LC-MS
122	D	S	GC-MS

APPENDIX 7. Methods used by participants for determining pesticides

IMIDACLOPRID

Determination									
Stationary phase									
Column type									
Partion Step (if any)	Clean up step	Internal Standard (if any)	Single Level Calibration	Multi-Level Calibration	Nº of points	Column type	Stationary phase	Determination	
Extraction Solvent									
Sample Weight (g)									
Recovery									
RL (mg/kg)									
Quantification Method									
Lab Code									
123 D S GC-ECD 0.05	30	1	GPC	Yes		250mx4mm	C-18	HPLC:UV	
124 D S HPLC 0.05	30	1				Yes	3	SGE	BPX-5
125 D M GC-MS 0.05	25	6	O	Yes	5	Capillary	RTX 200	GC-MS	
126	NO RESULTS								
127 D S 0.05	30	1	O	Yes	4	NUCLEOSIL 100	C-18	HPLC:UV	
128 D S HPLC-DAD 0.05	98	30	1	GPC	Yes	3	Purosphere RP18, 250x4mm 5um	HPLC-DAD	
129 D S 0.05	30	1	GPC	Yes	3	Supelcosil	LC-18	HPLC:UV	
130 D M 0.05	30	1	GPC	Yes	3	Waters	Nova-Pack C-18	HPLC:UV	

APPENDIX 7. Methods used by participants for determining pesticides

DITHIOCARBAMATES

								Lab Code
								Quantification Method
								Quantification using Standards in solvents or matrix
								Confirmation method
								RL (mg/kg)
								Recovery
								Sample Weight (g)
								Extraction Solvent
								Partition Step (if any)
								Clean up step
								Internal Standard (if any)
								Single Level Calibration
								Multi-Level Calibration
								Nº of points
								Column type
								Stationary phase
								Determination
001	D	S	0.3	91	50			
002/ 003	D	S	0.02	100	50	Head-Space	Dichloro methane	
004	NA							
005	D	S	GC-ECD 0.05	85	25	6		
006	NA							
007	NA							
008	NA							
009	D	UV	0.2 93.8	100	6			
010	NA							
011	NA							

APPENDIX 7. Methods used by participants for determining pesticides

DITHIOCARBAMATES

Lab Code									
Quantification Method									
Quantification using Standards in solvents or matrix									
Confirmation method									
RL (mg/kg)									
Recovery									
Sample Weight (g)									
Extraction Solvent									
Partition Step (if any)									
Clean up step									
Internal Standard (if any)									
Single Level Calibration									
Multi-Level Calibration									
Nº of points									
Column type									
Stationary phase									
Determination									
012	D	S	UV SCAN		70-110	50	Distillation	Yes	6
013	D	S	UV	0.4	80	50			UV
014	D	S	UV	0.2	100			Yes	3
015	D		UV	0.2	100				UV
016	NA								
017	D	S		0.01	5			Yes	5
018	D	S		0.019	15			Yes	8
019	D	S		0.05	50	Distillation			UV
020	D	S	UV	0.010	25			Yes	6
021	D	S		0.08	82.3	25	6		Photometry
022	NA								
023	D	S		0.05	10			Yes	8
									Photometry

APPENDIX 7. Methods used by participants for determining pesticides

DITHIOCARBAMATES

		Lab Code			
		Quantification Method			
		Quantification using Standards in solvents or matrix			
		Confirmation method			
		RL (mg/kg)			
		Recovery			
		Sample Weight (g)			
		Extraction Solvent			
		Partition Step (if any)			
		Clean up step			
		Internal Standard (if any)			
		Single Level Calibration			
		Multi-Level Calibration			
		Nº of points			
		Column type			
		Stationary phase			
		Determination			
024	D	Photometry	0.01	25	No
025	D		0.1	100	Yes
026	NA			6	5
027	D	M	GC-MS	0.02	10
028	D	S	Photometry	0.05	86
029	NA			100	Yes
030	D	S	Photometry	0.05	50
031	D	S		0.01	40
032	D	S	Photometry	0.02	45
033	D		UV	0.02	40
034	NA				Yes
					5
					UV
					UV
					Photometry 272.302.332 nm
					Spectrophotometric

APPENDIX 7. Methods used by participants for determining pesticides

		Lab Code	
		Quantification Method	
035	NA	Quantification using Standards in solvents or matrix	
036	NA	Confirmation method	
037	D M	GC-MS	0.05
038	ND M	GC-MS	0.03
039	D	Headspace/SnCl2	0.05
040	NA	CS2-photometric	50
041	D S	UV-Spectrophotometric	0.05
042	D S	UV	0.05
043	D S	UV	0.01
044	D S	UV	50
045	D S	Photometry	0.05
		RL (mg/kg)	
		Recovery	
		Sample Weight (g)	
		Extraction Solvent	
		Partition Step (if any)	
		Clean up step	
		Internal Standard (if any)	
		Single Level Calibration	
		Multi-Level Calibration	
		Nº of points	
		Column type	
		Stationary phase	
		Determination	

DITHIOCARBAMATES

APPENDIX 7. Methods used by participants for determining pesticides

DITHIOCARBAMATES

								Lab Code	
						Quantification Method			
						Quantification using Standards in solvents or matrix			
						Confirmation method			
						RL (mg/kg)			
						Recovery			
						Sample Weight (g)			
						Extraction Solvent			
						Partition Step (if any)			
						Clean up step			
						Internal Standard (if any)			
						Single Level Calibration			
						Multi-Level Calibration			
						Nº of points			
						Column type			
						Stationary phase			
						Determination			
						UV-Spectrophotometric			
						UV-VIS			
046	D	S	UV	0.01	106	20	Yes	8	
047	D	S	UV	0.01	10	6	Yes	8	
048	NA								
049	NA								
050	NA								
051	NA								
052	NA								
053	NA								
054	NA								
055	NA								
056	NA								
057	D	S	0.25						
058	NA								
059	ND		0.5						
060	D	S	0.2	90	45		Yes	7	
						UV 435nm			

APPENDIX 7. Methods used by participants for determining pesticides

DITHIOCARBAMATES

Lab Code									
Quantification Method									
Quantification using Standards in solvents or matrix									
Confirmation method									
RL (mg/kg)									
Recovery									
Sample Weight (g)									
Extraction Solvent									
Partition Step (if any)									
Clean up step									
Internal Standard (if any)									
Single Level Calibration									
Multi-Level Calibration									
Nº of points									
Column type									
Stationary phase									
Determination									
061	D		UV	0.5	83.2	50			UV
062	D	S	GC-ECD	0.01	85	50	Acetone ⁺ Methanol (1:1)		GC-ECD/NPD
063	NA								
064	D	S			50				
065	D	S		0.05	72.5				
066	D			0.4	50				
067	D	S			50				
068	NA								
069	D	S			77	50			
070	D	S		0.3	50	6			
071	D	S		0.3	100	6			

APPENDIX 7. Methods used by participants for determining pesticides

DITHIOCARBAMATES

Lab Code		Quantification Method		Confirmation method		RL (mg/kg)		Recovery		Sample Weight (g)		Extraction Solvent		Partition Step (if any)		Clean up step		Internal Standard (if any)		Single Level Calibration		Multi-Level Calibration		Nº of points		Column type		Stationary phase		Determination	
072	D	S	0.4	50		Yes	8												Yes	3	HP-5MS	GC-ECD	Internal								
073	D	S	0.5	50		Yes	8												Yes	4		MAS									
074																															
075	D	S	0.5																												
076	D	M	GC-ECD	0.05	1	HEAD-SPACE																									
077	NA																														
078	D	S		0.1	87	100	Acid Hydrolysis																								
079	D	M		0.05	50	6	Y																								
080																															
081	NA																														
082	D	S		0.05	100	6													Yes	5			UV								
083	NA																														
084	NA																														

APPENDIX 7. Methods used by participants for determining pesticides

DITHIOCARBAMATES

Determination								
Stationary phase								
Column type								
Partion Step (if any)	Clean up step		Internal Standard (if any)		Single Level Calibration		Multi-Level Calibration	
Extraction Solvent	Sample Weight (g)		Nº of points		Nº of points		Nº of points	
Confirmation method	RL (mg/kg)		Recovery		Recovery		Recovery	
Quantification Method	Lab Code		Lab Code		Lab Code		Lab Code	
085 D S GC-MS 0.02 81 25 6	Yes	5	Capillary	Porapak	GC-FPD-MS			
086 D M GC/FPD 0.2 99 5 6	Yes	4	30x0.25x5	DB-5	Dithiocarbamates			
087 D S GC/MS 0.04 4 5	Y	4	SPB-1 Sulphur	GC-FPD(HS)				
088 NA								
089 D S 0.05 50	Y	4						
090 ND S 0.2 50 6	Y	9						
091 NA								
092 D D UV 0.05 5 6	Y	5						
093 D S UV 0.05 15 4	Y	4	30m/25x25	CPSIL8CB	GC-MS			
094 D M 20 6	Y	4	30mx0.53x0.25	BP5	GC-FPD			
095 D M GC-FPD 0.025 80 50 6	Y	2	502.2(30m.0.53mmID.3μ mdf	RTX-Crossbond Phenylmethyl Polysiloxane	GC-FPD			

APPENDIX 7. Methods used by participants for determining pesticides

DITHIOCARBAMATES

Lab Code											
Quantification Method											
Quantification using Standards in solvents or matrix											
Confirmation method											
RL (mg/kg)											
Recovery											
Sample Weight (g)											
Extraction Solvent											
Partition Step (if any)											
Clean up step											
Internal Standard (if any)											
Single Level Calibration											
Multi-Level Calibration											
Nº of points											
Column type											
Stationary phase											
Determination											
096	D	S	GC-TID	0.05	79	50	6(iso-octane)			Y	4
097	D	M	GC-FPD	0.02	84	25	6			Y	6
098	D	M	GC-MS	0.05		25	6			Yes	3
099	D	S	GC-MS	0.05	86	50	6			Yes	4
100	D	S	GC-MS	0.05	95	50	L			Yes	3
101	NA										
102	NA										
103	NA										
104	NA										
105	NA										
106	D			0.05	50					Yes	5
107	D	S	UV	0.05	90.00	200				Yes	5
108	NA										
109	NA										
110	D	S		0.05	96.70	50	6			Yes	6
											Spectrophotometry

APPENDIX 7. Methods used by participants for determining pesticides

DITHIOCARBAMATES

Lab Code									
Quantification Method									
Quantification using Standards in solvents or matrix									
Confirmation method									
RL (mg/kg)									
Recovery									
Sample Weight (g)									
Extraction Solvent									
Partition Step (if any)									
Clean up step									
Internal Standard (if any)									
Single Level Calibration									
Multi-Level Calibration									
Nº of points									
Column type									
Stationary phase									
Determination									
111	D		0.05	84.00	10	4	SPE (Silica)	Yes	5
112	D	S	0.05	80	40			Yes	3
113	NA								
114	NA								
115	NA								
116	NA								
117	D	S	GC-MS	0.05	100.8	50	6	No	
118	NA							Yes	6
119	D	M	GC-MS	0.05	30	6	Yes	No	
120	NA							Yes	3
121	D	M	GC-MS	0.001	5		Yes	Capillary	DB-5MS
122	D	S	GC-FPD	0.1	73.5	50	6	No	GC-MS
							Yes	4	HP-5MS
									GC-ECD/NPD
									Spectrophotometric Method
									HP-VOC
									GC-ITD
									Spectrophotometry

APPENDIX 7. Methods used by participants for determining pesticides

DITHIOCARBAMATES

Determination										
Stationary phase										
Column type										
Single Level Calibration										
Partion Step (if any)	Internal Standard (if any)	Nº of points	Clean up step							
			Extraction Solvent							
			Sample Weight (g)							
			Recovery							
			RL (mg/kg)							
			Quantification Method							
			Quantification using Standards in solvents or matrix							
			Confirmation method							
123	D	S	0.05	25	Liquid/Gas (Headspace)	Thiophen	Yes	4	0.9mlx2mm	10%SE-30
124	D		0.05	100	6	O	Yes	6		Spectrophotometry
125	D	S	0.05	82	50	6	Yes	5	Packed	GC-FPD
126									SE30	
127	D	S	0.02	50	6		Yes	4	25mx0.53mmx1.0um	CP-Sil 8 CB
128	D	M	0.05	96	50		Yes	3	30mx0.53mmx1.5um	DB1
129	D	M	0.05	25	6		Yes	4	Chrompack	Tenax
130	D	M	0.1	30	6		Yes	4	Chrompack	CPSil 8CBD

OXYDEMETON-METHYL

APPENDIX 7. Methods used by participants for determining pesticides

Lab Code									
Quantification Method									
Quantification using Standards in solvents or matrix									
Confirmation method									
RL (mg/kg)									
Recovery									
Sample Weight (g)									
Extraction Solvent									
Partition Step (if any)									
Clean up step									
Internal Standard (if any)									
Single Level Calibration									
Multi-Level Calibration									
Nº of points									
Column type									
Stationary phase									
Determination									
001	D	M	LC-MS/MS	0.01	97	10	6		
002/03	NA							Yes	4
004	D	S	LCMS	0.02	100	25	Acetone. Petroleumether. Dichloromethane		Phenomenex Aqua
005	NA							Yes	6
006	NA								100x2.1
007	D	M	GC-NPD	0.07	86	25	2		C18
008	ND	M		0.1		25	2	Yes	5
009	D	M	LC-MS/MS	0.01	90.0	50	4	Yes	5
010	NA						Yes		DB
011	NA								1701/05/01/35

OXYDEMETON-METHYL

		Lab Code			
		Quantification Method			
		Quantification using Standards in solvents or matrix			
		Confirmation method			
		RL (mg/kg)			
		Recovery			
		Sample Weight (g)			
		Extraction Solvent			
		Partition Step (if any)			
		Clean up step			
		Internal Standard (if any)			
		Single Level Calibration			
		Multi-Level Calibration			
		Nº of points			
		Column type			
		Stationary phase			
		Determination			
012	NA				
013	NA				
014	NA				
015	NA				
016	ND	S	0.02	50	3
017	ND	S	GC-MS	0.01	40
018	D	M		0.02	10
019	D	M	6 Transitions	0.005	95
020	D	M		0.010	10
021	D	M	LC-MS/MS	0.02	116.2
022	D	M		0.01	93
023	D	M	LC-MS/MS	0.01	10

OXYDEMETON-METHYL

Determination									
Stationary phase									
Column type									
Partion Step (if any)	Clean up step	Internal Standard (if any)	Single Level Calibration	Multi-Level Calibration	Nº of points				
Extraction Solvent									
Sample Weight (g)	Recovery								
RL (mg/kg)									
Confirmation method	Quantification using Standards in solvents or matrix	Quantification Method	Lab Code						
024 D M LC-MS/MS 0.005	10 Acetonitril	Yes	5	Aqua	C18	LC-MS/MS			
025 D M LC-MS/MS 0.008	20 6	Yes	2x4	HPLC(RP)	RP(C18)	LC-MS/MS			
026 NA									
027 NA									
028 D M GC-NPD 0.05 <70%	10 5 Others	Yes	7	K _S	HPSMS	GC-MS			
029 D M LC-MS/MS 0.01	10 6 LL	Yes	3-5	HPLC	Phenomenex Aqua	LC-MS/MS			
030 D M LC-MS 0.02	10 Methanol	Yes	3	Synergi-polar RP		LC-MS			
031 D S GC-MS 0.02	10 5 SPE	Yes	2	Capillary-LC	HP5 RP18	GC-ECD HPLC-DAD/ HPLC-FL			
032 D M LC-MS/MS 0.01	88.7 10 6(Methanol) 1:4. SPE	Yes	5	Aqua5μ	C18	LC-MS/MS			
033 NA									
034 NA									

APPENDIX 7. Methods used by participants for determining pesticides

OXYDEMETON-METHYL

Determination												
Stationary phase												
Column type												
Partion Step (if any)	Clean up step	Internal Standard (if any)	Single Level Calibration	Multi-Level Calibration	Nº of points	Column type	Stationary phase	Determination	Partion Step (if any)	Clean up step	Internal Standard (if any)	Single Level Calibration
Extraction Solvent	Sample Weight (g)	Recovery	Sample Weight (g)	Extraction Solvent	Partion Step (if any)	Clean up step	Internal Standard (if any)	Single Level Calibration	Extraction Solvent	Sample Weight (g)	Recovery	Sample Weight (g)
Partion Step (if any)	Clean up step	Internal Standard (if any)	Single Level Calibration	Multi-Level Calibration	Nº of points	Column type	Stationary phase	Determination	Partion Step (if any)	Clean up step	Internal Standard (if any)	Single Level Calibration
Sample Weight (g)	Extraction Solvent	Partion Step (if any)	Clean up step	Internal Standard (if any)	Single Level Calibration	Multi-Level Calibration	Nº of points	Column type	Stationary phase	Determination	Partion Step (if any)	Clean up step
RL (mg/kg)	Recovery	Sample Weight (g)	Extraction Solvent	Partion Step (if any)	Clean up step	Internal Standard (if any)	Single Level Calibration	Multi-Level Calibration	Nº of points	Column type	Stationary phase	Determination
Lab Code	Quantification Method	Quantification using Standards in solvents or matrix	Confirmation method	RL (mg/kg)	Recovery	Sample Weight (g)	Extraction Solvent	Partion Step (if any)	Clean up step	Internal Standard (if any)	Single Level Calibration	Multi-Level Calibration
035	D	M	GC-MS	0.02	50	3	GFC		Yes	4	HP-5MS	GC-MS/ECD/FPD
036	NA											
037	NA											
038	D	M	LC-MS/MS	0.01	90	20	2		Yes	3	RP18 LC	C18 Aquabμ
039	D	S		0.01	10	5	L	PSA	Methoxyfenozide	Yes	6	Aqua (phenomex)
040	ND	S	GC-ECD	0.01	340	2		O+GPC	Parathion	Yes		C18
041	D	M	LC-MS	0.02	90	10	Methanol					GC-ECD
042	D	M	GC-MS	0.05	100	2		GPC	EPN	Yes	3	Capillary
043	D	M	LC-MS/MS	0.006	10	6 (Methanol)		SPE		Yes	2	DB5, DB1701, DB35 (GC-ECD, GC-NPD, GC-FPD, GC-MS (single-quad))
044	D	M	GC-MS	0.02	100	2		GPC		Yes	5	OV17, DB6
045	D	M	GC-NPD, GC-MS	0.02	100	2		GFC		Yes	5	GC-NPD
												DB5 DB1701

OXYDEMETON-METHYL

Determination									
Stationary phase									
Column type									
Particule Step (if any)	Clean up step	Internal Standard (if any)	Single Level Calibration	Multi-Level Calibration	Nº of points	Column type	Stationary phase	Determination	
Extraction Solvent	Sample Weight (g)	Recovery	Sample Weight (g)	Extraction Solvent	Particule Step (if any)	Clean up step	Internal Standard (if any)	Single Level Calibration	Multi-Level Calibration
Partion Step (if any)	Sample Weight (g)	Recovery	Sample Weight (g)	Extraction Solvent	Particule Step (if any)	Clean up step	Internal Standard (if any)	Single Level Calibration	Multi-Level Calibration
Confirmation method	RL (mg/kg)	Recovery	Sample Weight (g)	Extraction Solvent	Particule Step (if any)	Clean up step	Internal Standard (if any)	Single Level Calibration	Multi-Level Calibration
Quantification using Standards in solvents or matrix	Quantification Method	Recovery	Sample Weight (g)	Extraction Solvent	Particule Step (if any)	Clean up step	Internal Standard (if any)	Single Level Calibration	Multi-Level Calibration
Lab Code	Quantification Method	Recovery	Sample Weight (g)	Extraction Solvent	Particule Step (if any)	Clean up step	Internal Standard (if any)	Single Level Calibration	Multi-Level Calibration
046 D M GC-FPD 0.1 102 50 3 GPC	047 D M GC-NPD 0.02 100 3 GPC	048 D S GC-FPD,MS 0.02 50 1 GPC	049 NA NA NA NA NA NA	050 NA NA NA NA NA NA	051 NA NA NA NA NA NA	052 NA NA NA NA NA NA	053 NA NA NA NA NA NA	054 NA NA NA NA NA NA	055 NA NA NA NA NA NA
056 NA NA NA NA NA NA	057 NA NA NA NA NA NA	058 NA NA NA NA NA NA	059 NA NA NA NA NA NA	060 NA NA NA NA NA NA					

APPENDIX 7. Methods used by participants for determining pesticides

071	070	069	068	067	996	065	064	063	062	061	Lab Code
NA	NA	NA	NA	NA	NA	ND	NA	NA	NA	ND	Quantification Method
											Quantification using Standards in solvents or matrix
											Confirmation method
						0.01				0.05	RL (mg/kg)
											Recovery
											Sample Weight (g)
											Extraction Solvent
											Partition Step (if any)
											Clean up step
											Internal Standard (if any)
											Single Level Calibration
											Multi-Level Calibration
											Nº of points
											Column type
											Stationary phase
											Determination

OXYDEMETON-METHYL

OXYDEMETON-METHYL

APPENDIX 7. Methods used by participants for determining pesticides

		Lab Code	
		Quantification Method	
		Quantification using Standards in solvents or matrix	
		Confirmation method	
		RL (mg/kg)	
		Recovery	
		Sample Weight (g)	
		Extraction Solvent	
		Partition Step (if any)	
		Clean up step	
		Internal Standard (if any)	
		Single Level Calibration	
		Multi-Level Calibration	
		Nº of points	
		Column type	
		Stationary phase	
		Determination	
NO RESULTS			
072	NA		
073	NA		
074			
075	NA		
076	D	M	GC-MS
076			0.02
076			10
076			5
076			Yes
076			O
077	ND		
077			0.05
078	D	M	GC-MS
078			0.05
078			60
078			50
079	NA		
079			1
079			GPC
080			
081	NA		
082	NA		
083	NA		
084	NA		

OXYDEMETON-METHYL

APPENDIX 7. Methods used by participants for determining pesticides

Lab Code									
Quantification Method									
Quantification using Standards in solvents or matrix									
Confirmation method									
RL (mg/kg)									
Recovery									
Sample Weight (g)									
Extraction Solvent									
Partition Step (if any)									
Clean up step									
Internal Standard (if any)									
Single Level Calibration									
Multi-Level Calibration									
Nº of points									
Column type									
Stationary phase									
Determination									
085	D	M	GC-MS	0.02	79	15	4	Yes	4
086	D	M		0.05	72	15	4	Yes	4
087	D	M	GC-MS	0.05		10	4	Yes	3
088	NA							HP-5	
089	NA	M							
090	ND	M	LC-MS/MS	0.2	79	15	1	TIP(QC)	Yes
091	NA								4
092	NA								3 µm
093	NA	S						C-18	LC-MS/MS
094	NA								
095	D	M	LC-MS/MS	0.01	88	75	1	Yes	LC-MS/MS

OXYDEMETON-METHYL

APPENDIX 7. Methods used by participants for determining pesticides

Determination											
Stationary phase											
Column type											
Particule size (µm)	Flow rate (ml/min)	Temperature (°C)	Wavelength (nm)	Detector	Wavelength (nm)						
103	ND	0.02	20	3	LL	Yes	2	HP-5	GC-ECD/NPD		
104	NA										
105	NA										
106	NA										
107	NA										
108	NA										
109	NA										
110	NA										

APPENDIX 7. Methods used by participants for determining pesticides

OXYDEMETON-METHYL

122	121	120	119	118	117	116	115	114	113	112	111	Lab Code
NA	Quantification Method											
												Quantification using Standards in solvents or matrix
								M				Confirmation method
												RL (mg/kg)
												Recovery
												Sample Weight (g)
												Extraction Solvent
												Partition Step (if any)
												Clean up step
												Internal Standard (if any)
												Single Level Calibration
												Multi-Level Calibration
												Nº of points
												Column type
												Stationary phase
												Determination

OXYDEMETON-METHYL

APPENDIX 7. Methods used by participants for determining pesticides

Determination											
Stationary phase											
Column type											
Partion Step (if any)	Clean up step	Internal Standard (if any)	Single Level Calibration	Multi-Level Calibration	Nº of points	Column type	Stationary phase	Determination	Quantification Method		
Extraction Solvent									Quantification using Standards in solvents or matrix		
Sample Weight (g)									Confirmation method		
Recovery									RL (mg/kg)		
Lab Code											
123	D	S	0.1	30	1	GPC	Yes	30mmx0.53mm	CP-Sil 8 CB	GC-NPD	
124	D	S	0.02	30	1				SGE	BPX-5	GC-NPD
125	D	S	GC-MS	0.02	110	25	1				
126	NO RESULTS										
127	NA										
128	D	S	GC-ECD	0.05	90	30	1				
129	NA										
130	D	M		0.02	30	1					

PROCYRIDONE

Lab Code	Quantification Method					
	Quantification using Standards in solvents or matrix					
Confirmation method						RL (mg/kg)
Recovery						
Sample Weight (g)						Extraction Solvent
Partition Step (if any)						
Clean up step						Internal Standard (if any)
Single Level Calibration						
Multi-Level Calibration						Nº of points
Column type						
Stationary phase						Determination
001 D S GC-MS 0.01 97 15 4	PCB97	Yes	3	60x0.25x0.1 60x0.25x0.15	DB5 DB1701	ECD NPD MS
002/0 03 D M GC-MS 0.05 93 15 4	Nitrofen+ Mirex	Yes	5	CAP	Restate CLP	GC-ECD
004 D GC-MS 0.02 31 20 6	SPE	Yes	5	90x0.83x0.5	RTX-WMA	GC-NPD
005 D M GC-MS 0.1 85 20 1						
006 NA						
007 D M GC-ECD 0.01 90 25 2	GPC	Yes	5	J&W 0.32/0.25mmID	DB5 DB1701	GC-ECD, GC-NPD, GC-ITD
008 D M	0.01 79 25 2	GPC	Yes	5	DB	1701/05/01/35
009 D M GC-MS 0.02 113.0 50 4 Yes	LL Aldrin Ditallowphos	Yes		Capillary	H-P-5 DB-1701P	NPD ECO
010 D M GC-EC 0.05 100 3 LL no	Aldrin	Yes	3	Capillary	DB 5 DB1701	GC-ECD GC-NPD
011 D S GC-MS 0.01 62 25 1 SPE		Yes	4	30mx0.53mmx1.5μm	DB5, DB608	GC-ECD, GC-MS

PROCYDIDONE

Determination									
Stationary phase									
Column type									
Partion Step (if any)	Clean up step	Internal Standard (if any)	Single Level Calibration	Multi-Level Calibration	Nº of points	Column type	Stationary phase	Determination	
Extraction Solvent									
Sample Weight (g)	Recovery								
RL (mg/kg)									
Confirmation method	Quantification using Standards in solvents or matrix	Quantification Method	Lab Code						
012 D M GC-MS 0.01 70-110 50 3	Ethy Acetate- Dichloromethane	SPE	Yes	8	HP5MS				GC-MS
013 D S GC-ELCD 0.01 78 25									GC-ECD/ELCD
014 D S GC-MS 0.02 40 3		LL	Yes	3	GC-Capillary	DB5			GC-ECD/NPD-MS
015 D S GC-MS 0.01 50 1/DCM (80/20)		SPE Florisil	Yes	3	Capillary 0.32	DB5, DB35			GC-ECD
016 D S 0.02 50 3	Yes	SPE	Yes	4	AT1701/AT1/DB05-MS	GC-ECD/PFPD/NPD/ MS			
017 D S GC-MS 0.01 40 2		GPC	Yes	5	DB5/DB1701				GC-ECD/PND
018 D S GC-MS 0.015 100 2		GPC	Yes	5	DB-5/1/1701				GC-ECD/NPD/FPD
019 D M GC-TOF 0.01 99 10 5		PCB 138	Yes	1	DB5				GC-MS
020 D M GC-MS 0.010 25 2		GPC	Yes	3	FSOT	HP-5-MS			GC-MS
021 D S GC-MS 0.01 98.2 50 2		GPC	Yes			BD1/5/35/1301. HP50/1701/5MS			GC-ECD/NPD
022 D S 0.02 97 50 6; Acetone	Clohexane/ Ethyl Acetate	GPC	Trans-HCEO	Yes	Capillary	DB5/210			GC-ECD
023 D M GC-MS 0.02 50 2		GPC		Yes	5	30mmx0.25mm			GC-MS

PROCYDIDONE

APPENDIX 7. Methods used by participants for determining pesticides

Lab Code									
Quantification Method									
Quantification using Standards in solvents or matrix									
Confirmation method									
RL (mg/kg)									
Recovery									
Sample Weight (g)									
Extraction Solvent									
Partition Step (if any)									
Clean up step									
Internal Standard (if any)									
Single Level Calibration									
Multi-Level Calibration									
Nº of points									
Column type									
Stationary phase									
Determination									
024	D	M	GC-MS/ECD	0.03	10	Acetonitril	PCB 209	Yes	9
025	D	M	GC/MS	0.01	50	3	GPC	Yes	3
026	D	S	GC-MS	0.02	10	5	SPE	Yes	3
027	D	M	GC-MS	0.02	100	2	GPC	EPSILON-HCH	Varian VF-5MS
028	D	M	GC-MSD	0.01	10 ⁴	10	5	Others	30mx0.25x0.25
029	D	M	GC-MS	0.005	100	2	Triphenyl-Phosphat	DB-5. Factor Four	DB-5. Factor Four
030	D	M	GC-ECD	0.01	50	6	GPC SPE	Yes	7
031	D	S	GC-MS	0.01	50	2	Bromophos-Ethyl	GC	HP5-MS DB17
032	D	M	GC-MS	0.01	94.1	30	Ethyl/Acetate/Ciclohexane (2:1)	Yes	3-5
033	D	M	GC-ECD	0.01	100	2	GPC	Yes	25m x 0.32mmx0.52μm
									Ultra2
									GC-ECD/NPD

PROCYMDONE

APPENDIX 7. Methods used by participants for determining pesticides

Determination											
Stationary phase											
Column type											
Partion Step (if any)	Clean up step	Internal Standard (if any)	Single Level Calibration	Multi-Level Calibration	Nº of points	Column type	Stationary phase	Determination	Partion Step (if any)	Clean up step	Internal Standard (if any)
Extraction Solvent	Sample Weight (g)	Recovery	Sample Weight (g)	Extraction Solvent	Partion Step (if any)	Clean up step	Internal Standard (if any)	Single Level Calibration	Multi-Level Calibration	Nº of points	Column type
Confirmation method	Quantification using Standards in solvents or matrix	Quantification Method	Lab Code	Lab Code	Partion Step (if any)	Clean up step	Internal Standard (if any)	Single Level Calibration	Multi-Level Calibration	Nº of points	Column type
034 D M GC-MS 0.01	100 2	PCB 153	Yes	2	30mx0.3mmx0.25µm	DB5 OJ1701	GC-ECD				
035 D M GC-MS 0.01	50 3	GPC	Yes	4	HP-5MS		GC-MS/ECD/FPD				
036 D S GC-MS 0.02	101 3	GPC	Yes				GC-ECD/NPD/MS				
037 D M GC-MS 0.02	100 2	GPC	Yes	4	HP5		GC-MS				
038 D M GC-MS 0.01	89 20	Triphenyl-Phosphat	Yes	3	Capillary-GC	DB17-MS	GC-MS				
039 D S GC-MS 0.02	45 3	Parathion	Yes	3	Restex 30x0.25x0.25	Rtx-CLPesticides I+II	GC-NPD				
040 D S GC-ECD 0.02	340 2	O+GPC	Parathion	Yes		Capillary	GC-ECD				
041 D S GC-MS 0.02	86 75	Acetone Followed by Cyclohexan and Ethyl Acetate	GPC	Yes	3	HP5					
042 D M GC-MS 0.01	100 2	GPC	EPN	Yes	3	Capillary	GC-ECD, GC-NPD, GC-ECD, GC-NPD, GC-FPD, GC-MS (single-quad)	DB5, DB1701, DB35			
043 D S GC-MS 0.01	50 3	LL	Yes			Capillary	DB5				
044 D M 0.01	100 2	GPC	Yes	5			DB5				GC-MS

PROCYDIDONE

APPENDIX 7. Methods used by participants for determining pesticides

Lab Code										
Quantification Method										
Quantification using Standards in solvents or matrix										
Confirmation method										
RL (mg/kg)										
Recovery										
Sample Weight (g)										
Extraction Solvent										
Partition Step (if any)										
Clean up step										
Internal Standard (if any)										
Single Level Calibration										
Multi-Level Calibration										
Nº of points										
Column type										
Stationary phase										
Determination										
045	D	M	GC-ECD. GC-NPD. GC-MS	0.02	100	2	GPC	Yes	5	DB5 DB1701 GC-ECD. GC- NPD.GC-MS
046	D	M	GC-ECD	0.007	101	50	3	GPC	Yes	Capillary DB17. HP5 GC
047	D	M	GC-ECD	0.005	100	2		GPC	Yes	3. 5 Capillary DB5, DB1701 GC-ECD. GC- NPD.GC-FFD
048	D	S	GC-ECD. MS	0.01	7	4	Yes	Yes	3	Capillary DB5 GC-ECD. GC-MS
049	D	M	GC-MS	0.05	79.7	15	4		2	DB5MS GC-ECD.GC-IT
050	NA									
051	D	M	GC-ECD	0.01	104	25	6	Yes	5	DB-5MS 95% Dymethyl- Polysiloxane GC-ECD
052	NA									
053	NA									
054	D	M	GC-ECD	0.02	90	25	6	Yes	0	Yes HP5MS 5%Phenyl GC-ECD
055	D	M	GC-ECD	0.02	113	15	4		5	HP1701 Me Siloxane GC-ECD
056	D	M	GC-ECD	0.1	93.75	15	4	GPC	Yes	5 HP5MS GC-MS
057	D	S	GC-MS	0.02	20	6		GPC- SPE Yes	3	DB5 Me Polisiloxan GC-MS
058	D	M	GC-MS	0.03	15	1		GPC Yes	3	HP5MS 5%PHSi GC-MS

PROCYDOME

APPENDIX 7. Methods used by participants for determining pesticides

Determination									
Stationary phase									
Column type									
Partion Step (if any)	Clean up step	Internal Standard (if any)	Single Level Calibration	Multi-Level Calibration	Nº of points	Column type	Stationary phase	Determination	
Extraction Solvent									
Sample Weight (g)									
Recovery									
RL (mg/kg)									
Confirmation method									
Quantification using Standards in solvents or matrix									
Quantification Method									
Lab Code									
059	D	S	GC-MS	0.02	5	1	SPÉ		Yes
060	D	S	GC-NPD	0.05	101.1	10	6(Dichloromethane)		Yes
061	D	M	GC-MS	0.02	81.7	15	6(Dichloromethane)		Yes
062	D	S	GC-ECD	0.01	110.2	50	Acetone+Methanol (1:1)	Empore Disk C8	Yes
063	D	S	GC-ECD	0.01	92	25	3	EXTRRELUT 20	Yes
064	D	S	GC-MS		111.3	10	6	O	Yes
065	D	S	GC-MS	0.005	103.5	10	Dichloromethane		Yes
066	D	S	GC-MS	0.05	104	25	5	Yes	SPÉ
067	D	M	GC-MS		10	1	EXTRRELUT	GPC	Yes
068	D	M	GC-MS	0.05	95.5	5	6	1	TFF
069	D	S	GC-MS		81	5	1	ASE(Dionex)	Ronnel+Azob Bencene+Triphenyl Phosphate
									5% Phenyl Silicone
									GC-ECD/NPD/FPD
									GC-MS/HPLC-FL
									MDN-SS

PROCYDOME

APPENDIX 7. Methods used by participants for determining pesticides

Determination										
Stationary phase										
Column type										
Nº of points										
Single Level Calibration										
Multi-Level Calibration										
Clean up step										
Internal Standard (if any)										
Partion Step (if any)										
Extraction Solvent										
Sample Weight (g)										
Recovery										
RL (mg/kg)										
Quantification using Standards in solvents or matrix										
Quantification Method										
Lab Code										
070	D	S	GC-ECD	0.01	15	6	GPC	M-series	Yes	25m ID 0.32 mm 0.25um
071	D	M	GC-MS	0.01	15	6	GPC		Yes	SE 54; OV1701
072	D	S		0.01	100	1	GPC	Diclofenon	Yes	HP1. HP5
073	D	S	GC-NPD80	0.01	10	Dichloromethane			Yes	SE30/SE54
074	NO RESULTS									
075	D	S	GC-MS	0.02	10	3	SPE	Fenchloraphos	Yes	DB5
076	D	M	GC-MS	0.01	10	5	Yes	O	Yes	HP-5MS
077	D	S	GC-MS	0.02	61	25	2	GPC	Yes	4
078	D	M	GC-MS	0.02	94	50	1	GPC	Yes	Narrowbore
079	D	M	GC-MS	0.02	15	4			Yes	1
080	NO RESULTS									
081	D	M	GC-MS	0.01	20	1			Yes	3-6
082	D	M	GC-MS	0.05	15	4			Yes	HP-5MS
										5% Ph. Me S
										GC-ECD/FID/TD
										GC-ECD

PROCYDOME

Determination									
Stationary phase									
Column type									
Partion Step (if any)	Clean up step	Internal Standard (if any)	Single Level Calibration	Multi-Level Calibration	Nº of points	Column type	Stationary phase	Determination	Procymidone
Extraction Solvent	Sample Weight (g)	Recovery	RL (mg/kg)	Lab Code	Quantification Method	Quantification using Standards in solvents or matrix	Confirmation method	Lab Code	Procymidone
LL Phase Separation	Sample Weight (g)	Recovery	RL (mg/kg)	083 D M GC-MS 0.005	15	4	Varian CP 3800	Low Bleed Fused Silica	GC-MS
084 D M GC-MS/MS 0.01	10	1	C/P-Sil 8	085 D M GC-MS 0.02	98	15	4	PhMSil 5%	GC-MS/MS
086 D M GC-MS/MS 0.02	86	15	4	087 D M GC-MS 0.02	10	4	30x25x25	DB-5,DB-17	GC/FPD/ECD/MS
088 D M MS 0.03	15	4	HP-5	089 D M GC-MS/MS 0.02	15	4	NarrowBore	HP5-MS	GC-ECD/NPD/FPD/MS
090 D M GC-MS/MS 0.02	86	15	1	TPP(QC)	Yes	3	Capillary	BP5	GC-MS/MS
091 D M GC-MS/MS 0.05	15	1	TIP. Anthracene	092 D M GC-MS/MS 0.02	5.2	4	HP-5MS	DB-5 ms	GC-MS/MS
093 D S GC-MS 0.02	15	4	Factor Four	094 D M MS/MS	15	4	GC-ITD	GC-MS/MS-ECD	GC-ECD/GC-NPD/GC-FPD
			GC-MS				GC-MSL8CB	CPSIL8	GC-ECD/GC-NPD/GC-FPD
			30mx0.25x0.2						

PROCYDIDONE

APPENDIX 7. Methods used by participants for determining pesticides

Lab Code									
Quantification Method									
Quantification using Standards in solvents or matrix									
Confirmation method									
RL (mg/kg)									
Recovery									
Sample Weight (g)									
Extraction Solvent									
Partition Step (if any)									
Clean up step									
Internal Standard (if any)									
Single Level Calibration									
Multi-Level Calibration									
Nº of points									
Column type									
Stationary phase									
Determination									
095	D	M	GC-MS	0.02	85	75	1	GPC	Yes
096	D	M	GC-ITD	0.05	108	15	4		Yes
097	D	M	GC-MS	0.02	98	30	1	GPC	Triphenyl Phosphate
098	D	M	GC-MS	0.02		10	1	SPE	Yes
099	D	M	GC-MS	0.01	81	30	1	O	
100	D	M	GC-MS	0.02	87	10	1	GPC	
101	D	M	GC-MS	0.02	96	50	5	SPE	Yes
102	D	M	GC-ITD	0.01	95	50	1	GPC	Yes
103	D	S	GC-MS	0.02	91	20	3	LL	
104	D	M	GC-MS	0.01	109	75	1	1/20	GPC ALDRIN
105	NA								
106	D	M	GC-MS	0.02	7.5	1		Ethyl-Parathion	Yes
107	D	M	GC-MS	0.02	90.60	20	4		
108	D	S	GC-ECD	0.02	95	5	3		
109	D	S	GC-EC	0	91	100	3		Yes

PROCYDOME

APPENDIX 7. Methods used by participants for determining pesticides

Lab Code	Quantification Method		Confirmation method		RL (mg/kg)	Recovery	Sample Weight (g)	Extraction Solvent	Partition Step (if any)	Clean up step	Internal Standard (if any)	Single Level Calibration	Multi-Level Calibration	Nº of points	Column type	Stationary phase	Determination		
	Quantification using Standards in solvents or matrix																		
110	D	M	GC-MS	0.01	86.80	20	3	SPE		Yes	5	DB-5				GC-ECD,GC-NPD			
111	D	S		0.05	85.50	10	4	SPE (Silica)		Yes	5	HP-5MS	5%Phenyl				GC-ECD/NPD		
112	D	S	GC-NPD	0.02	95	100	3	O		Yes			HP1	MeSilicone			GC-ECD		
113	D	M	GC-TID	0.025	84	15	4	NO	Hexachloro-Benzene	Yes	5	DB-5MS				GC-TID			
114	D	S	GC-ECD	0.02		18	1	GPC		Yes	4	Capillary	HP5			GC-ECD			
115	D		GC/MS	0.02		50	4	Dichloro-methane	LL	Yes		HP(30x0.25x0.25)	5MS			GC/MS			
116	D	S	GC/MS	0.01	114	50	2	SPE		Yes			HP5-MS				GC/MS		
117	D	M	GC-MS	0.02	108.1	20	4	Yes	GPC		Yes					GC/MS			
118	D	M	GC-MS	0.02	95	10	6	SPE	Yes	Yes	3	DB-35MS	35%Phenyl			GC-MS			
119	D	M	GC-MS	0.01		10	1	NO		Yes		Capillary	DB-5MS			GC-MS			
120	NA																		
121	D	M	GC-MS	0.01		25	1	GPC		Yes	5	Capillary	DB-5MS	(single quad. GC-MS ECD,NPD)					

PROCYRIDONE

APPENDIX 7. Methods used by participants for determining pesticides

Determination													
Stationary phase													
Column type													
Lab Code	Quantification Method	Quantification using Standards in solvents or matrix	Confirmation method	RL (mg/kg)	Recovery	Sample Weight (g)	Extraction Solvent	Partition Step (if any)	Clean up step	Internal Standard (if any)	Single Level Calibration	Multi-Level Calibration	Nº of points
122 D S GC-MS 0.02	85.40	30	1	GPC		Yes	3	Chromopack 50mmx0.25mm x0.25mm	CPSL-8CB	GC-ECD			
123 D S GC-MS 0.02		30	1	GPC		Yes		30mmx0.32mm	CP-SII 8 CB	GC-ECD			
124 D S 0.02		30	1	GPC		Yes	3	Restek	STX-CLP	GC-ECD			
125 D S GC-MS 0.02	94	25	1	GPC		Yes	5	Capillary	RTX 17025	GC-ECD			
126 NO RESULTS													
127 D S GC-MS 0.01		30	1	GPC		Yes	3	50mmx0.32mm x0.25μm	CP-SII 8 CB	GC-ECD			
128 D S GC-ECD 0.02	106	30	1	GPC		Yes	3	30mmx0.25mm x0.25μm	RTX-CLP	GC-NPD			
129 D S 0.02		30	1	GPC		Yes	3	Restek	Stx-Cl-Pest.	GC-ECD			
130 D M GC-MS 0.05		30	1	GPC		Yes	3	Restek	RTX-5MS	GC-ECD			

APPENDIX 7. Methods used by participants for determining pesticides

THIABENDAZOLE

Lab Code	Quantification Method						Determination
	Confirmation method						
001	D	M	LC-MS/MS	0.05	97	15	RL (mg/kg)
002/0 03	D	S	HPLC-DAD	0.1	122	15	Recovery
004	D		LCMS	0.1	100	50	Sample Weight (g)
005	D	S		0.04	90	50	Extraction Solvent
006	0.22	M	LC-MS/MS	0.05	80	10	Partition Step (if any)
007	D	M	LC-MS/MS	0.01	86	10	Clean up step
008	D	S		0.05	90	25	Internal Standard (if any)
009	D	M	LC-MS/MS	0.01	97.0	50	Single level calibration
010	ND			0.5	100	3	Multi-level calibration
011	D	S	GC-MS	0.01	70	77	Nº of points
							Column type
							Stationary phase

APPENDIX 7. Methods used by participants for determining pesticides

THIABENDAZOLE

Determination											
Stationary phase											
Column type											
Partion Step (if any)	Clean up step	Internal Standard (if any)	Nº of points	Column type	Stationary phase	Determination					
Extraction Solvent	Sample Weight (g)	Recovery									
Partion Step (if any)	Clean up step	Internal Standard (if any)	Nº of points	Column type	Stationary phase	Determination					
Sample Weight (g)	Recovery	Extraction Solvent									
RL (mg/kg)											
Quantification Method	Quantification using Standards in solvents or matrix	Confirmation method									
Lab Code											
012	D	S	UVSCAN	0.01	70-110	50	1			Yes	6
013	D	S	HPLC-DAD	0.02	90	25	1				
014	D	S	HPLC-UV	0.05	75	1		LL	Yes		C18
015	D	S		0.04	25	4 (1/1/1)					HPLC-DAD
016	D	S		0.02	10	4		SPE	Yes	2	150x6x6µm
017	D	S	GC-MS	0.01	100	2		LL			DE613 Shodex
018	D	M	GC-MS	0.01	10	5		SPE	Yes	4	SHODEX DF-613 Polyalkyl Metacrylate
019	D	M	2 Transitions	0.005	93	10	5			5	250mmx4mm RP-18
020	D	M	LC-MS/MS	0.010	10	6					RP-18
021	d	s	LC-DAD	0.02	79.3	25	6	SPE	Yes	3	Aqua 3 C18
022	D	M		0.01	93	10	6 ₁ (Methanol)	SPE:LL	Yes	7	LICHROSPHER 100RP8 150mmx4.6mm. 5µm HPLC-FL
023	D	M	LC-MS/MS	0.01	10	10	Methanol	CEM ELUT	Yes	7	2.1x125mm; 3µm RP-C18 LC-MS/MS
									Yes	5	2.1x100mm Atlantis C18 LC-MS/MS

THIABENDAZOLE

APPENDIX 7. Methods used by participants for determining pesticides

Determination									
Stationary phase									
Column type									
Partion Step (if any)									Clean up step
Extraction Solvent									Internal Standard (if any)
Sample Weight (g)									Single level calibration
Recovery									Multi-level calibration
RL (mg/kg)									Nº of points
Confirmation method									Column type
Quantification Method									Internal Standard (if any)
Quantification using Standards in solvents or matrix									Single level calibration
Lab Code	D	M	LC-MS/MS	0.005	10	Acetonitril			
024	D	M	LC-MS/MS	0.005	10	Acetonitril		Yes	5
025	D	M	LC-MS/MS	0.006	20	6		Yes	2x4
026	NA								HPLC(RP)
027	D	M	GC-MS	0.05	100	2			RPC(C18)
028	D	S	HPLC-FL	0.05	92.4	10	1		
029	D	M	GC-MS	0.02	100	2	1	Yes	7
030	D	M	GC-MS	0.02	20	Acetone		Yes	3-5
031	D	S	GC-MS	0.05	30	4		Yes	4
032	D	M	LC-MS/MS	0.01	94.2	10	6(Methanol)		Capillary
033	D	S	HPLC-JV	0.03	100	2		Yes	3

APPENDIX 7. Methods used by participants for determining pesticides

THIABENDAZOLE

Determination									
Stationary phase									
Column type									
Partion Step (if any)	Clean up step	Internal Standard (if any)	Single level calibration	Multi-level calibration	Nº of points	Column type	Stationary phase	Determination	
Extraction Solvent									
Sample Weight (g)									
Recovery									
RL (mg/kg)									
Confirmation method									
Quantification using Standards in solvents or matrix									
Quantification Method									
Lab Code									
034	D	S	HPLC-F	0.05	100	2	GPC LL	Yes	250x4x5
035	D	M	LC-MS/MS	0.01	10	5	O	Yes	5 Aqua 5µ. 50x2mm
036	NA								RP18
037	D	S	GC-MS	0.05	100	2	GPC	Yes	HP5
038	D	M	LC-MS/MS	0.01	73	20	Triphenyl- Phosphat	Yes	GC-MS
039	D	S		0.01	10	5	PSA Methoxi- Fenoizide	Yes	LC-MS/MS
040	NA								Aqua (phenomex)
041	D	M	LC-MS	0.05	125	10	Atlantis	Yes	C18
042	D	M	GC-MS	0.05	100	2	GC-ECD, GC-NPD, GC-FPD, GC-MS (single-quad)	Yes	LC-MS/MS
043	D	M	LC-MS/MS	0.006	10	6 (Methanol)	Capillary	3	DB5, DB1701, DB35
044	D	M		0.01	100	2	SPE	Yes	C18
								4	RP8
									LC-MS/MS

APPENDIX 7. Methods used by participants for determining pesticides

THIABENDAZOLE

Determination									
Stationary phase									
Column type									
Partion Step (if any)	Clean up step	Internal Standard (if any)	Single level calibration	Multi-level calibration	Nº of points				
Extraction Solvent									
Sample Weight (g)									
Recovery									
RL (mg/kg)									
Confirmation method									
Quantification using Standards in solvents or matrix									
Quantification Method									
Lab Code									
045	D	M	LC-MS/MS	0.05	10	5	SPE	Yes	3
046	D	M	GC-PND	0.05	105	50	3	Yes	Luna
047	D	S	HPLC-DAD	0.02	50	2	LL	Yes	C18
048	NA								LC-MS/MS
049	NA								GPC
050	ND			0.1	95	25	1	Yes	Capillary
051	NA								DB17. HP5
052	NA								GC
053	NA								HPLC-DAD
054	ND			0.1	25	1.6	O	Yes	RP18-5μ
055	NA								
056	D	S	HPLC-DAD	0.05	103.7	15	4	Yes	
057	D	S	HPLC-UV	0.04	20	1	LL	Yes	Shodexde131
058	ND	M	GC-MS	0.03	15	1	GPC	Yes	P/M/A Gel
									HPLC-DAD
									HP5MS
									5%PHSi
									GC-MS

APPENDIX 7. Methods used by participants for determining pesticides

THIABENDAZOLE

Determination									
Stationary phase									
Column type									
059	NA								
060	NA								
061	D	M	GC-MS	0.05	72.6	15	6(Dichloromethane)		
062	ND	S	HPLC-DAD	0.05	71.2	10	Dichloromethane	LC-NH ₂ Columns	Yes
063	D	S	GC-NPD	0.05	85	50	1	L-L (Acid- Base Partition)	Yes
064	D	S	GC-MS		76	50	1	LL	Yes
065	D	S	GC-MS	0.02	85.4	10	Dichloromethane		Yes
066	D	S		0.02	75	25	5		Yes
067	D	M	HPLC-DAD		10	1	EXTRRELUT	GPC	Yes
068	D	M	GC-MS	0.05	100	5	6	TF	Yes
069	NA							C1P SIL 8CB	5
								FUSED SILICA	
								GC-MS/HPLC-FL	

APPENDIX 7. Methods used by participants for determining pesticides

THIABENDAZOLE

Determination											
Stationary phase											
Column type											
Lab Code	Clean up step										
Quantification Method	Internal Standard (if any)										
Quantification using Standards in solvents or matrix	Single level calibration										
Confirmation method	Multi-level calibration										
RL (mg/kg)	Nº of points										
Recovery	Column type										
Sample Weight (g)	Stationary phase										
Extraction Solvent	Determination										
Partition Step (if any)	Determination										
Clean up step	Determination										
Internal Standard (if any)	Determination										
Single level calibration	Determination										
Multi-level calibration	Determination										
Nº of points	Determination										
Column type	Determination										
Stationary phase	Determination										
Determination	Determination										
NO RESULTS											
070	D	S	GC-MS	0.05	10	3	SPE	Fenchlorphos	Yes		
071	D	M	HPLC-FL	0.01	50	1	LL		Yes	5	250mmx4.6. 5um
072	D	S		0.02	110	1	LL		Yes	6	150mmx4.6. 5um
073	NA						GPC	Diclofenon	Yes		PILRP-S
074											HPLC-UV
075	D	S	GC-MS	0.05	10	3	SPE	Fenchlorphos	Yes		
076	D	M	GC-MS	0.1	10	5	Yes	O	Yes	3	HP-5MS
077	D	S	GC-MS	0.5	10	25	2	GPC	Yes	4	HP-5MS
078	D	M	GC-MS	0.1	50	50	1	GPC	Yes	4	5%Phenyl-Methyl Siloxane
079	NA										Narrowbore
080											HP 50
081	NA										GC-NPD
082	NA										GC-MS

APPENDIX 7. Methods used by participants for determining pesticides

THIABENDAZOLE

Determination									
Stationary phase									
Column type									
083	NA								
084	NA								
085	D	M	GC-MS	0.04	79	15	4	Yes	4
086	D	M	LC-MS/MS	0.05	95	15	4	Yes	4
087	D	S	HPLC-DAD	0.03		10	1	pH Changes	30x25x25
088	NA								DB-5, DB-17
089	NA								GC/FPD/ECDD/MS
090	D	M	LC-MS/MS	0.05	92	15	1	TTP(QC)	HPLC-DAD
091	NA								
092	NA								
093	D	S	HPLC-UV	0.05	15	4		Yes	4
094	NA								30m/25x25
									CPSIL8CB
									GC-MS

THIABENDAZOLE

Determination									
Stationary phase									
Column type									
Partion Step (if any)	Clean up step	Internal Standard (if any)	Single level calibration	Multi-level calibration	Nº of points				
Extraction Solvent	Sample Weight (g)	Recovery	Sample Weight (g)	Extraction Solvent	Partion Step (if any)	Internal Standard (if any)	Single level calibration	Multi-level calibration	Nº of points
Confirmation method	RL (mg/kg)	Quantification using Standards in solvents or matrix	Quantification Method	Lab Code	Lab Code	Internal Standard (if any)	Single level calibration	Multi-level calibration	Nº of points
LC-MS/MS	LC-MS/MS	LC-MS/MS	LC-MS/MS	LC-MS/MS	LC-MS/MS	RP-HPLC	RP-HPLC	RP-HPLC	RP-HPLC
Altima C18.5mm	Altima C18.5mm	Altima C18.5mm	Altima C18.5mm	Altima C18.5mm	Altima C18.5mm	C-18	C-18	C-18	C-18
LC-MS/MS	LC-MS/MS	LC-MS/MS	LC-MS/MS	LC-MS/MS	LC-MS/MS	30%oPhenyl	30%oPhenyl	30%oPhenyl	30%oPhenyl
LC-MS/MS	LC-MS/MS	LC-MS/MS	LC-MS/MS	LC-MS/MS	LC-MS/MS	ZB5	ZB5	ZB5	ZB5
GC-MS	GC-MS	GC-MS	GC-MS	GC-MS	GC-MS	C-18	C-18	C-18	C-18
GC-MS/MS	GC-MS/MS	GC-MS/MS	GC-MS/MS	GC-MS/MS	GC-MS/MS	HP 5/HP 50	HP 5/HP 50	HP 5/HP 50	HP 5/HP 50
See Column Type	See Column Type	See Column Type	See Column Type	See Column Type	See Column Type	GC-ECD,GC-NPD	GC-ECD,GC-NPD	GC-ECD,GC-NPD	GC-ECD,GC-NPD
ODS2	ODS2	ODS2	ODS2	ODS2	ODS2	C-18	C-18	C-18	C-18
HPLC/DAD	HPLC/DAD	HPLC/DAD	HPLC/DAD	HPLC/DAD	HPLC/DAD	A ^T -5MS	A ^T -5MS	A ^T -5MS	A ^T -5MS
Narrowbore	Narrowbore	Narrowbore	Narrowbore	Narrowbore	Narrowbore	GC-MS	GC-MS	GC-MS	GC-MS
GC-MS	GC-MS	GC-MS	GC-MS	GC-MS	GC-MS	C-18	C-18	C-18	C-18
L C-MS/MS	L C-MS/MS	L C-MS/MS	L C-MS/MS	L C-MS/MS	L C-MS/MS	X-terra 100 mm	X-terra 100 mm	X-terra 100 mm	X-terra 100 mm
LC-MS	LC-MS	LC-MS	LC-MS	LC-MS	LC-MS	C-18	C-18	C-18	C-18
NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
109	NA	NA	NA	NA	NA	NA	NA	NA	NA
108	NA	NA	NA	NA	NA	NA	NA	NA	NA
107	D	S	LC-MS	0.05	85.50	20	4	Yes	6
106	D	M	LC-MS/MS	0.05	7.5	1		Yes	C-18
105	NA	NA	NA	NA	NA	NA	NA	NA	NA
104	D	M	GC-MS	0.01	77	75	1	Yes	C-18
103	D	S	HPLC/DAD	0.05	89	50	1	Yes	C-18
102	NA	NA	NA	NA	NA	NA	NA	NA	NA
101	ND	M	GC-MS	0.1		50	5	Yes	HP 5/HP 50
100	D	M	LC-MS/MS	0.05	87	10	1	Yes	ZB5
99	D	M	GC-MS	0.05	65	30	1	Yes	4
98	D	M	LC-MS/MS	0.05	10	1		Yes	4
97	D	M	LC-MS/MS	0.02	89	10	5	Yes	5
96	D	S	GC-ITD	0.01	97	7.5	4(+Na ₂ SO ₄)	Yes	4
95	D	M	LC-MS/MS	0.01	108	75	1	Yes	4

APPENDIX 7. Methods used by participants for determining pesticides

THIABENDAZOLE

APPENDIX 7. Methods used by participants for determining pesticides

Lab Code	Quantification Method					
	Quantification using Standards in solvents or matrix					
Confirmation method						
110	RL (mg/kg)					
	Recovery					
111	Sample Weight (g)					
	Extraction Solvent					
112	Partition Step (if any)					
	Clean up step					
113	Internal Standard (if any)					
	Single level calibration					
114	Multi-level calibration					
	Nº of points					
115	Column type					
	Stationary phase					
116	Determination					
117	ND	M	GC-MS	0.05	112.3	20
118	D	M	GC-MS	0.02	92	10
119	D	M	LC-MS/MS	0.01	10	5
120	NA					
121	D	M	LC-MS	0.001	12.5	5

APPENDIX 7. Methods used by participants for determining pesticides

THIABENDAZOLE

Protocol for the European Proficiency Test 06



<http://www.ual.es/GruposInv/EUPT6>



Instructions

To be able to participate in the European Proficiency Test 6 it is essential that you have returned via fax or e-mail the application form. The Organisation will **ONLY** send samples to laboratories that have completed and returned their application forms.

Once the completed application form has been received, the Organisation will send a unique **LABORATORY CODE** to each participant via e-mail. This code must be used by the laboratories when returning results and when communicating with members of the Organising Team. This code will only be known by the participant, the Organising Team and the Commission.

Together with this code, the protocol will be sent and 3 forms (**Form 1, 2 and 3**). The participant laboratories will need these forms during the course of the proficiency test.

Each one has a reporting deadline; please ensure you adhere strictly to these deadline. The reporting forms must be sent directly to the Organiser. The official language used in this Proficiency Test will be English.

Communications between participating laboratories during the test are forbidden unless they report combined official data.

A list of all the possible pesticides that could be present in the test material is enclosed with this protocol. The MPRLs given in this list, refer to the "lower limit of analytical determination" given in the European Commission's Directive for MRLs. For pesticides with a residue definition that includes metabolites/degradation products, there is only one MPRL in the list, the one for the parent compound.

General Characteristics

Objectives

The objective of the proficiency test is to obtain information of the quality, accuracy and comparability of the pesticide residue data sent to the Commission. Participating laboratories will be provided with an assessment of their analytical performance and the reliability of their data compared to other laboratories.

ANNEX 1. Protocol and instructions. List of pesticides to be sought.

Steps to Follow

The Proficiency Test is made up of the following 8 steps that are essential for the generation of satisfactory results:

1. Preparation of the test materials. Homogeneity and stability testing performed by the Organisation.
2. Return of the application forms by the participating laboratories.
3. Production of a detailed Protocol for the test by the Organisers.
4. Distribution of a code to each participant to allow them access to the protocol and the forms.
5. Shipment of the test material, together with the blank.
6. The participant laboratories are responsible for reporting their data to the Organiser using the forms supplied by the stipulated deadline.
7. The Organiser will evaluate the results at the end, once the deadline has passed.
8. The Organiser will send a Final Report to each participant laboratory. This report will include information regarding the design of the test, the homogeneity and stability test results, a record of the shipped samples, a statistical evaluation of the participant's results, graphical displays of the results and conclusions. Any other relevant information considered of interest will also be included.

Evaluation of the Results

The statistics used for the treatment and assessment of the data will be described in detail in the Final Report. A short summary of how the results will be treated is given below.

The results could be:

– ***False Positives***

These are the results that show the presence of pesticides that were not actually present in the test material. However if a number of laboratories detect the same additional pesticide or if the concentrations are close to the reporting levels, decisions will be taken on a case-by-case basis.

– ***False Negatives***

These are results for pesticides that were not reported but have been shown by the majority of laboratories to be present in the test material, and were present at level higher than the Reporting Limit (RL) reported by the laboratory.

ANNEX 1. Protocol and instructions. List of pesticides to be sought.

– *Establishing the true concentration (μ)*

The true concentration in all cases will be determined by the median of the results (evaluated as valid, following application of a test for outliers). Therefore there will be a median value for every pesticide present.

– *Establishing the assigned value for the standard deviation*

The assigned standard value (δ) will be fixed by the Organiser.

$$\text{Where } \delta = b_i * \mu_i \quad \text{being } b_i = \% \text{FFP}/100$$

An assigned value will be established based on the Fit-For-Purpose (FFP) Standard Deviation model. An average fixed value of 25% will be chosen. The Organizer may increase this value for certain difficult pesticide-crop-concentration combinations, after consultation with the committee of experts and based on the experience of passed Proficiency Tests results.

Other tests, to calculate a target standard deviation, such as Horwitz and robust statistics (Qn method), will also be used in parallel for comparison.

– *z-Scores*

This parameter is calculated using this formula:

$$z_i = (x_i - \mu_i) / \delta_i$$

where x_i is the value reported by the laboratories, μ_i the assigned value and δ_i the standard deviation of the concentration, for each pesticide.

z-Score value will be interpreted in the following way:

$$\begin{aligned} |z| \leq 2 & \text{ acceptable} \\ 2 < |z| \leq 3 & \text{ questionable} \\ |z| > 3 & \text{ unacceptable} \end{aligned}$$

The values considered as false negative results will be assigned a value equal to the Reporting Limit given by the laboratory, resulting a z-Score value not greater than 3.5. A z-Score will not be assigned to false positive results. The organizer will consider whether these values should appear in the graphs or not.

– *Combined z-Score values*

In order to evaluate each laboratory's performance, taking into account all the pesticides analysed, two ways will be used to combine the z-Scores obtained: the re-scaled sum of z-Scores (RSZ) and the sum of squared z-Scores (SSZ).

Laboratories may be excluded from calculation of these parameters depending on their minimum number of pesticide sought and/or reported.

The equations are:

$$\begin{aligned} \text{RSZ} &= \sum z / (m)^{1/2} \\ \text{SSZ} &= \sum z^2 \end{aligned}$$

ANNEX 1. Protocol and instructions. List of pesticides to be sought.

These summed z-Score results are considered to be less important than the individual z-Scores. Therefore the organizer retains the right to not use them if he feels they are not helpful. Besides, a descriptive performance evaluation will be worked out, based on scope of analysis, number of false positives and false negatives and number of too high Reporting Limit.

Organisation Address

The official postal address of the organizer, phone number, fax number and e-mail are as follows:

**Universidad de Almería
Edificio Químicas CITE I
Ctra. Sacramento s/n
04120 Almería - Spain**

Phone Number: 00 34 950015034/ 0034950015645

Fax Number: 00 34 950015645

E-mail: pmedina@ual.es or amadeo@ual.es

On-Line News

The latest information will currently be updated and found on the web address:

www.ual.es/GruposInv/EUPT6/

European Proficiency Test 06

Introduction

This proficiency test is based on pesticides residues analysis in tomatoes. The tomatoes were grown in Almería.

The pesticide treatments were carried out with formulated aerosols. An additional spiking in the laboratory following harvest of the crop was required for some pesticides.

The samples were chopped, frozen (using liquid nitrogen), ground, mixed thoroughly and sub-sampled into polyethylene bottles.

For the homogeneity test, 11 of these bottles, now containing the test material, will be randomly chosen and analysed by an independent laboratory to check for homogeneity.

The material, will be stored frozen (-20°C) prior to shipment to participants.

Two bottles, again randomly chosen, will be analysed in over a time period to confirm the stability of the pesticides in the test material (firstly when the test materials are shipped, and then a few days after the deadline for receipt of results from the participants). These results will not be included in the statistical analysis of the test.

The aim is only to check the stability during the shipping process and the duration of the test.

Calendar

The following table shows the program for this EUPT 06

Activity	Date
- Selection of pesticides/crop combinations to be tested. - Protocol. Selection of characteristics of the web page.	1 st March 2004
- Selection of participating laboratories and contact points in each country.	16 th April 2004
- Growing crop and field treatment.	1 st May 2004
- Spiking (if needed), homogenisation, and storage stability testing.	10 th May 2004

ANNEX 1. Protocol and instructions. List of pesticides to be sought.

Activity	Date
- Distribution of the protocol.	11 th May 2004
- Sample distribution.	17 th -18 th May 2004
- Deadline for receiving results by the organizers.	7 th July 2004
- Preliminary Report.	13 th September 2004
- Final Report.	12 th October 2004

Participant Laboratories

It is up to the contact points of each country to choose the laboratories that should participate, although it is a requirement that a laboratory should contribute results to the national monitoring program of a EU Member State. It is up to those participants to fill in and return the application form so the organiser has all their details before the deadline. The Organisation will not be responsible if a laboratory does not get notification of the web page address and the protocol necessary to take part in the test.

Amount of Sample

About 400g of tomato test material will be shipped together with 400g of 'blank' tomato packed in boxes surrounded with dry ice. The courier costs are charged to the participants. In case of the receipt of a spoiled test material, it will be possible to request a replacement. However, again, the courier cost will still be charged to participants. Form 1 should be used to request additional test material and an explanation for the reasons for the request must be given. There will only be a limited amount of test sample and laboratories should not ask for more than they need.

Application Form (using the EUPT06 web page)

Using the web page (www.ual.es/GruposInv/EUPT6/), the participating laboratories should fully complete the application form and send it via fax or mail.

Shipping of Samples

The shipment of samples will follow an e-mail message to participants warning that the test material is about to be dispatched (or fax if no mail has been given). This could vary depending on the country and the use, or not, of delivery points in some specific countries. The time to reach each laboratory will vary according to the location of the country and the town (from 2-5 days).

Form 1

Once the laboratory has received the test materials they must complete Form 1, filling in the date of reception, and any other observation that they consider important about the adverse condition of the sample. Form 1 must then be sent by fax or e-mail within two days of receipt of the test materials otherwise, the organizer will assume that the shipment and the reception was satisfactory. Please note that you must include the laboratory code assigned to you on this form.

Analyses And Results Forms (Form 2)

Significant Decimals

The results will be expressed in mg/kg in the following way:

- concentrations <0.100 mg/kg, to be expressed with two significant figures (three decimals, i.e. 0.058 mg/kg).
- concentrations > 0.100 mg/kg, to be expressed with three significant figures, i.e. 0.156, 1.64, 10.3 mg/kg.

In cases where a pesticide has not been detected, its concentration should be expressed as < x mg/kg, where x is the Reporting Limit, RL, (of the laboratory), expressed to one significant figure.

Correction of Results

The results should **not** be corrected using recovery factors. If the laboratory usually corrects the results for the recoveries, they must provide the correction factor used for each pesticide. This information must be sent together with the results in Form 2.

Samples Material for Analysis (Form 2)

The test material contains a certain number of pesticides form the List of Pesticides. It should not be assumed that only pesticides registered for use on tomatoes are present.

Each laboratory must undertake only **ONE** analysis for each pesticide and only report **ONE** result for the pesticides in the sample material, using their normal routine analytical procedures. This do not exclude that more than one method has to be used to cover all compounds present.

The analytical procedure used must be reported using Form 2. The results, expressed as concentration levels in mg/kg, (whether it has been used calculated from a standard in pure solvent or in a matrix extract) must also be reported together with the limits of quantification for each pesticide.

This form must be sent to the Organiser before 7th of July. Results received after this date will not be included in the statistical treatment, or in the final report.

Analytical Procedures Used (Form 3)

A brief summary of the analytical procedures used is required from each laboratory on Form 3.

If more than one method has been used, please mark them with different letters or codes in Form 2 and use as many copies of Form 3 as are needed (one for each method).

The organizer must receive Form 3 by mail or by fax before 7th July 2.004.

Sample Manipulation Advises

The test material should remain frozen until it is analysed.

- Allow the test material to defrost in the refrigerator the afternoon before the analysis.
- Once defrosted, be sure to mix the content in the bottle thoroughly, to ensure homogeneity of the contents, before taking the analytical portion.

Annexes

- Form 1
- Form 2
- Form 3
- List of Pesticides

FORM 1

Laboratory Code:

Test material code:

Date of reception:

*(check the bottle for the blank
and the test material)*

EUPT06-Lab-

EUPT06-blank-

EUPT06-sample-

<input type="text"/>	<input type="text"/>
----------------------	----------------------

Observation about the condition of the sample (either the test material or the blank):

Loses:

YES

NO

Frozen:

YES

NO

Please, fill in this form and send it back by fax (0034950015645) or e-mail (pmedina@ual.es) once you have received the test material.

Date:

Signature:

FORM 2 (RESULTS)

Laboratory Code: Date:

Test material code:

Pesticide	Quantification Method(1)	Analytical Procedure(2)	Conc. (mg/kg)(3)	Quantification Using Standards in solvents or matrix (4)	Confirmation method (5)	RL (mg/kg) (6)	Recovery (7)
Acephate							
Acrinathrin							
Aldicarb							
Azinphos-Methyl							
Azoxystrobin							
Bifenthrin							
Bromopropylate							
Captan + Folpet							
Carbendazim							
Carbofuran							
Chlorothalonil							
Chlorpyrifos							
Chlorpyrifos-methyl							
Cypermethrin							
Cyprodinil							
Deltamethrin							
Diazinon							
Dichlofluanid							
Dicofol							
Dimethoate							
Diphenylamine							
Endosulfan							
Esfenvalerate							
Fenvalerate							

ANNEX 1. Protocol and instructions. List of pesticides to be sought.

Pesticide	Quantification Method(1)	Analytical Procedure(2)	Conc. (mg/kg)(3)	Quantification Using Standards in solvents or matrix (4)	Confirmation method (5)	RL (mg/kg) (6)	Recovery (7)
Fenhexamid							
Imazalil							
Imidacloprid							
Iprodione							
Kresoxim-methyl							
Lambda-cyhalothrin							
Lindane							
Malathion							
Dithiocarbamates (expressed as CS ₂)							
Mecarbam							
Metalaxyll							
Methamidophos							
Methidathion							
Methiocarb							
Methomyl							
Mevinphos							
Myclobutanil							
Oxydemeton-methyl							
Parathion							
Penconazole							
Permethrin							
Phorate							
Phosalone							
Pirimicarb							
Pirimiphos-methyl							
Procymidone							
Propyzamide							
Spiroxamine							
Thiabendazole							
Tolyfluanid							
Triadimefon							
Triazophos							
Vinclozolin							

ANNEX 1. Protocol and instructions. List of pesticides to be sought.

- (1) If the substance is not included in your analysis, fill **NA**
If the substance is detected, fill **D**
If the substance is NOT detected, fill **ND**
- (2) Write the same code as you use in Form 3 for the analytical method used, e.g. A, B, C...
- (3) Concentration of **ONE** analysis **ONE** result (**NO DUPLICATES**)
Record the levels for all compounds according to the residue definition given in the table
- (4) Standards: **S** = standard/calibration in pure solvent, **M** = standard/calibration in matrix extract
- (5) Give the determination technique used e.g. **GC-FPD, HPLC-UV, GC-MS, LC-MS, LC-MS/MS**
- (6) **RL** Reporting Limit, must be given for all pesticides
For pesticides with metabolites/degradation products included in the MRL definition, give the "Reporting Limit" for the global compound (see definition in the pesticide list).
- (7) The results reported should not be corrected using recovery factors. If the laboratory usually corrects the results for the recoveries, they must express the correction factor used for each pesticide.

Please send this Form before the 7th July 2004

Signature:

FORM 3 (ANALYTICAL PROCEDURES USED)

Laboratory Code:

Date:

Fill in one of this forms for every different analytical procedure used

Analytical Procedure (2): _____

Weight sample (g): _____ Extraction solvent/s (7): _____

Partition step (if any): _____ Cleanup step (8): _____

Internal standard (if any): _____ Single level calibration: _____

Multi-level calibration: _____ Number of points: _____

Column type: _____ Stationary phase: _____ Determination (9): _____

Reference Method: _____

Signature:

Please send this Form before the 7th July 2004

(7) Denoted as **1** = ethyl acetate, **2** = acetone followed by cyclohexane and ethyl acetone, **3** = acetone followed by dichloromethane, **4** = acetone followed by dichloromethane and petroleum ether, **5** = acetonitrile, **6** = others.

(8) Clean-up: **GPC** = gel permeation chromatography, **SPE** = solid phase extraction, **LL** = liquid-liquid partition, **NO** = no clean-up, **O** = other clean-up method

(9) Determination Technique: e.g. **GC-ECD**, **GC-NPD**, **GC-FPD**, **GC-MS** (single-quad), **GC-ITD**, **HPLC-FL**, **HPLC-UV**, **HPLC-DAD**, **LC-MS**, **LC-MS/MS**

European Proficiency Test 06

Pesticide LOD's



<http://www.ual.es/GruposInv/EUPT6>

Pesticides, which could be present in the test material, with the "limit of analytical determination" (MPRL) given in the MRL residue definition.

Name	LOD's (mg/kg)
Tomato	
Acephate	0.02
Acrinathrin	0.05
Aldicarb (Aldicarb+ Aldicarb Sulfone+ Aldicarb Sulfoxide)	0.05
Azinphos-Methyl	0.05
Azoxystrobin	0.05
Bifenthrin	0.05
Bromopropylate	0.05
Captan + Folpet	0.1
Carbendazim (Benomyl + Carbendazim + Thiophanate-methyl)	0.1
Carbofuran (Carbofuran + 3-Hydroxy-Carbofuran)	0.1
Chlorothalonil	0.01
Chlorpyrifos	0.05
Chlorpyrifos-methyl	0.05
Cypermethrin	0.05
Cyprodinil	0.05

ANNEX 1. Protocol and instructions. List of pesticides to be sought.

Name	LOD's (mg/kg)
	Tomato
Deltamethrin	0.05
Diazinon	0.02
Dichlofluanid	0.05
Dicofol	0.02
Dimethoate (Dimethoate + Omethoate) Expressed as Dimethoate	0.02
Diphenylamine	0.05
Endosulfan ($\alpha+\beta$ Endosulfan + Endosulfan Sulfate) Expressed as Endosulfan	0.05
Esfenvalerate	0.02
Fenvalerate	0.02
Fenhexamid	0.05
Imazalil	0.02
Imidacloprid	0.05
Iprodione	0.02
Kresoxim-methyl	0.05
Lambda-cyhalothrin	0.02
Lindane	0.01
Malathion (Malathion + Malaoxon) Expressed as Malathion	0.05
Maneb Group (Maneb + Mancozeb + Metiram + Propineb + Zineb) Expressed as Dithiocarbamate (CS_2)	0.05
Mecarbam	0.05

ANNEX 1. Protocol and instructions. List of pesticides to be sought.

Name	LOD's (mg/kg)
Tomato	
Metalaxyll	0.05
Methamidophos	0.02
Methidathion	0.02
Methiocarb (Methiocarb + Methiocarb sulfone + Methiocarb sulfoxide) Expressed as Methiocarb	0.05
Methomyl (Methomyl + Thiodicarb) Expressed as Methomyl	0.05
Mevinphos	0.01
Myclobutanil	0.02
Oxydemeton-methyl (Oxydemeton-methyl + Demeton-S-Methylsulfon) Expressed as Oxydemeton-methyl	0.02
Parathion	0.05
Penconazole	0.05
Permethrin	0.05
Phorate (Phorate + Oxygenated Analogue + Sulfones + Sulfoxides) Expressed as Phorate	0.05
Phosalone	0.05
Pirimicarb	0.05
Pirimiphos-methyl	0.05
Procymidone	0.02
Propyzamide	0.02
Spiroxamine	0.05
Thiabendazole	0.05

ANNEX 1. Protocol and instructions. List of pesticides to be sought.

Name	LOD's (mg/kg)
	Tomato
Tolyfluanid	0.05
Triadimefon (Triadimefon + Triadimenol)	0.1
Triazophos	0.02
Vinclozolin	0.05

¹ The MPRLs refer to the "limit of analytical determination" given in the European Commission's Directives for the MRLs of the parent compounds. If no MPRL is listed in these Directives default value of 0.05 has been used.

ANNEX 2. List of laboratories invited to participate in PT6.

COUNTRY	City	Laboratory Name	Reported Results
AUSTRIA	Innsbruck	AGES Competence Centre for Pesticide Residues Innsbruck	yes
AUSTRIA	Vienna	Competence Centre Residue-Analysis- AGES	yes
AUSTRIA	Vienna	Austrian Agency for Health and Food Safety (ILUM-VIENNA)	yes
BELGIUM	Zwijnaarde	FYTOLAB	yes
BELGIUM	Bruxelles	IPH (Scientific Institute of Public Health)	yes
BELGIUM	Marloie	Laboratory of Hormonology	yes
CYPRUS	Nicosia	Pesticide Residue Laboratory. Department of Agriculture	yes
CYPRUS	Nicosia	State General Laboratory	yes
CZECH REPUBLIC	Praha 5	Czech Agriculture and Food Inspectorate	yes
CZECH REPUBLIC	Prague 10	National Institute of Public Health	yes
CZECH REPUBLIC	Prague 6	Institute of Chemical Technology	yes
DENMARK	Soeborg	Danish Institute for Food and Veterinary Research	yes
DENMARK	Copenhagen V	Fodevarerregion Kobenhavn	yes
ESTONIA	Saku (Harjumaa)	Agricultural Research Centre, Lab for Residues and Contaminants	yes
ESTONIA	Tartu	Health Protection Inspectorate	yes
FINLAND	Espoo	Finnish Customs Laboratory	yes
FINLAND	Helsinki	Enviroment Centre of Helsinki	yes
FRANCE	Talence	Laboratoire Interrég. de la Répression des Fraudes	yes
FRANCE	Illkirch-Graffenstaden	DGCCRF-Laboratoire de Strasbourg	yes
FRANCE	Montpellier Cedex 5	Laboratoire de la DGCCRF	yes
FRANCE	Villeneuve d'Ascq	D.G.C.C.R.F.- L59	yes

ANNEX 2. List of laboratories invited to participate in PT6.

COUNTRY	CITY	Laboratory Name	Reported Results
FRANCE	Rennes	Laboratoire DGCCRF	yes
FRANCE	Massy Cedex	DGCCRF-Laboratoire Interrégional de Paris-Massy	yes
GERMANY	Wuppertal	Chemisches Untersuchungsinstitut Bergisches Land	yes
GERMANY	Recklinghausen	Gemeinsames Chemisches und Lebensmitteluntersuchungsamt des Kreises Recklinghausen und der Stadt Gelsenkirchen; CEL	yes
GERMANY	Fellbach	CVUA Stuttgart	yes
GERMANY	Erlangen	Bayer. Landesamt für Gesundheit und Lebensmittelsicherheit	yes
GERMANY	Berlin	BBGes-ILAT, FB 26	yes
GERMANY	Hamburg	Institut Fur Hygiene und Umwelt	yes
GERMANY	Kassel	Staatliches Untersuchungsamt Hessen-Standort Kassel-	yes
GERMANY	Braunschweig	Lebensmittelinstitut Braunschweig	yes
GERMANY	Hagen	Chemisches Untersuchungsamt der Stadt Hagen	yes
GERMANY	Aachen	Chemisches und Lebensmitteluntersuchungsamt Stadt Aachen	yes
GERMANY	Bielefeld	Chemisches Untersuchungsamt Bielefeld	yes
GERMANY	Dortmund	Chem. und Lebensmitteluntersuchungsamt	yes
GERMANY	Rostock	Landesveterinär- und Lebensmitteluntersuch Amt M-V	yes
GERMANY	Speyer	Landesuntersuchungsamt, Institut für Lebensmittelchemie	yes
GERMANY	Trier	Landesuntersuchungsamt-Institut für Lebensmittelchemie	yes
GERMANY	Oldenburg	LAVES Lebensmittelinstitut Oldenburg	yes
GERMANY	Leipzig	Landesuntersuchungsanstalt Sachsen Standort Leipzig	yes
GERMANY	Kiel	Landeslabor Schleswig-Holstein Aubenstelle Kiel 1	yes

ANNEX 2. List of laboratories invited to participate in PT6.

COUNTRY	CITY	Laboratory Name	Reported Results
GERMANY	Essen	CGI Essen/Oberhausen	yes
GERMANY	Dusseldorf	Amt für Verbrauchers.-Abt.Chem.- und Lebensmittelunters.	yes
GERMANY	Saarbrücken	LVGA	yes
GERMANY	Sigmaringen	Chemisches und Veterinäruntersuchungsamt Sigmaringen	yes
GERMANY	Potsdam	Landeslabor Brandenburg, Fachbereich L5	yes
GERMANY	Bremen	Landesuntersuch.für Chemie	yes
GERMANY	Erfurt	TLLV	yes
GERMANY	Chemnitz	LUA Chemnitz FG Pestizide	yes
GERMANY	Wiesbaden	Staatliches Untersuchungsamt Hessen, Standort Wiesbaden	yes
GERMANY	Münster	Chemisches Landes- und Staatliches Veterinäruntersuchungsamt	yes
GERMANY	Dresden	LUA Sachsen, Dresden	yes
GERMANY	Bonn	Amt fur Umweltschutz und Verbraucherschutz und Lokale Agenda	yes
GERMANY	Halle	Landesamt fur Verbraucherschutz, FB 3	yes
GREECE	Athens	General Chemical State Laboratory	yes
GREECE	Kifissia, Athens	Pesticide Residues Laboratory, Benaki Phytopathological Institute	yes
GREECE	Patras	Regional Center of Plant Protection and Quality Control of Patras	yes
GREECE	Thessaloniki	Regional Center of Plant Protection and Quality Control	yes
GREECE	Iraklion, Crete	Regional Centre of Plant Protection	yes
GREECE	Ioannina	Regional Centre of Plant Protection and Quality Control	yes
GREECE	Lycovrissi, Athens	Pesticide Residue Laboratory of Lycovrissi	yes

ANNEX 2. List of laboratories invited to participate in PT6.

COUNTRY	CITY	Laboratory Name	Reported Results
GREECE	Kavala	Ministry Rural Development and Food, Peripheral Center	yes
HUNGARY	Velence	Plant Protection and Soil Conservation Service of Féjer County	yes
HUNGARY	Hódmezovásárhely	Plant Protection and Soil Conservation Service of Csongrad County	yes
HUNGARY	Szolnok	Plant Protection and Soil Conservation Service	yes
HUNGARY	Kaposvár	Plant Protection and Soil Conservation Service of Somogy County	yes
HUNGARY	Békéscsaba	BIO-CHEM LABOR BT.	no
HUNGARY	Miskolc	Pesticide Residue Laboratory PPSCS-BAZ	yes
HUNGARY	Fácánkert	Plant Protection and Soil Conservation Station of Tolna County	yes
HUNGARY	Tanakajd	Pesticide Residue Analytical Laboratory	yes
HUNGARY	Nyiregyháza	Plant Protection and Soil Conservation Service	yes
IRELAND	Dublin	Pesticide Control Service	yes
ITALY	La Loggia (Torino)	A.R.P.A Piemonte - Polo Chimico Regionale Alimenti	yes
ITALY	Pordenone	ARPA - Friuli Venezia Giulia Pordenone	yes
ITALY	Gorizia	ARPA-Dipartimento Provinciale di Gorizia	yes
ITALY	Palermo	ARPA Sicilia Dipartamento di Palermo	yes
ITALY	Ragusa	AUSL N7 DAP RAGUSA ARPA SICILIA L.I.P SEZ CHEMICA	yes
ITALY	Sondrio	ARPA-Dip. Sondrio-UnitaÓrganizzativa Laboratorio	yes
ITALY	Roma	Instituto Superiore di Sanità-Dip. Ambiente e Connessa Prevenzione Primaria-Reparto antiparassitari	yes
ITALY	Trento	A.P.P.A. Settore Lab. e Controlli	yes
ITALY	Bolzano	Agenzia Ambiente Bolzano	yes

ANNEX 2. List of laboratories invited to participate in PT6.

COUNTRY	CITY	Laboratory Name	Reported Results
ITALY	Vicenza	A.R.P.A. Veneto - Dip. di Vicenza	yes
ITALY	Florence	A.R.P.A.T. Dip. Provinciale di Firenze	yes
ITALY	Cagliari	P.M.P. Area Chimica Farmacologica	yes
ITALY	Bari	ARPA Puglia - Dipartimento di Bari	yes
ITALY	Ferrara	ARPA Sezione di Ferrara	yes
ITALY	Chiesuol del Fosso (Ferrara)	ARPA Sezione di Ferrara	yes
ITALY	Arezzo	ARPAT DIP Arezzo	yes
ITALY	Verona	APRAV-Verona	yes
ITALY	Nuoro	P.M.P. Area Chimica	no
ITALY	Catania	ARPA Sicilia-DAP Catania	yes
ITALY	La Spezia	A.R.P.A.L. Laboratorio	yes
LATVIA	Riga	State Agency "Public Health Agency"	yes
LATVIA	Riga	State Veterinary Medicine Diagnostic Centre (SVMDC)	yes
LUXEMBOURG	Luxembourg	Laboratoire du Contrôle des Denrées Alimentaires, LNS	yes
NORWAY	Ås	The Norwegian Crop Research Institute, Pesticide Laboratory	yes
POLAND	Trzebnica	Pesticide Residue Laboratory	yes
POLAND	Sosnicowice	Plant Protection Institute	yes
POLAND	Poznan	Plant Protection Institute, Department of Pesticide Residue Research	yes
POLAND	Bialystok	Plant Protection Institute, Experimental Field Station in Bialystok	yes
POLAND	Rzeszów	Plant Protection Institute	yes

ANNEX 2. List of laboratories invited to participate in PT6.

COUNTRY	CITY	Laboratory Name	Reported Results
PORTUGAL	Oeiras	Direcção-Geral de Protecção das Culturas	yes
PORTUGAL	Camacha	Laboratório Agrícola Da Madeira	yes
ROMANIA	Bucharest	Laboratorui Central pentru Controlui Reziduurilor de Pesticide din Plante si Produse Vegetale	yes
SLOVAKIA	Bratislava	State Veterinary and Food Institute	yes
SLOVAKIA	Bratislava 42	Central Agricultural Control & Testing Institute	yes
SLOVENIA	Ljubljana	Institute of Public Health	yes
SLOVENIA	Ljubljana	Agricultural Insitute of Slovenia, Central Laboratories	yes
SLOVENIA	Maribor	Public Health Institute, Environmental Protection Institute	yes
SPAIN	Villava Navarra	Gobierno de Navarra - Negociado de Análisis Instrumental	no
SPAIN	Cabrilis	Laboratori Agroalimentri Generalitat de Catalunya	yes
SPAIN	Murcia	Laboratorio Agrario y de Medio Ambiente	yes
SPAIN	A Coruña	Laboratorio Agrario y Fitopatológico de Galicia	yes
SPAIN	Logroño	Consejería de Agricultura. Laboratorio Regional de la Grajera	yes
SPAIN	Aravaca (Madrid)	Laboratorio Arbitral Agroalimentario	yes
SPAIN	Burjassot (Valencia)	Agroalimentario Generalitat Valenciana	yes
SPAIN	Majadahonda (Madrid)	Centro Nacional de Alimentacion	yes
SPAIN	Zizurkil-Gipúzcoa	Laboratorio Agrario Diputación Foral de Gipuzkoa	yes
SPAIN	Sevilla	Laboratorio del Catice (SOIVRE) de Sevilla	yes
SPAIN	La Mojonera, Almeria	Laboratorio de Sanidad Vegetal de Almería	yes
SPAIN	Almeria	Laboratorio del Catice (SOIVRE) de Almería	yes

ANNEX 2. List of laboratories invited to participate in PT6.

COUNTRY	City	Laboratory Name	Reported Results
SPAIN	Murcia	S.O.I.V.R.E. Murcia	yes
SPAIN	Burgos	Laboratorio Agrario Regional. Dir. Gen. De Producción Agropecuaria	yes
SPAIN	Jaén	Laboratorio de Sanidad Vegetal. CAPJA	yes
SWEDEN	LIDKÖPING	AnalyCen Nordic AB	yes
THE NETHERLANDS	Amsterdam	VWA-KVV (Food and Consumer Product Safety Authority)	yes
UNITED KINGDOM	York	Central Science Laboratory (CSL)	yes
UNITED KINGDOM	Teddington, Middlesex	Laboratory of the Government Chemist Limited	yes
UNITED KINGDOM	Wolverhampton	Direct Laboratories	yes
UNITED KINGDOM	Edinburgh, Scotland	Scottish Agricultural Science Agency	yes