

Analysis of Fumigants in Cereals and Dry Fruits Applying GC-MS/MS

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Brief description:

The intension was to develop a multi-method to analyze residues fumigants in cereals and dried fruits together with carbon disulfide (hydrolysis product of dithiocarbamates) by GC-MS/MS. The fumigants were extracted with n-hexane. The extracts were filtered and subjected to gas chromatography quadrupole mass spectrometry (GC-MS/MS). The isotopic labeled substance chlorobenzene D5 was used as internal standard.

Compounds included: (carbon tetrachloride, trichloroethylene, ethylene chlorobromide, 1,3-dichloropropene (cis+trans), tetrachloroethylene, 1,1,2,2-tetrachloroethane, p-dichlorobenzene, naphthalene, 1,2-dibromo-3-chloropropane, dichlorvos, p-nitrochlorobenzene, azobenzene, chloropicrin and ethylene dibromide).

Table 1 lists the MRLs for the compounds.

Table 1 MRLs of fumigant residues

| Substance | Maximum residue level (mg kg ⁻¹)* |
|-----------------------------|---|
| 1,2-Dibromo-3-Chloropropane | 0.01** |
| 1,3-Dichloropropene | 0.01 (products of animal origin) |
| Azobenzene | 0.01** |
| Carbon Tetrachloride | 0.01** |
| Chloropikrin | 0.02 (tea. spices), 0.01 (cereals) |
| Dichlorvos | 0.01 |
| Ethylene Chlorobromide | 0.01** |
| Ethylene Dibromide | 0.01 |
| Naphthalene | 0.01** |
| p-Dichlorobenzene | 0.01** |
| p-Nitrochlorobenzene | 0.01** |
| Tetrachlorethane | 0.01** |
| Tetrachloroethylene | 0.01** |
| Trichlorethylene | 0.01** |

* (European Commission Directorate - General Health & Consumers (SANCO) I)

** Default MRL according to article 18 (1B) REGULATION (EC) NO 396/2005 (14)

Materials:

n-hexane of EMPLURA[®] grade (Merck Darmstadt, Germany)

helium 5.0 and argon 5.0 (Praxair) for GC-MS/MS

carbon disulfide (pro analysi grade) (Merck Darmstadt, Germany),

carbon tetrachloride (purity ≥ 99.5 %), trichloroethylene (purity ≥ 99.6 %), ethylene chlorobromide (purity ≥ 99.5 %), 1,3-dichloropropene (cis+trans), (purity ≥ 92 %), tetrachloroethylene (purity ≥ 99 %), 1,1,2,2-tetrachloroethane (purity ≥ 98.5 %),

p-dichlorobenzene (purity ≥ 99.5 %), naphthalene (purity ≥ 99.5 %), 1,2-dibromo-3-dichloropropane (purity ≥ 98.5 %), dichlorvos (purity ≥ 98 %), p-nitrochlorobenzene (purity ≥ 99.5 %) and azobenzene (purity ≥ 98.5 %) (Dr. Ehrenstorfer GmbH Augsburg, Germany), chloropicrin (purity ≥ 99 %) and ethylene dibromide (purity ≥ 99.6 %) (Sigma-Aldrich Munich, Germany)

internal standards chlorobenzene D5 (purity ≥ 99 %) and 1,1,2-tribromoethane (purity ≥ 99 %) (Sigma-Aldrich Munich, Germany).

The stock solutions of all standards (1 mg mL⁻¹ in n-hexane) were diluted to 10 µg mL⁻¹ and 1 µg mL⁻¹ (working solutions) with n-hexane. All solutions were stored in the dark at 4 °C.

Instrumentation:

Geno Grinder 2010 (SPEX Sample Prep, Metuchen, USA)

centrifuge Rotanta 460 (Hettich Tuttlingen, Germany)

electronic pipettes applicable for volumes of 10–100 µL and 100–1000 µL, respectively, and manual pipettes applicable for volumes of 1–10 mL (Eppendorf Hamburg, Germany)

volumetric pipettes (10 mL; 0,1 DIN B Ex 20 °C; Hirschmann Laborgeräte, Eberstadt, Germany).

50 mL PP (114×28 mm) single-use tubes with screw caps (Sarstedt Nümbrecht, Germany).

1.5 mL GC autosampler vials (Klaus Ziemer GmbH Langerwehe, Germany)
 6 mL single-use syringes from (Henke Sass Wolf Tuttlingen, Germany)
 disposable polyester syringe filters (0.45 µm pore size, 15 mm diameter) from (Machery-Nagel Düren, Germany).

ThermoScientific Trace 1310 GC system (ThermoScientific, Waltham, USA) combined with the mass spectrometer ThermoScientific TSQ 8000 (ThermoScientific, Waltham, USA), run in EI positive mode was used for the analysis of the fumigant extracts. The GC system was connected to a TriPlus RSH autosampler (ThermoScientific, Waltham, USA).

Sample Extraction:

Dry samples (e.g. wheat grains) were directly weighed without any milling and without adding water.

For the extraction 5 g ± 0.1 g of the samples were weighed into 50 mL PP tubes. 5 mL of n-hexane followed by 50 µL of the internal standard working solution (1 µg mL⁻¹) were added to the samples. Two isotopic labeled standards (chlorobenzene D5 and 1,1,2-tribromoethane) were used. The tube was closed and shaken by a mechanical shaker for one minute. The tube was centrifuged for 5 min at 4000 rpm afterwards filtered through a syringe filter (0.45 µm) into 50 mL PP tubes. 1 mL of the extract was transferred into vials. The extracts were applied GC-MS/MS measurement.

GC-MS/MS analysis

Injection: split-mode, split ratio of 1:5 (split flow: 5 mL/min) injection volume 2 µL.

| | Rate (°C/ s) | Temperature (°C) | Time (min) |
|-----------|--------------|------------------|------------|
| Injection | | 120 | 0,1 |
| Transfer | 14 | 250 | 5 |
| Cleaning | 10 | 300 | 10 |

Flow: helium with a constant flow rate of 1 mL/min

Column: 30 m; 0,20 mm; 1,12 μm Agilent HP VOC column (Agilent, Waldbronn, Germany)

Pre-column 10 m; 0,25 mm Fused-Silica (Agilent, Waldbronn, Germany).

Oven program:

| Ramp | Rate ($^{\circ}\text{C}/\text{min}$) | Temperature ($^{\circ}\text{C}$) | Hold Time (min) |
|---------|--|------------------------------------|-----------------|
| Initial | | 45 | 2 |
| 1 | 5 | 80 | 5 |
| 2 | 125 | 280 | 10 |

MS/MS parameters: EI ionization: positive mode, MS/MS detection selected reaction monitoring (SRM) mode. transferline 350 $^{\circ}\text{C}$, source temperature 280 $^{\circ}\text{C}$

The mass transitions for each compound are shown in the Appendix.

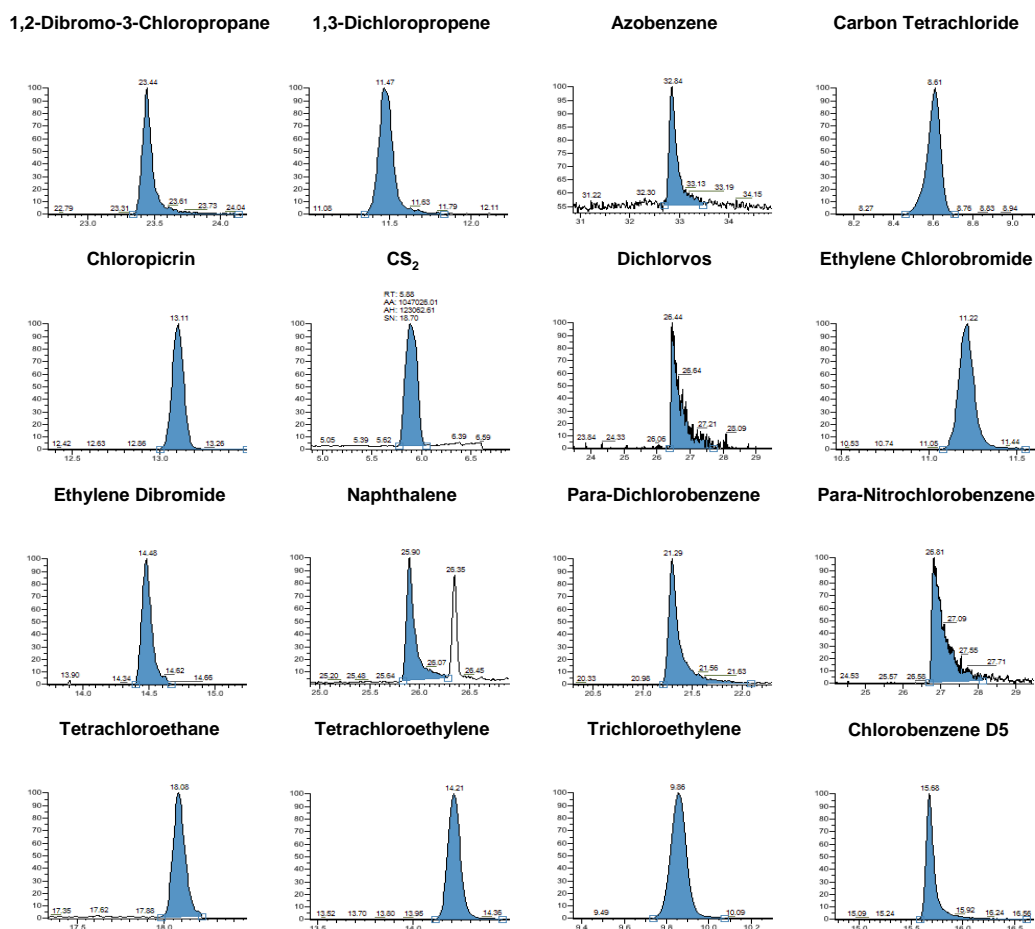
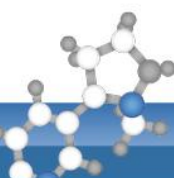


Figure 1 Chromatograms of the fumigants on wheat matrix, 0.1 mg kg⁻¹



Validation:

The validation in wheat and raisin matrix (n=5) showed satisfying recoveries between 70–120 % and relative standard deviations (RSDs) well below 10 % for all compounds (see tables 2 and 3). except CS₂. The substance is very volatile and therefore discriminated. An internal standard with similar losses such as CS₂ would be beneficial.

Table 2 Recoveries for Raisin

| Compound | Raisin | | | | | | |
|-----------------------------|--------------|-----|-----|-----|-----|-------------|-----------|
| | Recovery (%) | | | | | Average (%) | RSD (%) |
| 1,2-Dibromo-3-Chloropropane | 103 | 98 | 102 | 102 | 101 | 101 | 2 |
| 1,3-Dichloropropene | 95 | 93 | 92 | 92 | 93 | 93 | 1 |
| Azobenzene | 106 | 102 | 114 | 127 | 144 | 118 | 13 |
| Carbon Tetrachloride | 86 | 85 | 85 | 85 | 88 | 86 | 1 |
| Chloropicrin | 96 | 95 | 94 | 95 | 97 | 95 | 1 |
| CS ₂ | 52 | 54 | 51 | 53 | 53 | 52 | 2 |
| Ethylene Chlorobromide | 92 | 91 | 95 | 92 | 93 | 93 | 2 |
| Ethylene-Dibromide | 96 | 97 | 99 | 98 | 99 | 98 | 1 |
| Naphthalene | 102 | 97 | 101 | 99 | 95 | 99 | 3 |
| p-Nitrochlorobenzene | 108 | 104 | 102 | 98 | 98 | 102 | 4 |
| p-Dichlorobenzene | 107 | 99 | 101 | 106 | 107 | 104 | 3 |
| Tetrachloroethane | 103 | 92 | 98 | 95 | 101 | 98 | 4 |
| Tetrachloroethylene | 100 | 98 | 98 | 100 | 100 | 99 | 1 |
| Trichloroethylene | 91 | 90 | 88 | 90 | 91 | 90 | 1 |

Table 3 Recoveries for Wheat

| Compound | Wheat | | | | | | |
|-----------------------------|--------------|----|----|----|----|-------------|----------|
| | Recovery (%) | | | | | Average (%) | RSD (%) |
| 1,2-Dibromo-3-Chloropropane | 97 | 96 | 98 | 95 | 98 | 97 | 1 |
| 1,3-Dichloropropene | 79 | 83 | 83 | 90 | 82 | 83 | 4 |
| Azobenzene | 97 | 95 | 97 | 89 | 92 | 94 | 3 |
| Carbon Tetrachloride | 77 | 81 | 79 | 85 | 75 | 79 | 4 |
| Chloropicrin | 87 | 87 | 89 | 91 | 85 | 88 | 3 |
| CS ₂ | 52 | 58 | 53 | 61 | 51 | 55 | 7 |
| Ethylene Chlorobromide | 82 | 78 | 80 | 85 | 80 | 81 | 3 |
| Ethylene-Dibromide | 85 | 87 | 89 | 87 | 89 | 87 | 2 |

| | | | | | | | |
|----------------------|----|-----|-----|-----|----|-----------|----------|
| Naphthalene | 94 | 103 | 98 | 101 | 99 | 99 | 3 |
| p-Nitrochlorobenzene | 90 | 98 | 92 | 91 | 97 | 94 | 4 |
| p-Dichlorobenzene | 97 | 99 | 101 | 90 | 95 | 96 | 4 |
| Tetrachloroethane | 91 | 91 | 93 | 94 | 94 | 93 | 1 |
| Tetrachloroethylene | 90 | 91 | 91 | 93 | 91 | 91 | 1 |
| Trichloroethylene | 79 | 84 | 82 | 87 | 79 | 82 | 4 |

The limit of quantification (LOQ) and the limit of detection (LOD) were estimated from the undiluted extracts as the level showing a signal-to-noise ratio of at least 10 to 1 (LOQ) respectively 3 to 1 (LOD). **Table** shows the LOQ and LOD of the compounds for solvent and matrix.

Table 4 LOQs and LODs of the fumigants in solvent and matrix

| Fumigant | Matrix | LOQ (mg kg ⁻¹) | LOD (mg kg ⁻¹) |
|-----------------------------|--------------|----------------------------|----------------------------|
| 1,2-Dibromo-3-Chloropropane | n-Hexane | 0.01 | 0.005 |
| | Raisin/Wheat | 0.01 | 0.005 |
| 1,3-Dichloropropene | n-Hexane | 0.05 | 0.01 |
| | Raisin/Wheat | 0.05 | 0.01 |
| Azobenzene | n-Hexane | 0.01 | 0.01 |
| | Raisin/Wheat | 0.01 | 0.01 |
| Carbon Tetrachloride | n-Hexane | 0.001 | 0.001 |
| | Raisin/Wheat | 0.005 | 0.005 |
| Chloropicrin | n-Hexane | 0.005 | 0.005 |
| | Raisin/Wheat | 0.01 | 0.005 |
| | Wheat | 0.01 | 0.005 |
| CS ₂ | n-Hexane | 0.05 | 0.05 |
| | Raisin/Wheat | 0.05 | 0.05 |
| Dichlorvos | n-Hexane | 0.1 | 0.05 |
| | Raisin/Wheat | 0.1 | 0.1 |
| Ethylene Chlorobromide | n-Hexane | 0.05 | 0.05 |
| | Raisin/Wheat | 0.05 | 0.05 |
| Ethylene Dibromide | n-Hexane | 0.05 | 0.01 |
| | Raisin/Wheat | 0.05 | 0.05 |
| Naphthalene | n-Hexane | 0.05 | 0.01 |
| | Raisin/Wheat | 0.01 | 0.01 |
| Para-Nitrochlorobenzene | n-Hexane | 0.05 | 0.05 |
| | Raisin/Wheat | 0.05 | 0.05 |
| Para-Dichlorobenzene | n-Hexane | 0.01 | 0.005 |

| Fumigant | Matrix | LOQ (mg kg ⁻¹) | LOD (mg kg ⁻¹) |
|---------------------|--------------|----------------------------|----------------------------|
| | Raisin/Wheat | 0.01 | 0.005 |
| Tetrachloroethane | n-Hexane | 0.01 | 0.005 |
| | Raisin | 0.01 | 0.005 |
| | Wheat | 0.1 | 0.005 |
| Tetrachloroethylene | n-Hexane | 0.001 | 0.0005 |
| | Raisin/Wheat | 0.005 | 0.0005 |
| Trichloroethylene | n-Hexane | 0.001 | 0.005 |
| | Raisin/Wheat | 0.005 | 0.005 |

Discussion and Conclusions:

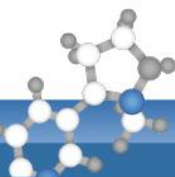
Since the analytes are volatile it is recommended to prepare the calibration solutions daily, and to analyze the extracts directly after extraction.

Linear range of the majority of the fumigants is between 0.005 to 2 ppm. The slopes of the calibration curves on matrix tend to be lower compared to the calibration curves on solvent (see **Fehler! Verweisquelle konnte nicht gefunden werden.**). An exception is trichloroethylene, where the slope of the calibration curve on matrix is in a similar range as the solvent.

Not for all fumigants LOQs were low enough to control the MRLs. An up scaling of the extraction conditions might be advantageous in this case.

Appendix:
Table 2 Mass transitions of the compounds

| Compounds | Retention Time (min) | Precursor Mass (u) | Product Mass (u) | Collision Energy (eV) |
|-----------------------------|----------------------|--------------------|------------------|-----------------------|
| CS ₂ | 5.6 | 77.7 | 77.9 | 10 |
| | | 75.9 | 75.9 | 10 |
| | | 75.9 | 44 | 40 |
| | | 77.7 | 45.8 | 40 |
| Pentafluorobenzene | 8.0 | 99 | 79.6 | 25 |
| | | 168 | 99 | 20 |
| Carbon Tetrachloride | 8.2 | 116.9 | 81.9 | 28 |
| | | 118.9 | 83.9 | 28 |
| | | 120.9 | 83.9 | 28 |
| Trichlorethylene | 9.3 | 94.9 | 60 | 25 |
| | | 129.9 | 95 | 10 |
| | | 131.8 | 96.9 | 10 |
| Ethylene Chlorobromide | 10.6 | 65 | 65 | 0 |
| | | 143.9 | 63 | 5 |
| | | 63 | 63 | 0 |
| Methylisothiocyanate (MITC) | 11.9 | 71.8 | 45 | 15 |
| | | 73 | 45 | 10 |
| 1.3-Dichloropropene | 12 | 110 | 75 | 5 |
| | | 111.9 | 77.2 | 5 |
| Chloropicrin | 12.5 | 116.9 | 82 | 30 |
| | | 120.9 | 83.9 | 30 |
| | | 118.9 | 81.9 | 30 |
| Tetrachloroethylene | 13.5 | 128.9 | 93.9 | 20 |
| | | 130.9 | 95.9 | 15 |
| | | 163.9 | 128.9 | 15 |
| | | 165.8 | 130.9 | 10 |
| Ethylene-Dibromide | 13.8 | 187.8 | 107 | 5 |
| | | 106.9 | 106.9 | 0 |
| | | 108.9 | 108.9 | 0 |



| Compounds | Retention Time (min) | Precursor Mass (u) | Product Mass (u) | Collision Energy (eV) |
|-----------------------------|----------------------|--------------------|------------------|-----------------------|
| Chlorobenzene D5 (ISTD) | 15.8 | 82 | 54 | 15 |
| | | 117 | 82.1 | 15 |
| | | 118.8 | 82.1 | 15 |
| Tetrachloroethane | 17.2 | 132.6 | 97 | 15 |
| | | 165.8 | 83 | 5 |
| | | 82.8 | 82.8 | 0 |
| P-Dichlorobenzene | 20.0 | 111 | 75.1 | 10 |
| | | 112.8 | 75 | 15 |
| | | 146 | 111 | 15 |
| | | 147.9 | 113 | 15 |
| 1.2-Dibromo-3-Chloropropane | 22.0 | 154.9 | 75 | 5 |
| | | 154.9 | 92.9 | 25 |
| | | 156.8 | 75 | 5 |
| | | 234 | 155 | 5 |
| 1.1.2-Tribromoethane (ISTD) | 22.0 | 184.9 | 104.9 | 15 |
| | | 186.9 | 104.9 | 15 |
| | | 186.9 | 106.9 | 15 |
| | | 188.8 | 106.9 | 15 |
| Naphthalene | 24.4 | 127.9 | 77.7 | 20 |
| | | 128 | 128 | 0 |
| Dichlorvos | 25.2 | 109 | 79 | 5 |
| | | 185 | 93.1 | 10 |
| | | 185 | 108.9 | 15 |
| Para-Nitrochlorobenzene | 25.5 | 111.1 | 75.1 | 10 |
| | | 156.9 | 99 | 15 |
| 1-Bromo-4-Nitrobenzene | 28.4 | 154.9 | 75.5 | 15 |
| | | 156.8 | 75.7 | 15 |
| | | 170.9 | 143 | 10 |
| Azobenzene | 29.7 | 105 | 77.1 | 5 |
| | | 182.1 | 105.1 | 5 |